DuoBody-EpCAMx4-1BB mediates conditional T-cell co-stimulation and promotes antitumor activity in preclinical models

<u>Sina Fellermeier-Kopf¹</u>*, Andreea Ioan-Facsinay²*, Andrea Imle¹, Annieck M. Diks², Kristina B. Nuermberger¹, Lars Guelen², Maren Köhne¹, Mischa Houtkamp², Aras Toker¹, Saskia M. Burm², Joshua Gamse³, Christine Janaitis¹, David Satijn², Alexander Muik¹, Tahi Ahmadi³, Özlem Türeci¹, Kristel Kemper², Uğur Şahin¹*, Esther Breij²*

¹BioNTech SE, Mainz, Germany; ²Genmab B.V., Utrecht, the Netherlands; ³Genmab US Inc., Princeton, NJ, USA. *These authors contributed equally.

Correspondence to: sina.fellermeier-kopf@biontech.de; aif@genmab.com

Conclusions

DuoBody[®]-EpCAMx4-1BB (BNT314/GEN1059) is a novel Fc-inert bispecific antibody designed to boost antitumor immune responses through EpCAMdependent 4-1BB agonistic activity.

In preclinical studies we found that:

- DuoBody-EpCAMx4-1BB induced 4-1BB signaling, thereby enhancing activation, proliferation, cytokine secretion, and cytotoxic activity of human T cells in vitro and ex vivo.
- 4-1BB agonistic activity of DuoBody-EpCAMx4-1BB was strictly conditional, *i.e.*, dependent on EpCAM binding.
- An Fc-inert hEpCAMxm4-1BB bispecific antibody exhibited antitumor activity in vivo, providing proof-ofprinciple for dual targeting of EpCAM and 4-1BB.
- DuoBody-EpCAMx4-1BB was well-tolerated in cynomolgus monkeys.

Based on these results, DuoBody-EpCAMx4-1BB is hypothesized to boost antitumor immunity in patients with EpCAM⁺ tumors. The clinical safety and preliminary antitumor activity of DuoBody-EpCAMx4-1BB will be investigated in patients with solid tumors in a first-inhuman trial

References 2022: 12(5):1248-1265

1. Chester et al., Immunotherapy targeting 4-1BB: mechanistic rationale, clinical results, and future strategies. Blood. 2018; 131(1):49-57 2. Muik et al., Preclinical Characterization and Phase I Trial Results of a Bispecific Antibody Targeting PD-L1 and 4-1BB (GEN1046) in Patients with Advanced Refractory Solid Tumors. Cancer Discov.

Acknowledgments

This study was sponsored by BioNTech SE and Genmab A/S. The authors would like to thank nantharaman Muthuswamy, Ann-Kathrin Wallisch, Bart-Jan de Kreuk, Bernadette Jesionek, Dennis Verzijl, Diana Schneider, Julia Mühl, Laura Smits de Vries, Merve Eken, Michiel Bolkenstein, Murielle van der Horst, Natalie Schwarz, Patrick Franken, and Patrick Goldberg for

their technical expertise and support. Schematic in Fig 6A was generated using biorender.com.

3. Balzar et al., The biology of the 17-1A antigen (Ep-CAM). J Mol Med (Berl). 1999; 77(10):699-712.

4. Huang et al., Functions of EpCAM in physiological processes and diseases (Review). Int J Mol Med. 2018; 42(4):1771-1785.

Disclosures Fellermeier-Kopf, Imle, Janaitis, Köhne, Muik, Nuermberger, Toker: Employee and stockholder of BioNTech. Sahin, Türeci: Management board member, employee and stockholder of BioNTech. Ahmadi, Breij, Burm, Diks, Gamse, Guelen, Houtkamp, Ioan-Facsinay, Kemper, Satijn: Employee and stockholder of Genmab.

clinical trials¹.

• DuoBody-EpCAMx4-1BB (BNT314/GEN1059) is an Fc-inert bsAb that is generated by controlled Fab-arm exchange (DuoBody platform) of an EpCAM-specific human IgG1 antibody and a 4-1BB-specific humanized IgG1 antibody (Fig 1). • DuoBody-EpCAMx4-1BB binds to immobilized recombinant human EpCAM and human 4-1BB with affinities (K_D) of 56 nM and 0.12 nM, respectively.

200

Figure 3: Immunohistochemistry analysis of EpCAM expression in tumor tissue microarrays. H-score was calculated based on manual scoring of EpCAM staining intensity by a certified pathologist (0 = no, 1 = low, 2 = intermediate, 3 = high staining)using the formula: $1 \times \%$ low pos. $+ 2 \times \%$ intermediate pos. $+ 3 \times \%$ high pos. cells. Symbols represent tissue cores of individual patients. AC, adenocarcinoma; CA, carcinoma; RCC, renal clear cell carcinoma; SCC, squamous cell carcinoma; PDAC, pancreatic ductal adenocarcinoma.

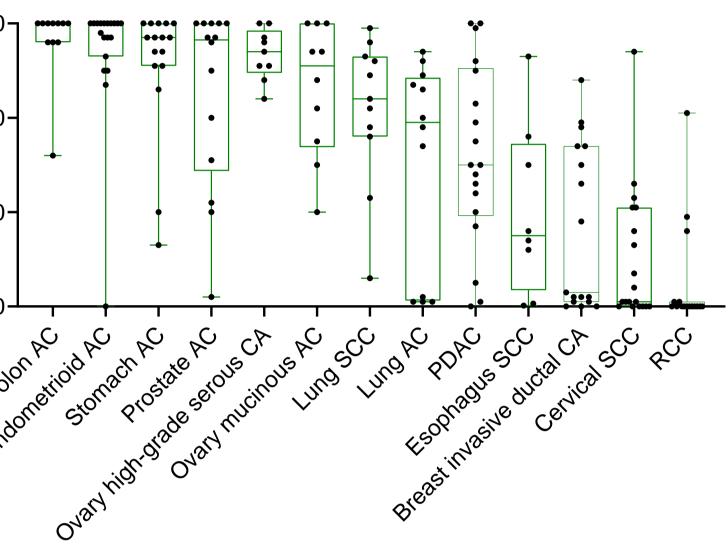
• 4-1BB is recognized as a promising target to enhance antitumor immunity, although classical 4-1BB agonistic antibodies have been associated with the occurrence of dose-limiting liver toxicity or have shown lack of efficacy as monotherapy in

• Bispecific antibodies (bsAbs) redirecting 4-1BB agonistic activity to the tumor microenvironment were shown to potently enhance antitumor immune responses in preclinical mouse models^{1,2}.

• EpCAM is highly expressed on the surface of tumor cells and accessible for antibody binding in many solid tumor types of epithelial origin, whereas in healthy epithelia EpCAM is largely shielded by tight junctions^{3,4}.

EpCAM expression in human solid tumors

• Immunohistochemistry analyses revealed solid tumor indications with high EpCAM expression (as shown by an H-score of >200) and indications with variable EpCAM expression (Fig 3).



Conditional 4-1BB agonistic activity in vitro

• DuoBody-EpCAMx4-1BB exhibited conditional (i.e., EpCAM-dependent) 4-1BB agonistic activity *in vitro* (**Fig 4A**).

• 4-1BB agonistic activity of DuoBody-EpCAMx4-1BB was observed in the presence of EpCAM⁺ tumor cell lines showing a range of EpCAM expression levels (Fig 4B).

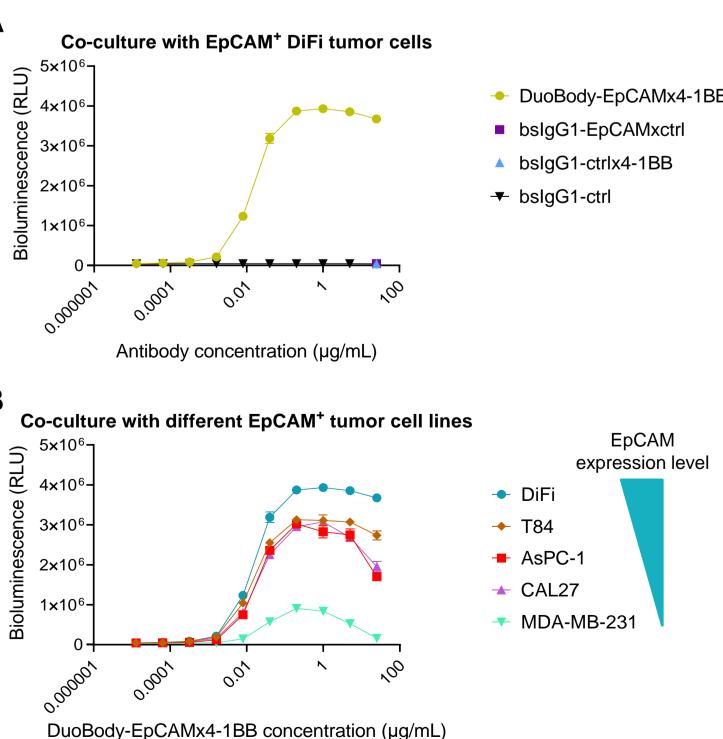
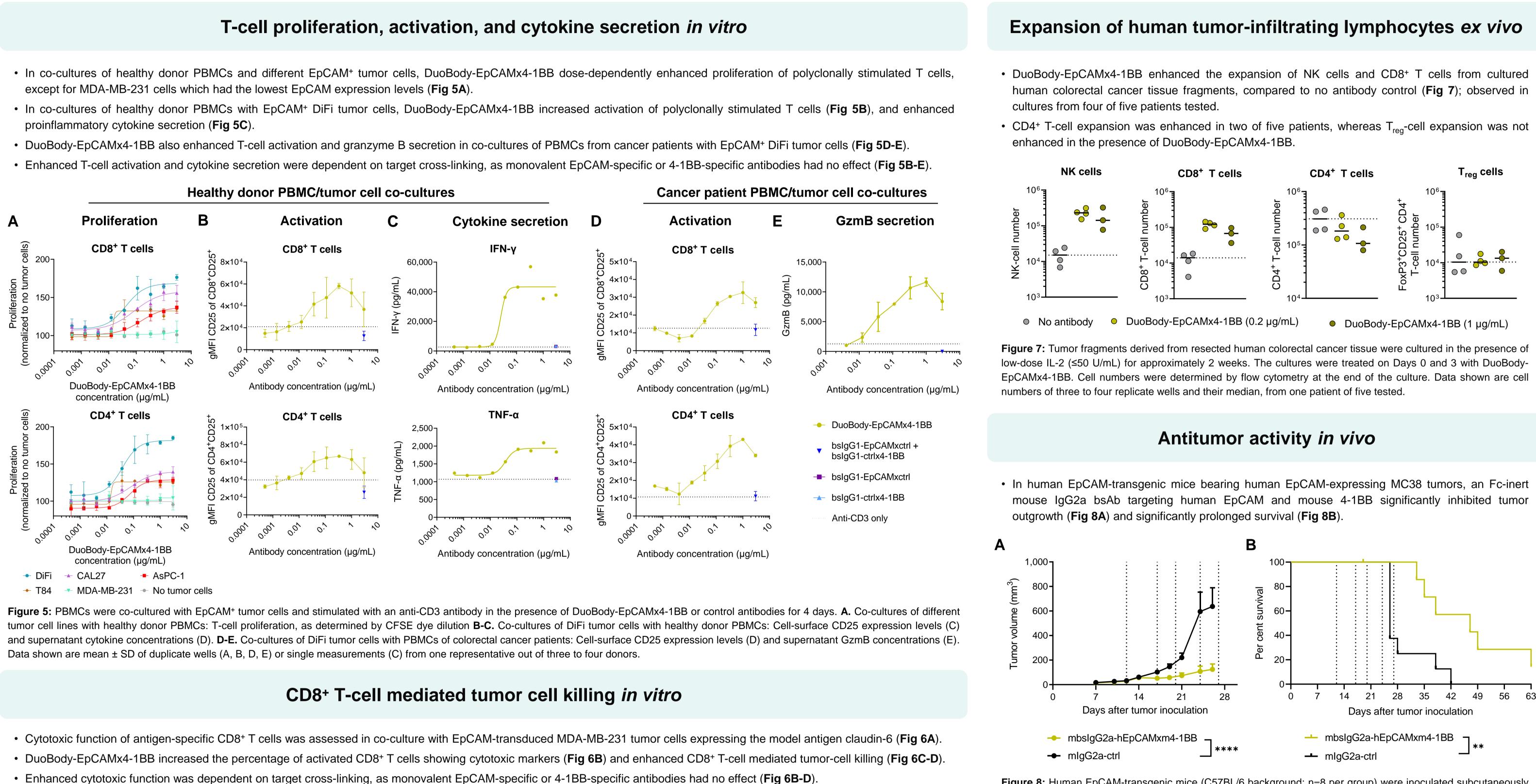
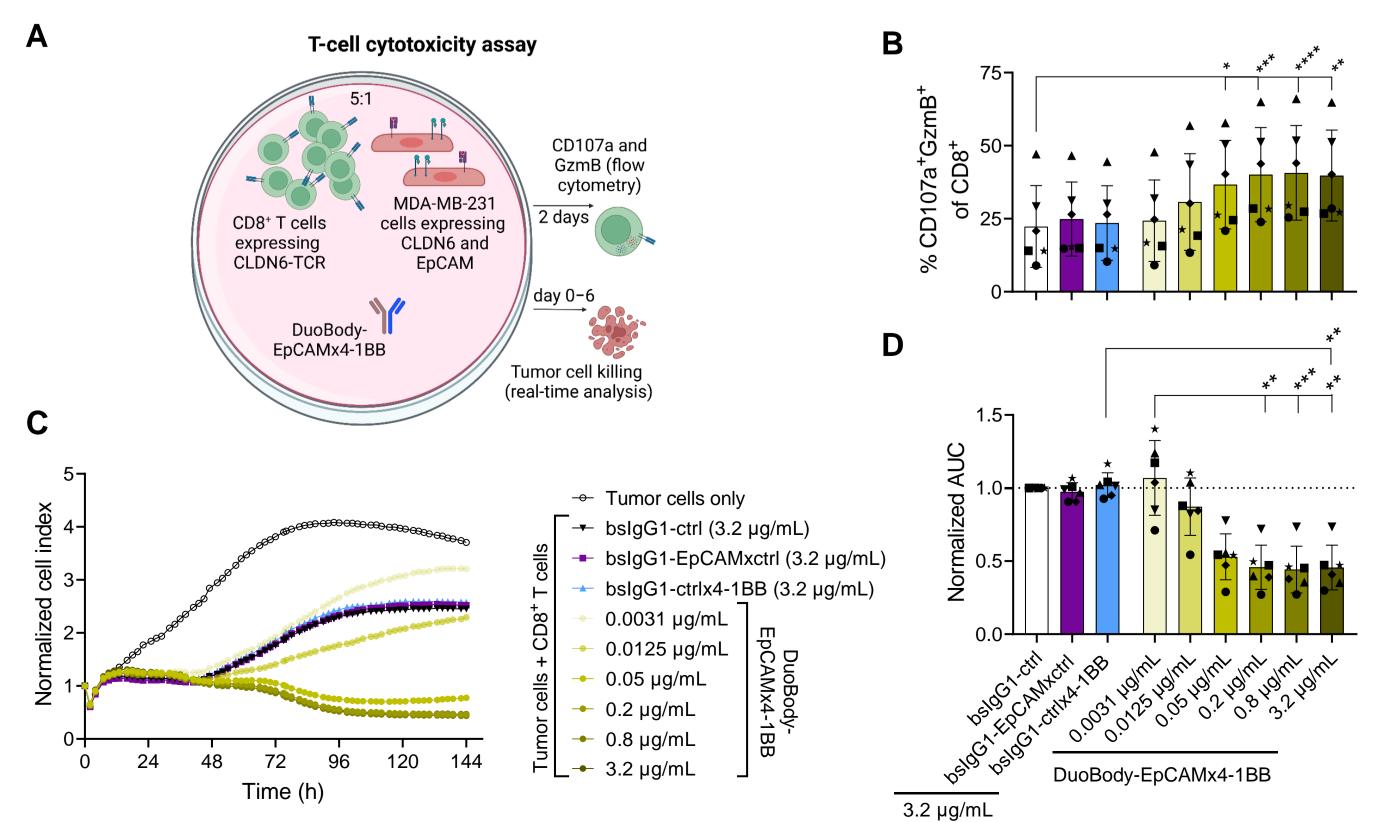


Figure 4: In reporter Jurkat T cells, activation of 4-1BB results in luciferase expression, which is measured by conversion of a bioluminescent substrate. A. 4-1BB agonistic activity of DuoBody-EpCAMx4-1BB in co-cultures of reporter Jurkat T cells and EpCAM+ DiFi tumor cells. **B.** 4-1BB agonistic activity of DuoBody-EpCAMx4-1BB in co-cultures of reporter Jurkat T cells with a panel of five EpCAM⁺ tumor cell lines. Data shown are mean bioluminescence ± SD from duplicate measurements. RLU, relative light units.

- proinflammatory cytokine secretion (Fig 5C).





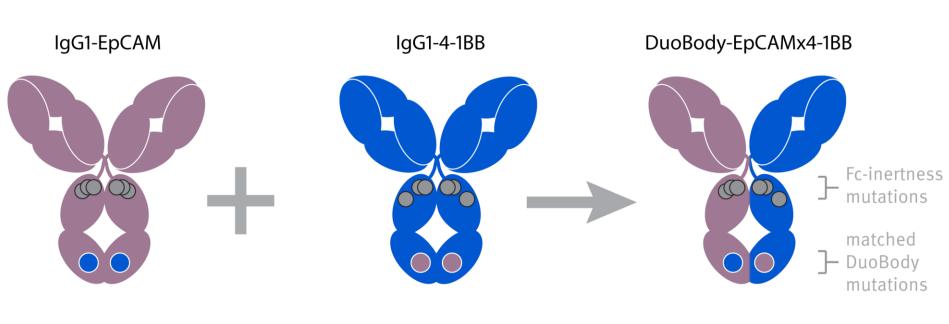


Figure 1: Generation of DuoBody-EpCAMx4-1BB by controlled Fab-arm exchange

Results

Figure 6: A. Healthy donor PBMC-derived CD8+ T cells engineered to express a CLDN6-specific TCR were co-cultured with CLDN6⁺ EpCAMtransduced MDA-MB-231 tumor cells in the presence of DuoBody-EpCAMx4-1BB or control antibodies. **B.** Percentages of CD107a⁺GzmB⁺ CD8⁺T cells were determined by flow cytometry after 2 days. Data represent mean ± SD of six donors. C. Cell index values derived from impedance measurements during the co-culture were determined as a measure of tumor-cell killing. Data normalized to the time point of coculture initiation are shown from one representative donor. **D.** AUC analysis of normalized cell-index curves of all six donors tested. The AUC of each treatment was normalized to bslgG1-ctrl-treated cultures from the same donor. ****, P<0.0001; ***, P<0.001; **, P<0.01; *, P<0.05; Friedman test with Dunn's multiple comparisons test. CLDN6, claudin-6

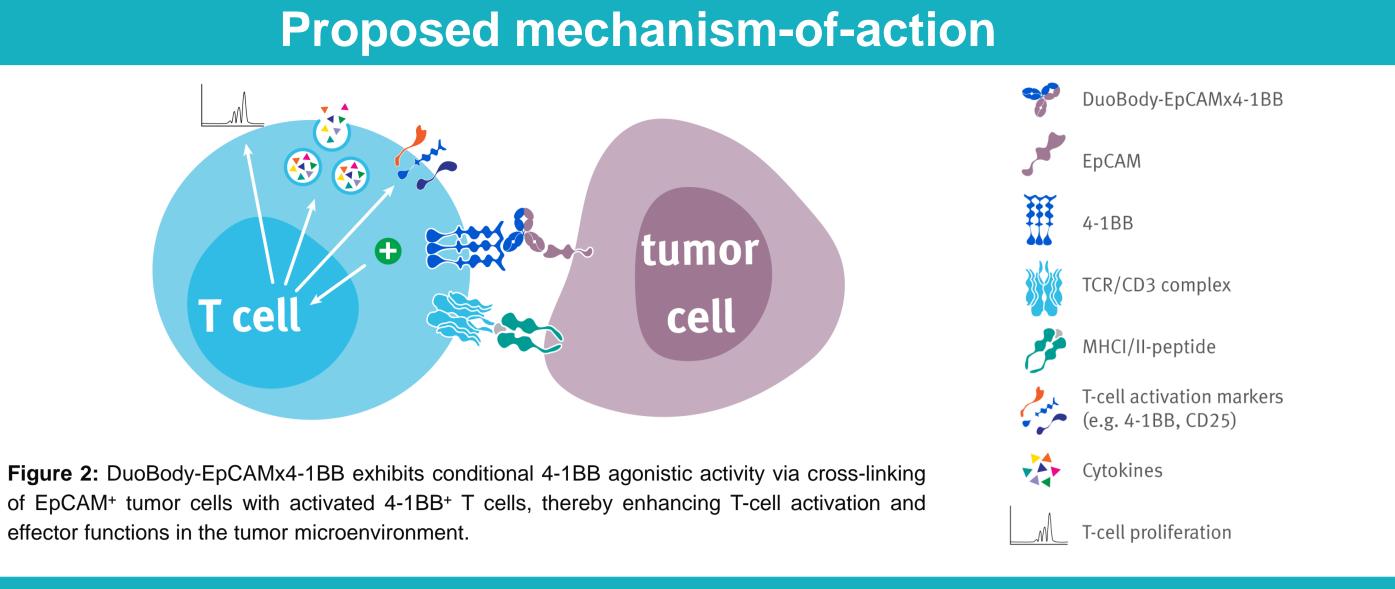


Figure 8: Human EpCAM-transgenic mice (C57BL/6 background; n=8 per group) were inoculated subcutaneously with 5 × 10⁵ human EpCAM-expressing MC38 tumor cells. Starting on Day 12 after inoculation (mean tumor volume of 31 mm³), the mice were treated intraperitoneally with 100 µg mbslgG2a-hEpCAMxm4-1BB or control antibody twice weekly for 2.5 weeks. A. Tumor volumes over time (group mean ± SEM). ****, P<0.0001; Two-Way ANOVA. B. Kaplan-Meier analysis of survival. **, P<0.01; Log-rank (Mantel-Cox) test. Dotted lines indicate treatment days.

Non-clinical safety in cynomolgus monkeys

- DuoBody-EpCAMx4-1BB was well-tolerated in cynomolgus monkeys at doses up to 50 mg/kg (the highest tested dose, QWx5) in a GLP-compliant repeat-dose toxicity study.
- Antibody exposure increased in a dose-proportional manner after the first dose (**Table 1**). Exposure was lower in most cases after the fourth dose, which was attributed to treatment-emergent anti-drug antibodies in all monkeys.

Table 1: Mean sex-combined toxicokinetic parameters

	Dose 1			Dose 4		
Dose (mg/kg)	5	25	50	5	25	50
C _{max} (mg/L)	148	810	1550	86.3	730	2010
AUC _{0-tau} (mg*d/L)	321	2040	4460	24.0	414	4070

 AUC_{0-tau} , area under the concentration-time curve during a single dosing interval; C_{max} , maximum observed concentration