

# Adoptive T cell therapy

John Haanen, MD, PhD

ESMO Asia 2015

NETHERLANDS  
CANCER  
INSTITUTE

ANTONI VAN LEEUWENHOEK

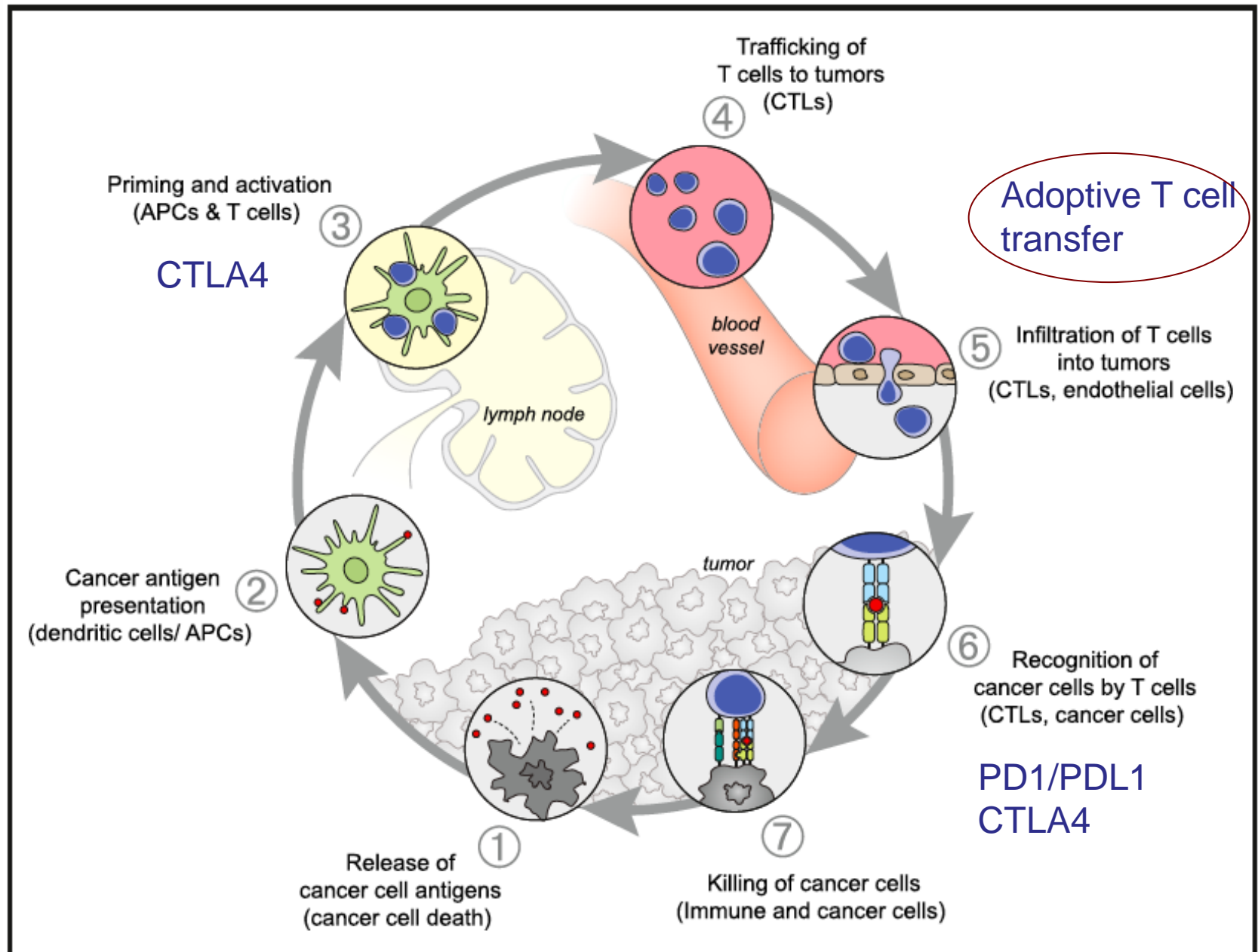


# Disclosures

Advisory role: BMS, Pfizer, Roche, MSD, Novartis  
Neon Therapeutics

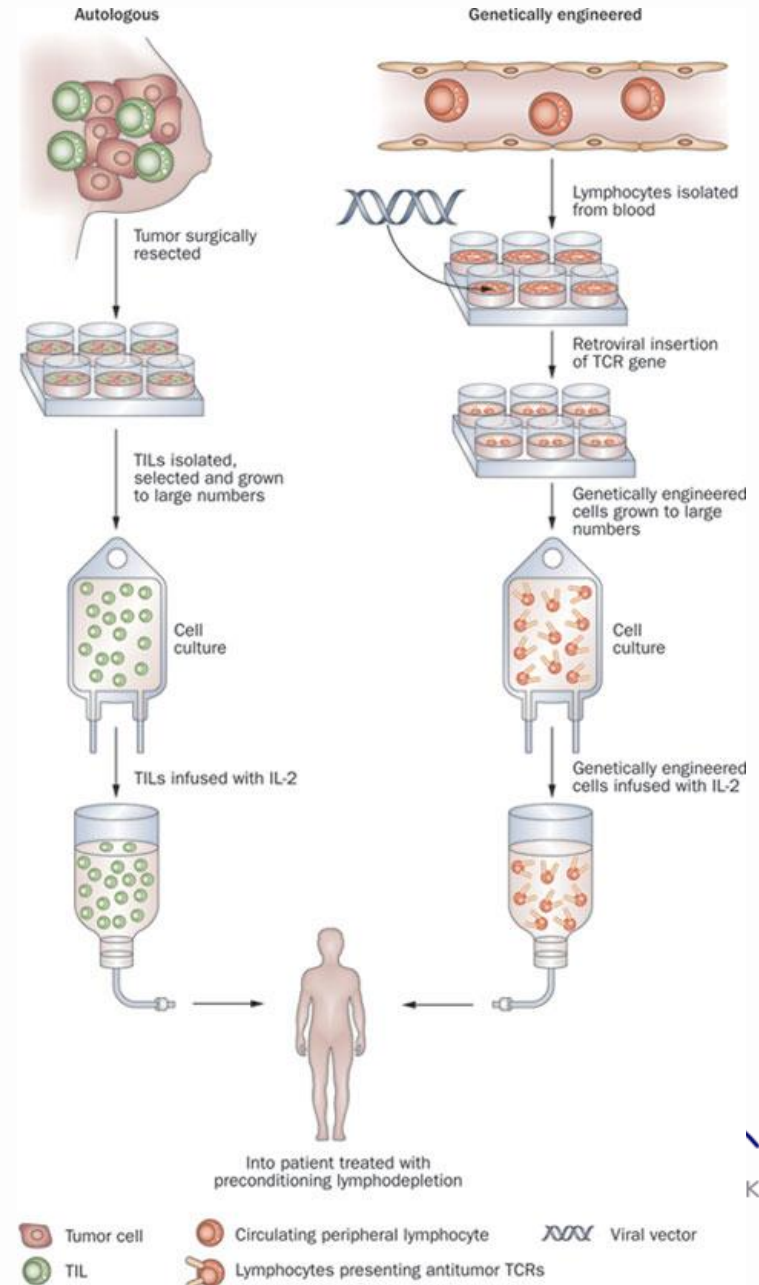
Research grant: GSK, BMS, MSD

# Cancer immunity cycle



# Different strategies available for adoptive cell therapy

- Tumor infiltrating lymphocytes: TIL therapy
- Genetically modified peripheral blood lymphocytes
  - inserting a tumor-reactive TCR
  - inserting a tumor-reactive CAR
    - using retroviral insertion
    - using lentiviral insertion
    - using transposon-based insertion



# Role for T cells in cancer

The NEW ENGLAND JOURNAL of MEDICINE

## ORIGINAL ARTICLE

### Intratumoral T Cells, Recurrence, and Survival in Epithelial Ovarian Cancer

Lin Zhang, M.D., Jose R. Conejo-Garcia, M.D., Ph.D.,  
Dionyssios Katsaros, M.D., Ph.D., Phyllis A. Gimotty, Ph.D.,  
Marco Massobrio, M.D., Giorgia Regnani, M.D.,  
Antonis Makrigiannakis, M.D., Ph.D., Heidi Gray, M.D.,  
Katia Schlienger, M.D., Ph.D., Michael N. Liebman, Ph.D.,  
Stephen C. Rubin, M.D., and George Coukos, M.D., Ph.D.

## PERSPECTIVES

### OPINION

### The immune contexture in human tumours: impact on clinical outcome

Wolf Herman Fridman, Franck Pagès, Catherine Sautès-Fridman and  
Jérôme Galon



### Type, Density, and Location of Immune Cells Within Human Colorectal Tumors Predict Clinical Outcome

Jérôme Galon, *et al.*  
*Science* **313**, 1960 (2006);  
DOI: 10.1126/science.1129139

## Cancer Research

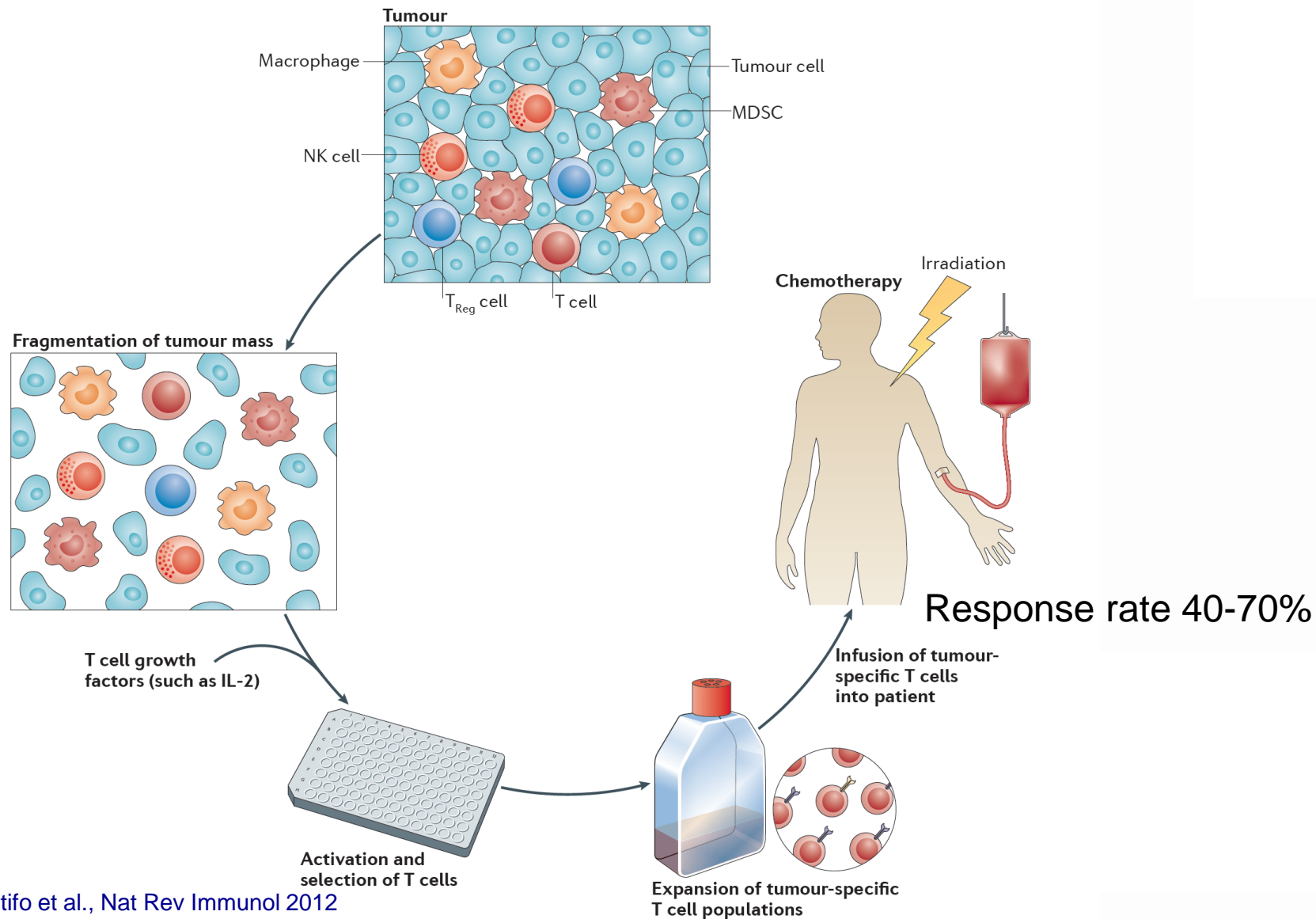


### Immunotype and Immunohistologic Characteristics of Tumor-Infiltrating Immune Cells Are Associated with Clinical Outcome in Metastatic Melanoma

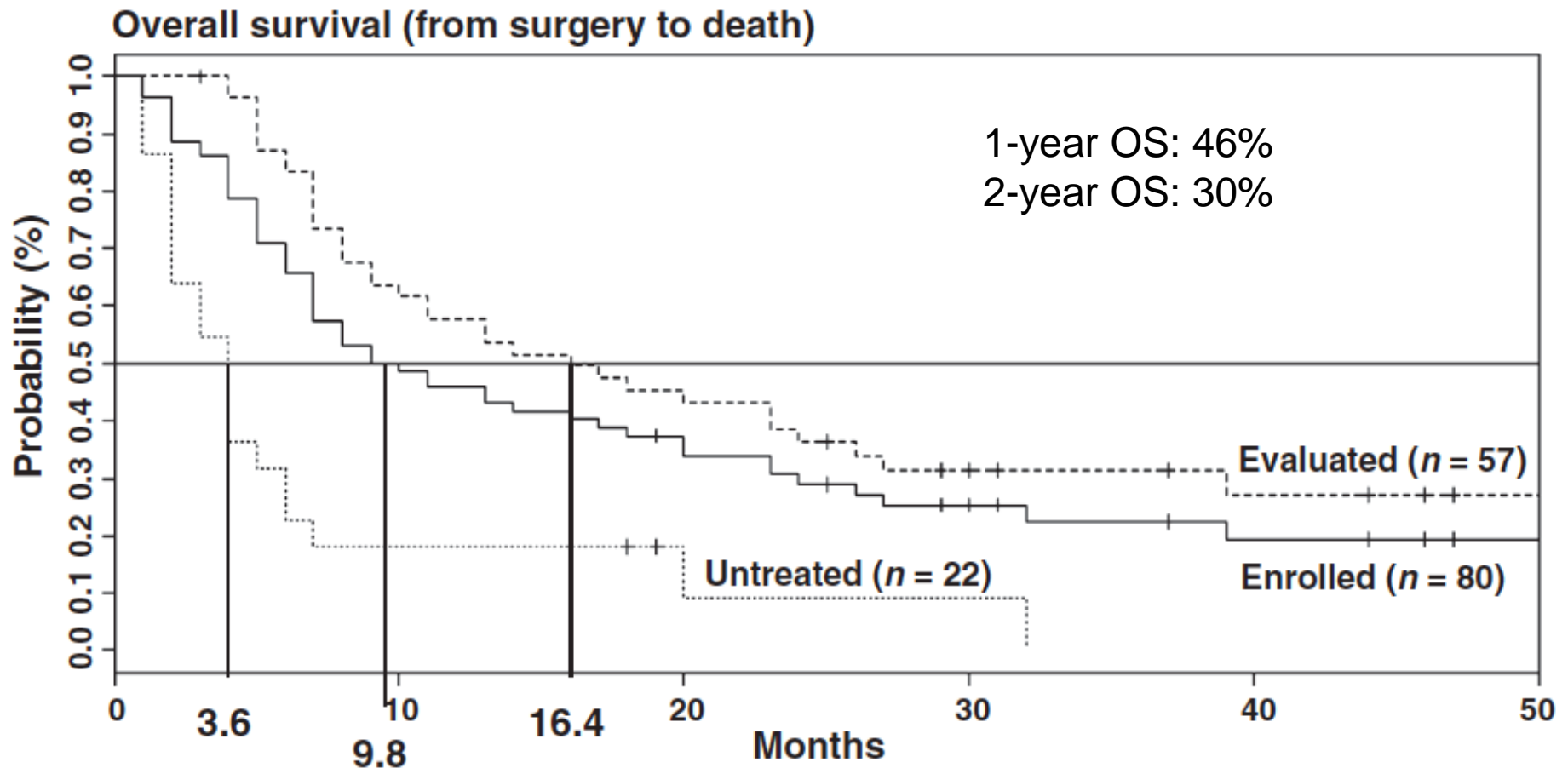
Gulsun Erdag, Jochen T. Schaefer, Mark E. Smolkin, et al.

*Cancer Res* 2012;72:1070-1080. Published OnlineFirst January 19, 2012.

# Tumor infiltrating lymphocytes: TIL therapy in melanoma



# Overall survival of metastatic melanoma patients treated with TIL (ITT analysis)

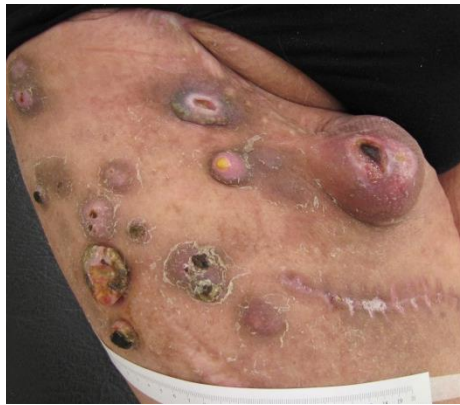


# Clinical data N10TIL003: ongoing complete response 4 years

Prior to TIL



3 wks post TIL



8 wks post TIL



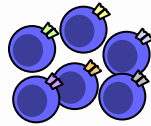
12 wks post TIL



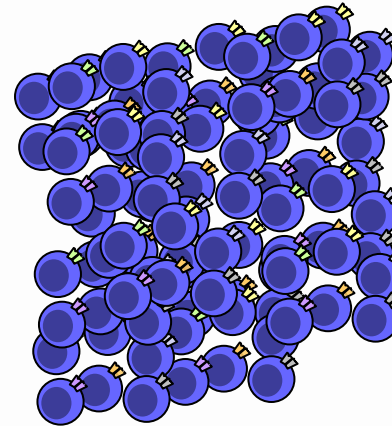
Biopsy at wk 7 showed no viable tumor cells

# Immunotherapy of melanoma: TIL therapy

TIL are grown  
from melanoma  
tumors



Rapid Expansion



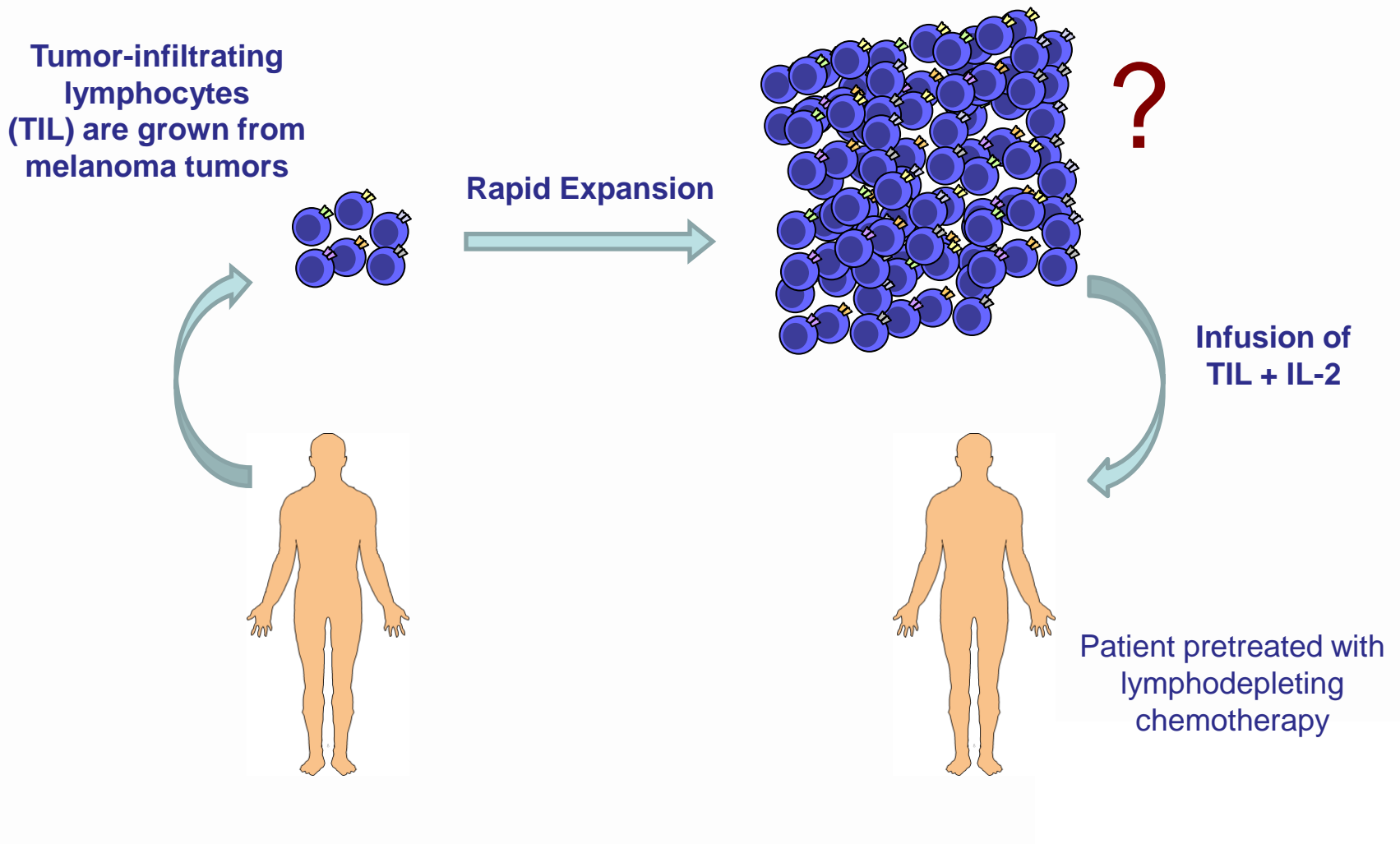
A few million T cells



$1 \times 10^{11}$  T cells

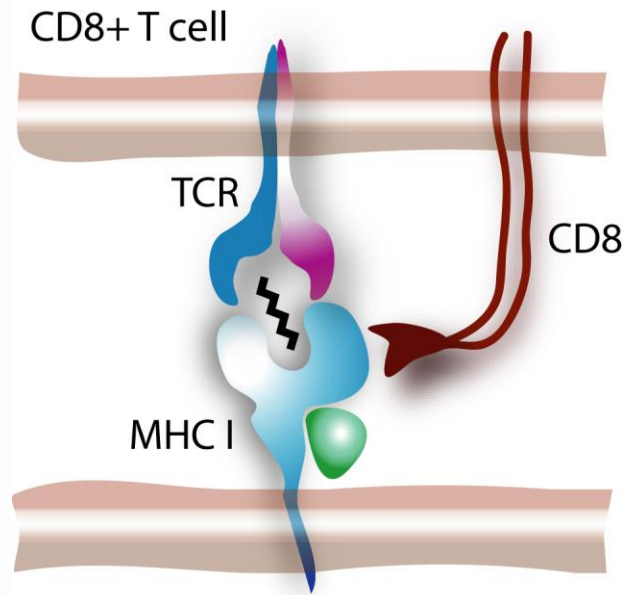


# The big unknown



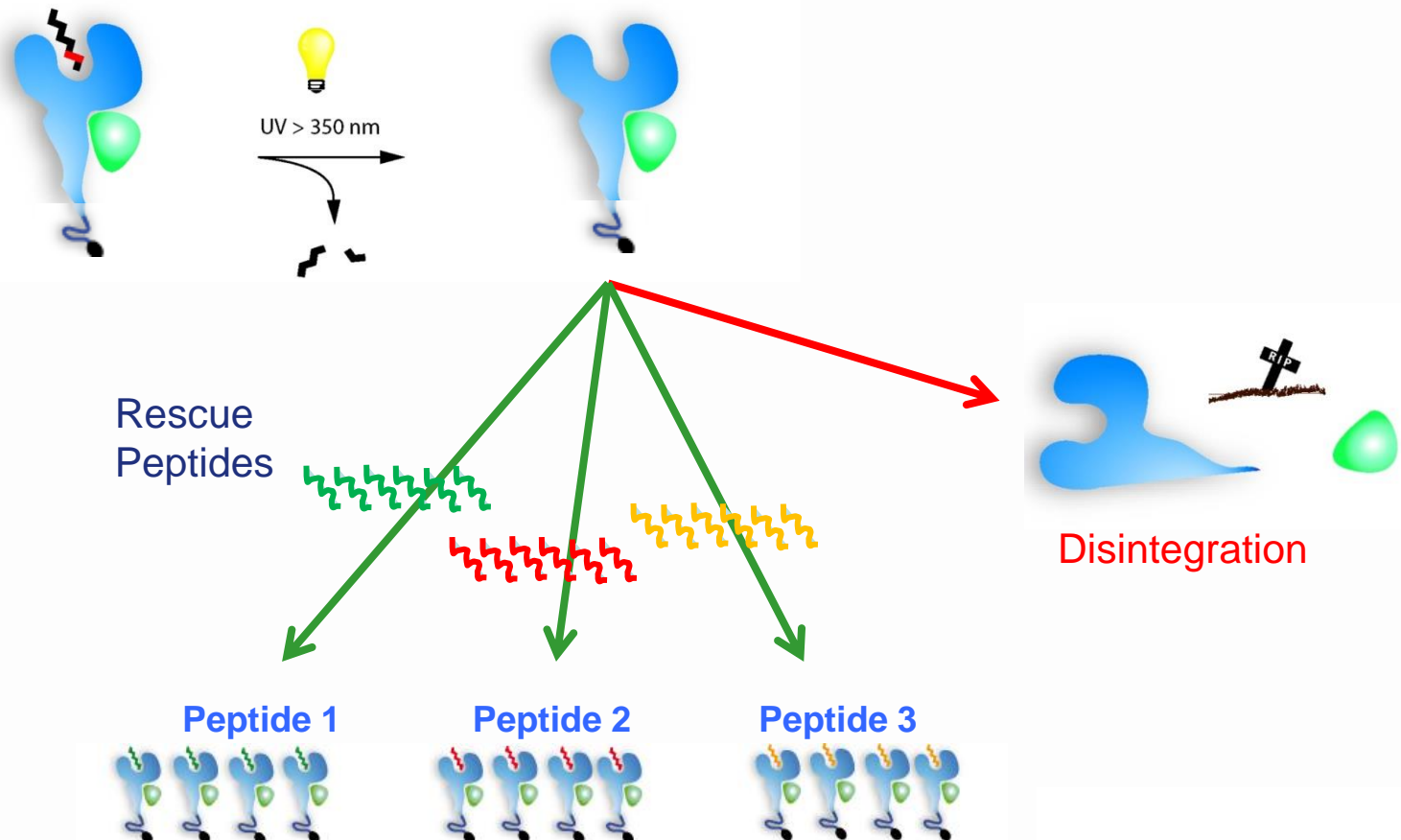
- Which cytotoxic T cells mediate cancer regression?
- Could we specifically boost their numbers?

# What could tumor-specific cytotoxic T cells detect on human cancer?



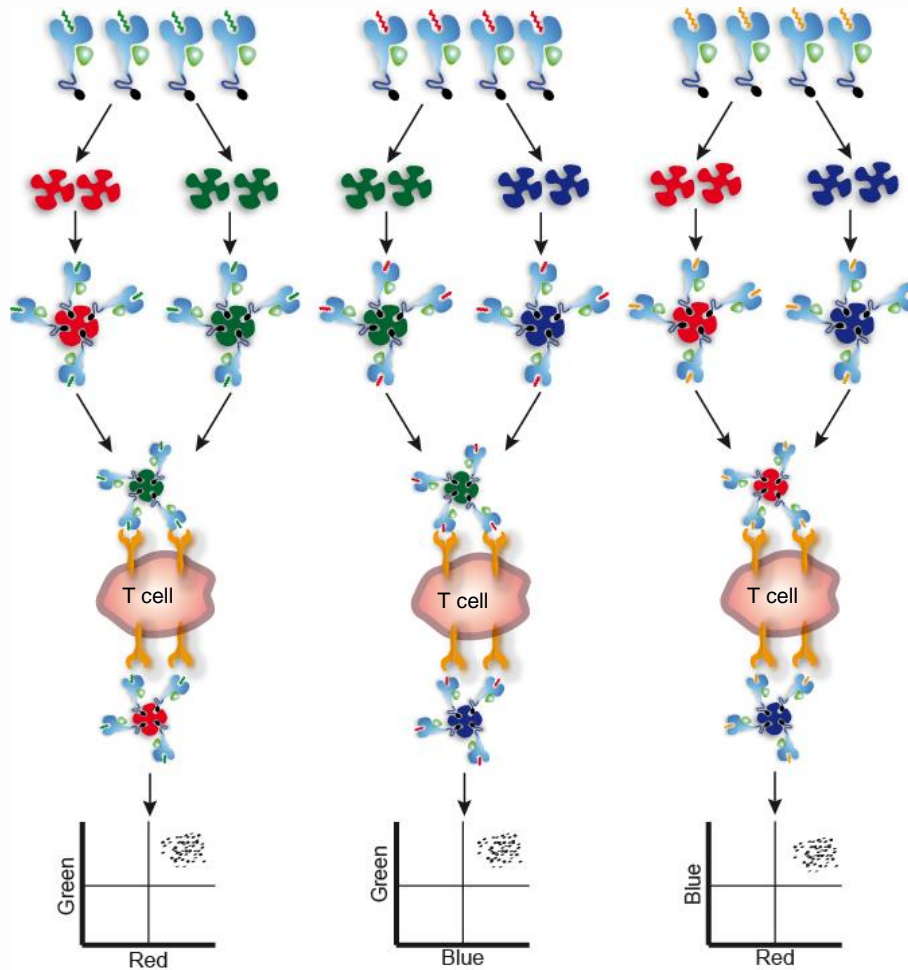
1. Self antigens (to which tolerance is incomplete)  
*Shared between patients*
2. 'Neo-antigens', epitopes that arise as a consequence of tumor-specific mutations  
*In large part patient-specific, hence generally ignored*

# Generation of pMHC multimers by UV-induced peptide exchange



Allows generation of 1000s of pMHC in parallel

## Self-assembling molecular codes



Generate fluorochrome conjugated MHC multimers

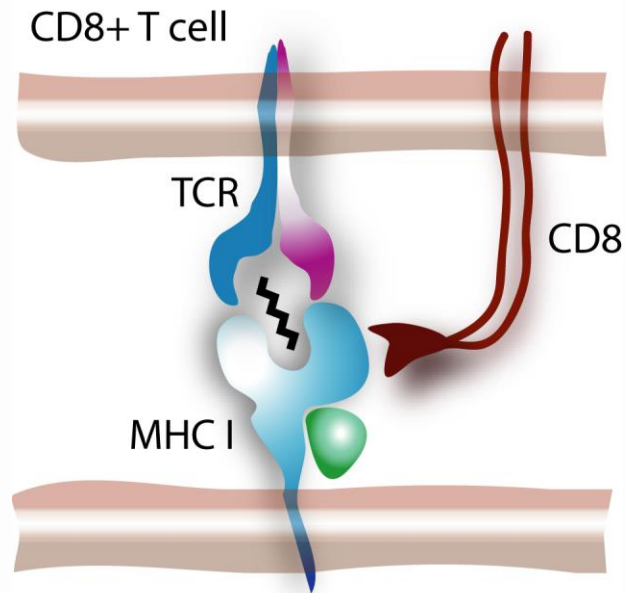
Mix to create a collection of differentially encoded MHC multimers

Assembly of combinatorial codes on T cell surfaces

Analysis by flow cytometry

Allows detection of 47 T cell responses in parallel

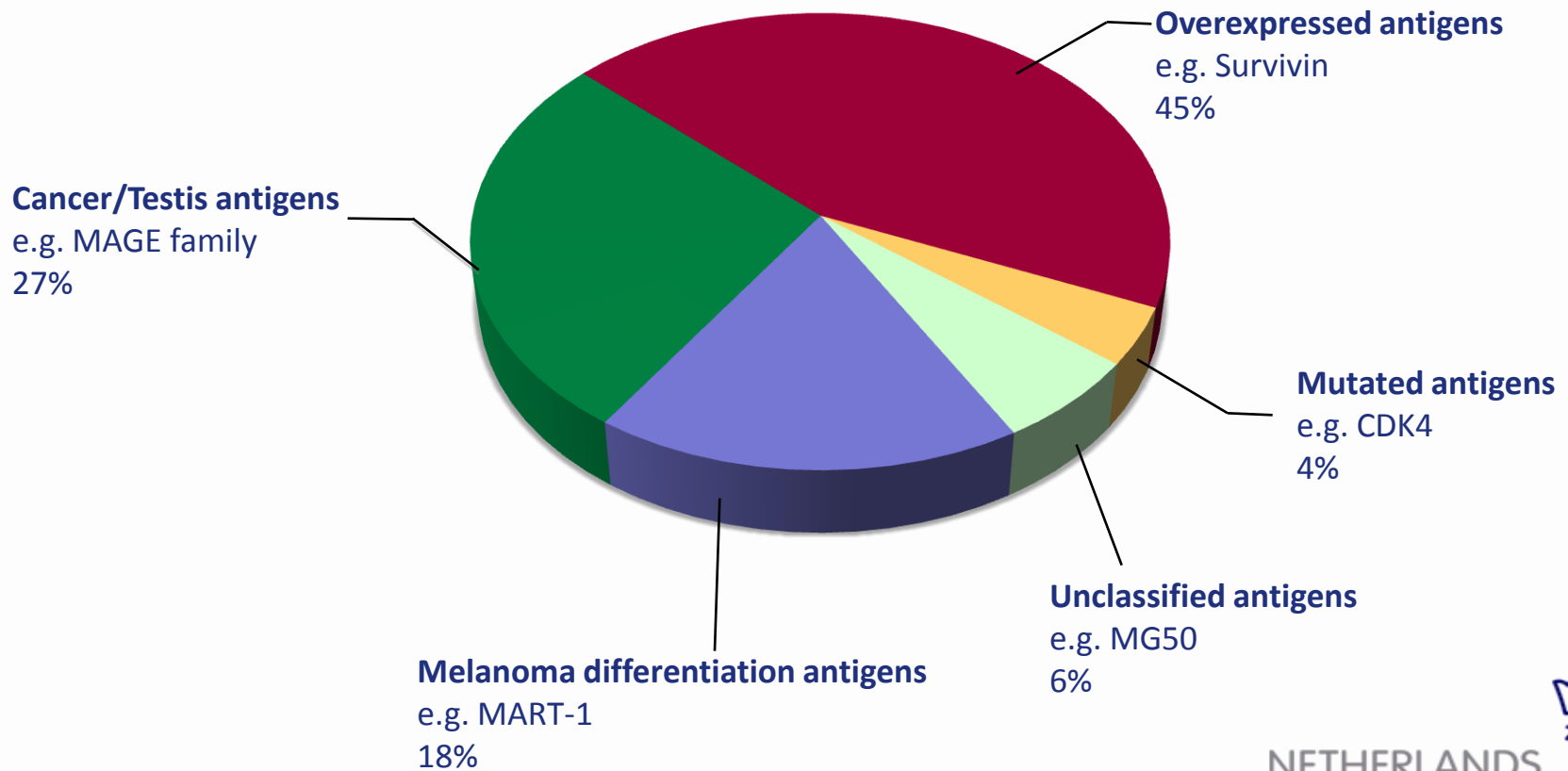
# What could tumor-specific cytotoxic T cells detect on human cancer?



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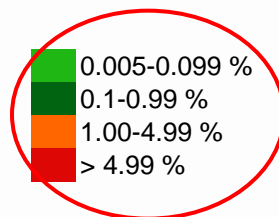
# Melanoma associated epitope panel

HLA-A2 restricted peptide panel includes 145 epitopes



# Visualizing the composition of TIL

T cell responses are very low magnitude



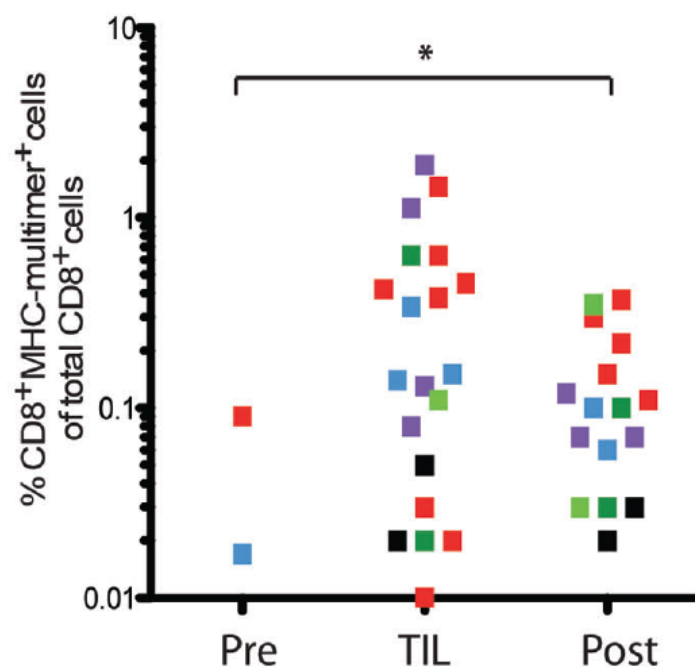
	Young CD8 enriched TIL (NIH)																	Young TIL (Ella)						Selected TIL													
Patient	LR	LN	KS	MV	CR	ER	MA	LD	SW	HE	AS	RE	OJ	AS	MG	NJ	BJ(f)*	SC*	57SV	63SM	51VS	60SD	41BA	52SD	14PA	09BY	31YR	PS	OM	BJ(m)	SM	ER	CR	BM	CJ	AD	
Mart-1 <sub>ELA</sub>																																					
gp100 <sub>IMD</sub>																																					
gp100 <sub>YLE</sub>																																					
gp100 <sub>VLY</sub>																																					
gp100 <sub>AML</sub>																																					
gp100 <sub>KTW</sub>																																					
TRP2 <sub>VYD</sub>																																					
TRP2 <sub>SVY</sub>																																					
NY-MEL 1 <sub>VLH</sub>																																					
CML28 <sub>AVL</sub>																																					
Mage A4 <sub>GVY</sub>																																					
MAGE A10 <sub>GLY</sub>																																					
MageB1, B2 <sub>FLW</sub>																																					
MAGE C2 <sub>LLF</sub>																																					
MAGE C2 <sub>ALK</sub>																																					
NY-ESO 1 <sub>SLL</sub>																																					
HERV K mel <sub>MLA</sub>																																					
SSX-2 <sub>KAS</sub>																																					
GnTV <sub>VLP</sub>																																					
GnT-V <sub>VLP10mer</sub>																																					
Meloe-1 <sub>TLN</sub>																																					
Telomerase <sub>RLF</sub>																																					
Bing 4 <sub>CQW</sub>																																					
Clinical response	NR	NR	NR	PR	PR	NR	NR	PR	PR	PR	NR	NR	NR	NR	PR	NR	NR	CR	PR	NR	PR	NR	NR	PR	CR	NR	NR	PR	PR	CR	PR	PR	NR	NR	CR	NR	

# TILs against shared tumor antigens

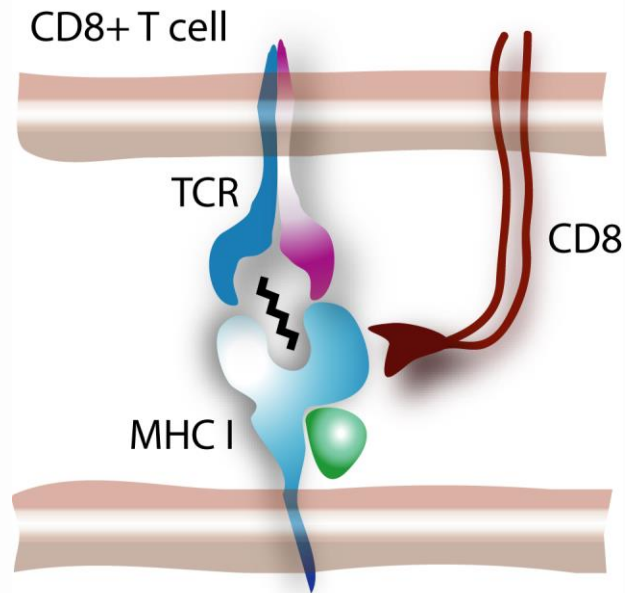
- In the majority of TILs T cells specific for shared antigens can be found
  - Melanocyte differentiation Ags (Mart-1, gp100, etc)
  - Cancer/Testis gene products (NY-eso-1, MAGE, SSX-2, etc)
  - Overexpressed Ags (Meloe etc.)
- Low frequency (mostly below 1%)
- No correlation with response

# TIL therapy broadens the tumor-reactive CD8<sup>+</sup> T cell compartment in melanoma patients

Pia Kvistborg,<sup>1,†</sup> Chengyi Jenny Shu,<sup>1,†</sup> Bianca Heemskerk,<sup>1</sup> Manuel Fankhauser,<sup>1</sup> Charlotte Albæk Thue,<sup>2</sup> Mireille Toebes,<sup>1</sup> Nienke van Rooij,<sup>1</sup> Carsten Linnemann,<sup>1</sup> Marit M. van Buuren,<sup>1</sup> Jos H.M. Urbanus,<sup>1</sup> Joost B. Beltman,<sup>3</sup> Per thor Straten,<sup>2</sup> Yong F. Li,<sup>4</sup> Paul F. Robbins,<sup>4</sup> Michal J. Besser,<sup>5,6</sup> Jacob Schachter,<sup>5</sup> Gemma G. Kenter,<sup>7</sup> Mark E. Dudley,<sup>4</sup> Steven A. Rosenberg,<sup>4</sup> John B.A.G. Haanen,<sup>1</sup> Sine Reker Hadrup<sup>2</sup> and Ton N.M. Schumacher<sup>1,\*</sup>

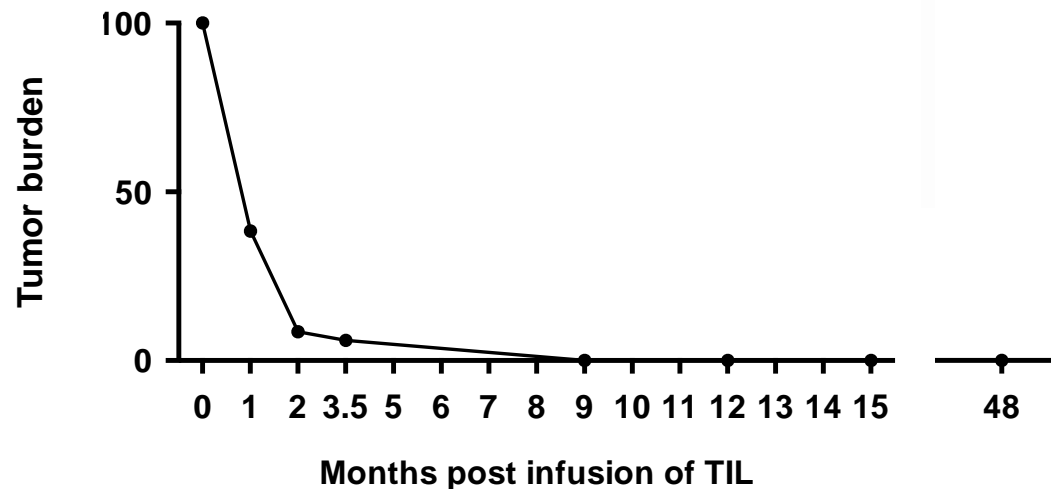
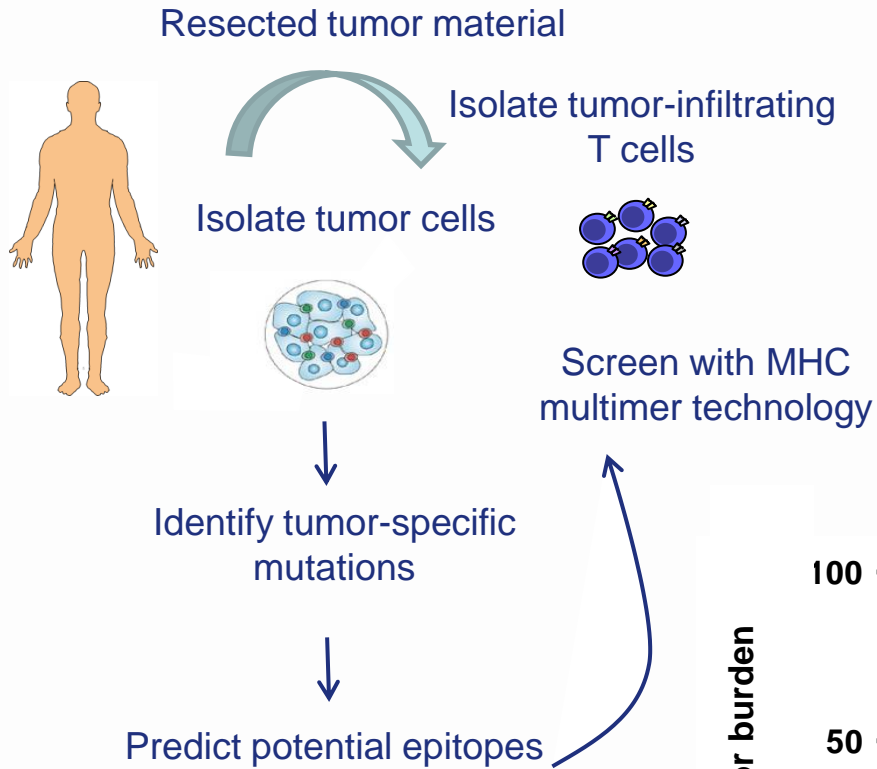


# What could tumor-specific cytotoxic T cells detect on human cancer?



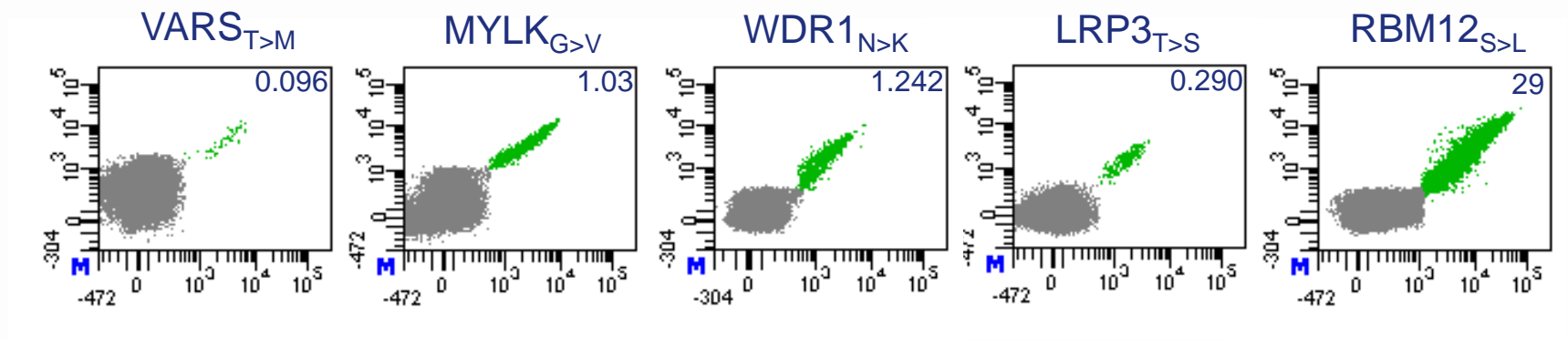
1. Self antigens (to which tolerance is incomplete)  
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# Pt 003: complete response upon TIL therapy



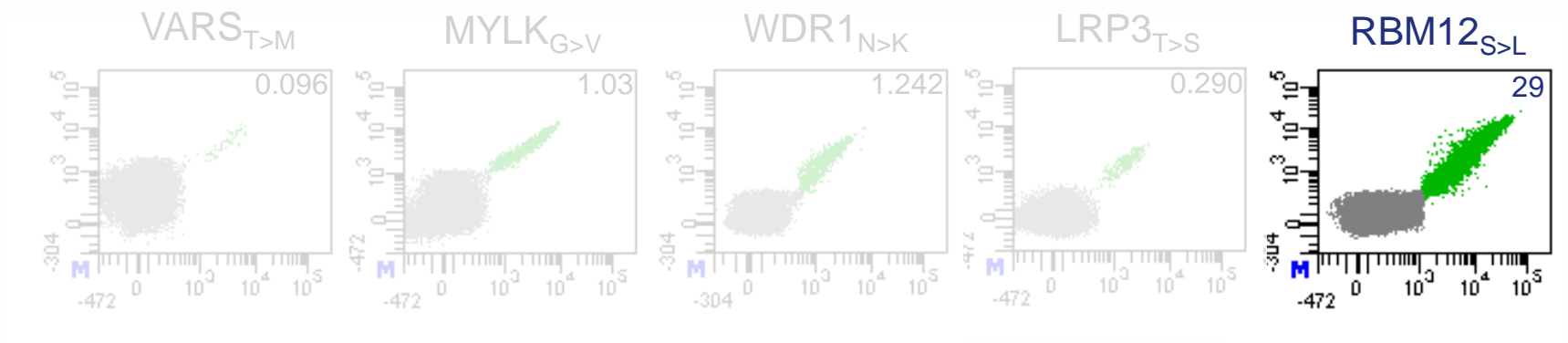
# Pt 003: complete response upon TIL therapy

TIL infusion product

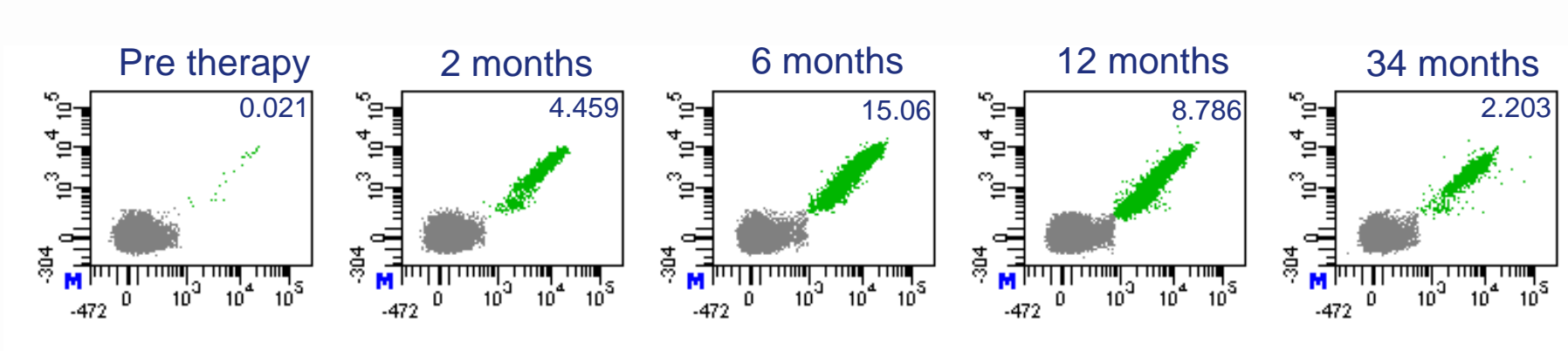


# Pt 003: complete response upon TIL therapy

## TIL infusion product

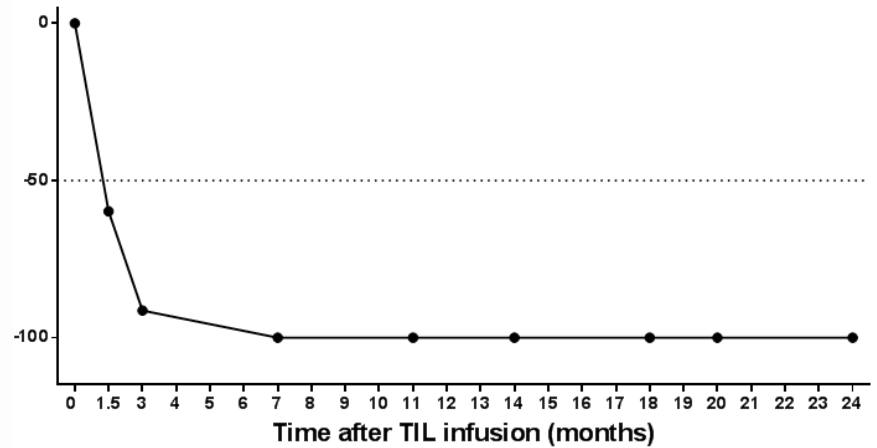
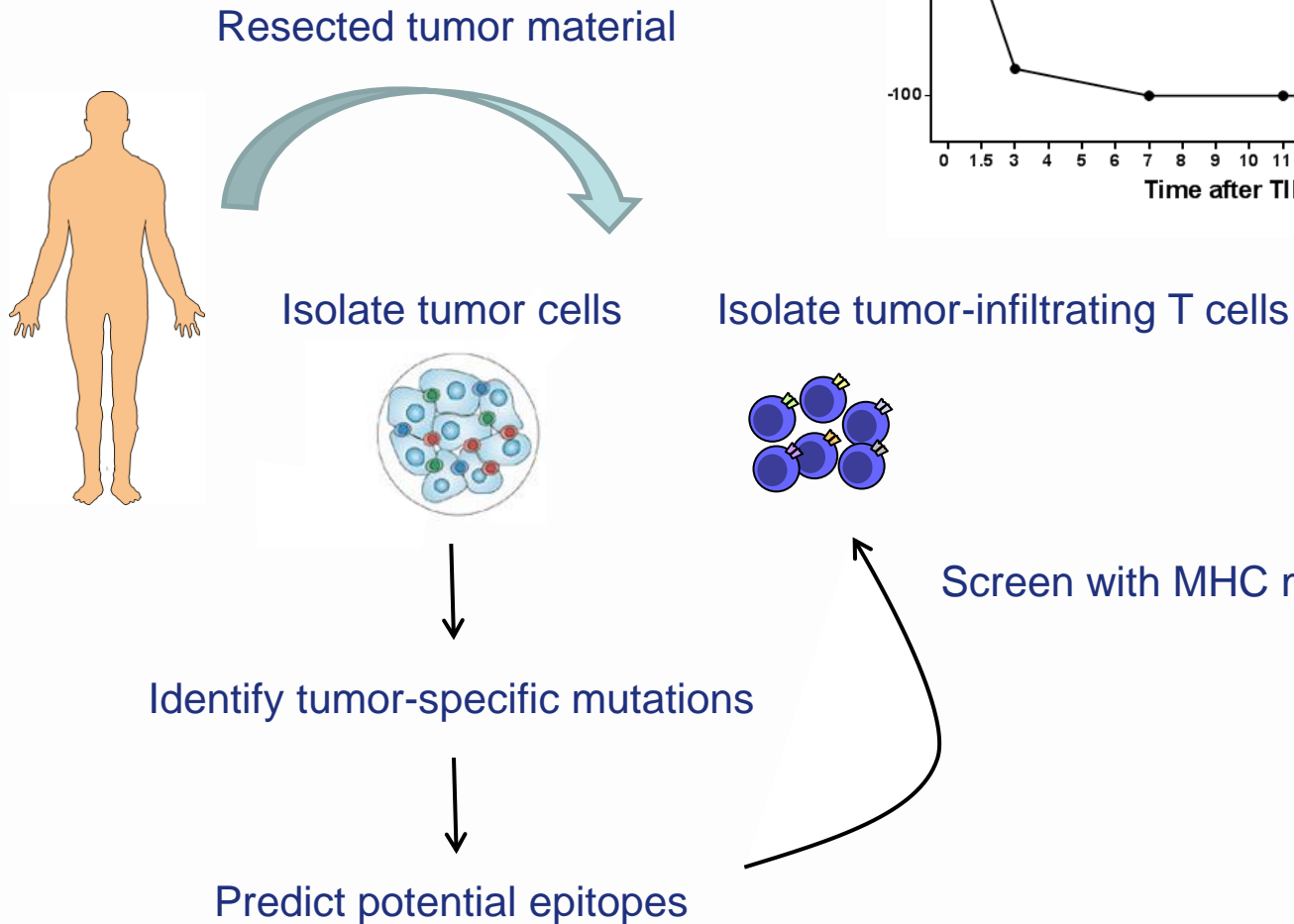


## Peripheral blood



>450 fold increase in neo-antigen specific T cell reactivity upon TIL therapy

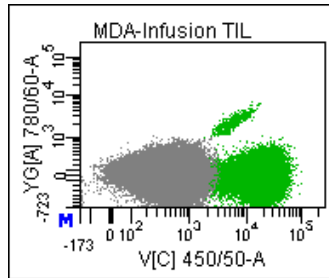
# Pt 008: CR upon TIL therapy



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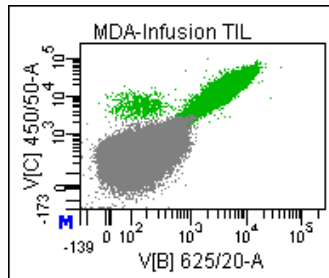
## Infusion TIL product

0.172%



**RASSF1<sub>R>C</sub>**

23%



**DHX33<sub>R>W</sub>**

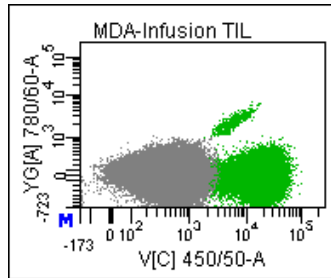
# Pt 008: CR upon TIL therapy

Infusion TIL product

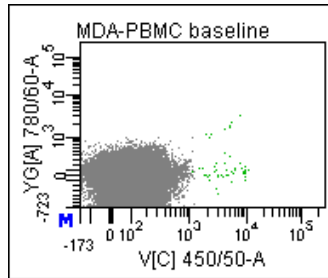
Pre-therapy PBMNC

D7 post-therapy

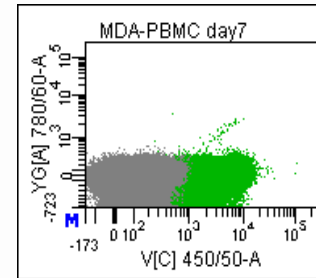
0.172%



0.002%

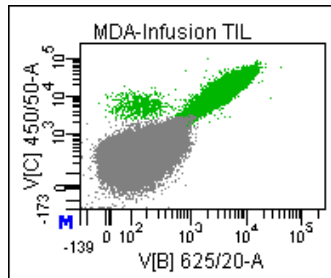


0.010%

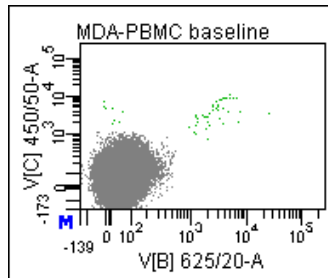


RASSF1<sub>R>C</sub>

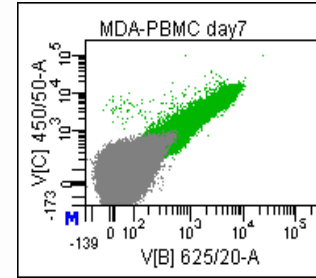
23%



0.009%



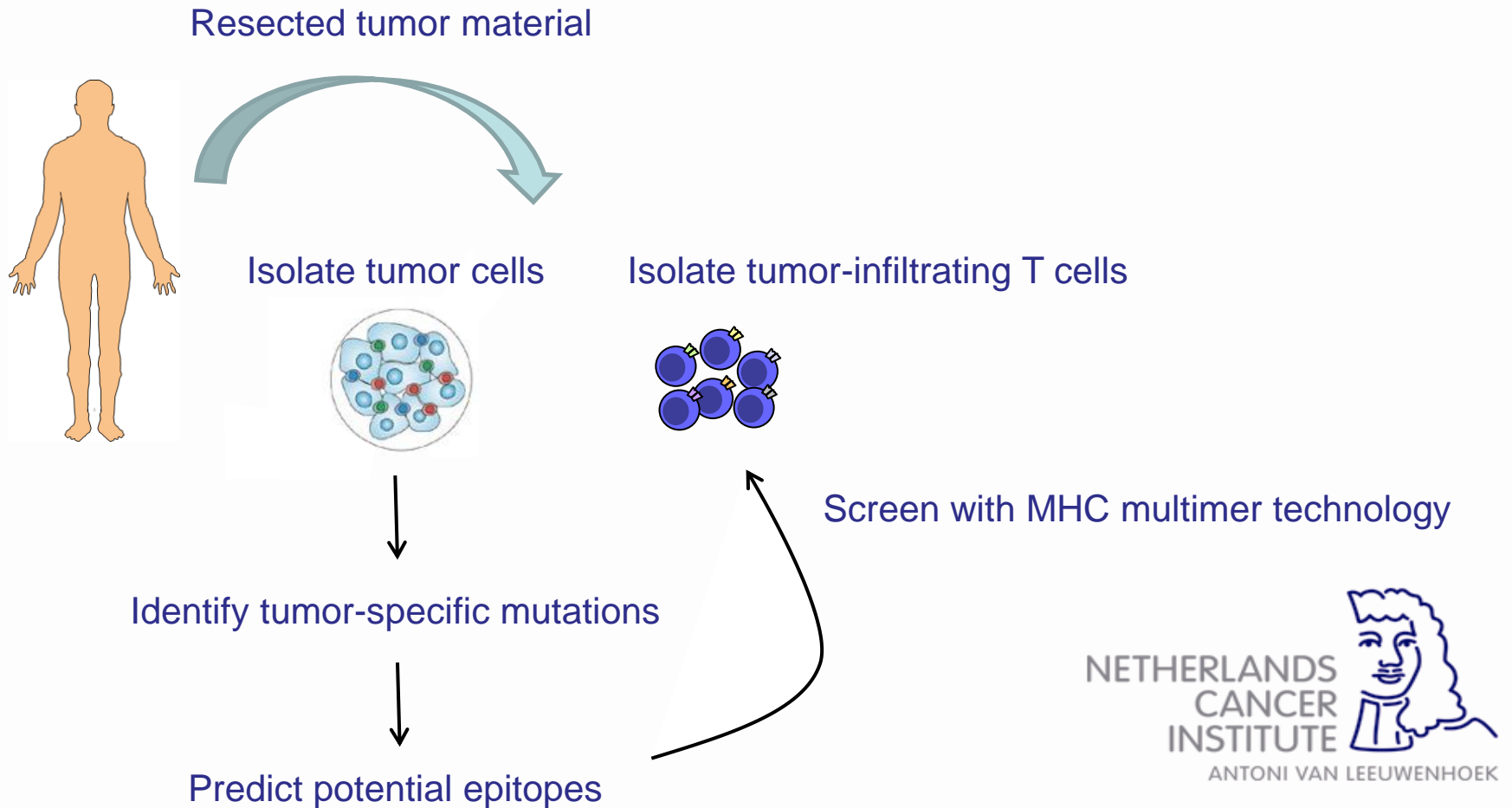
54%



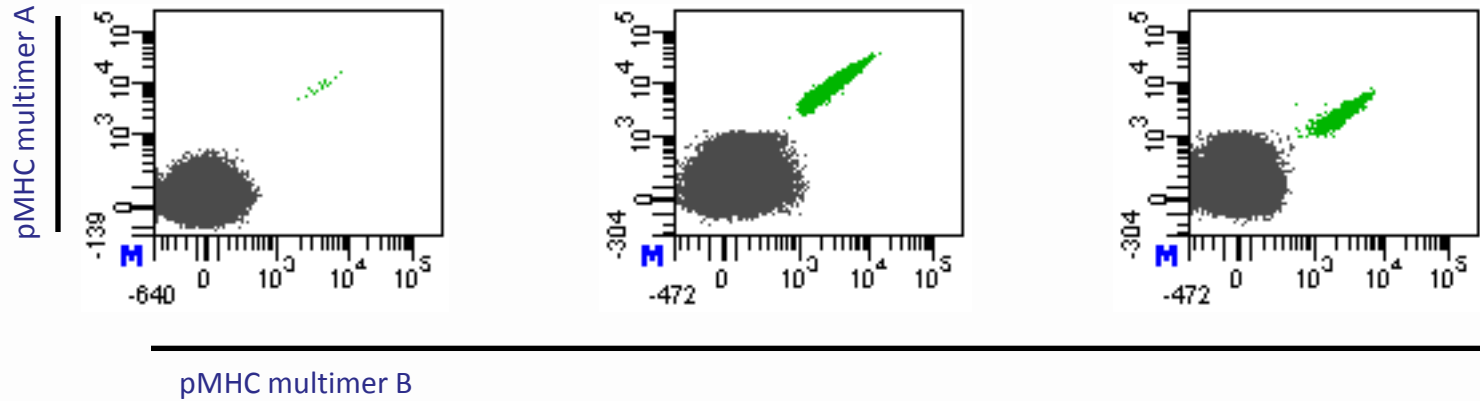
DHX33<sub>R>W</sub>

Major (>5000 fold) increase in neo-antigen specific T cell reactivity upon TIL therapy

# Pt 004:



# Pt 004:

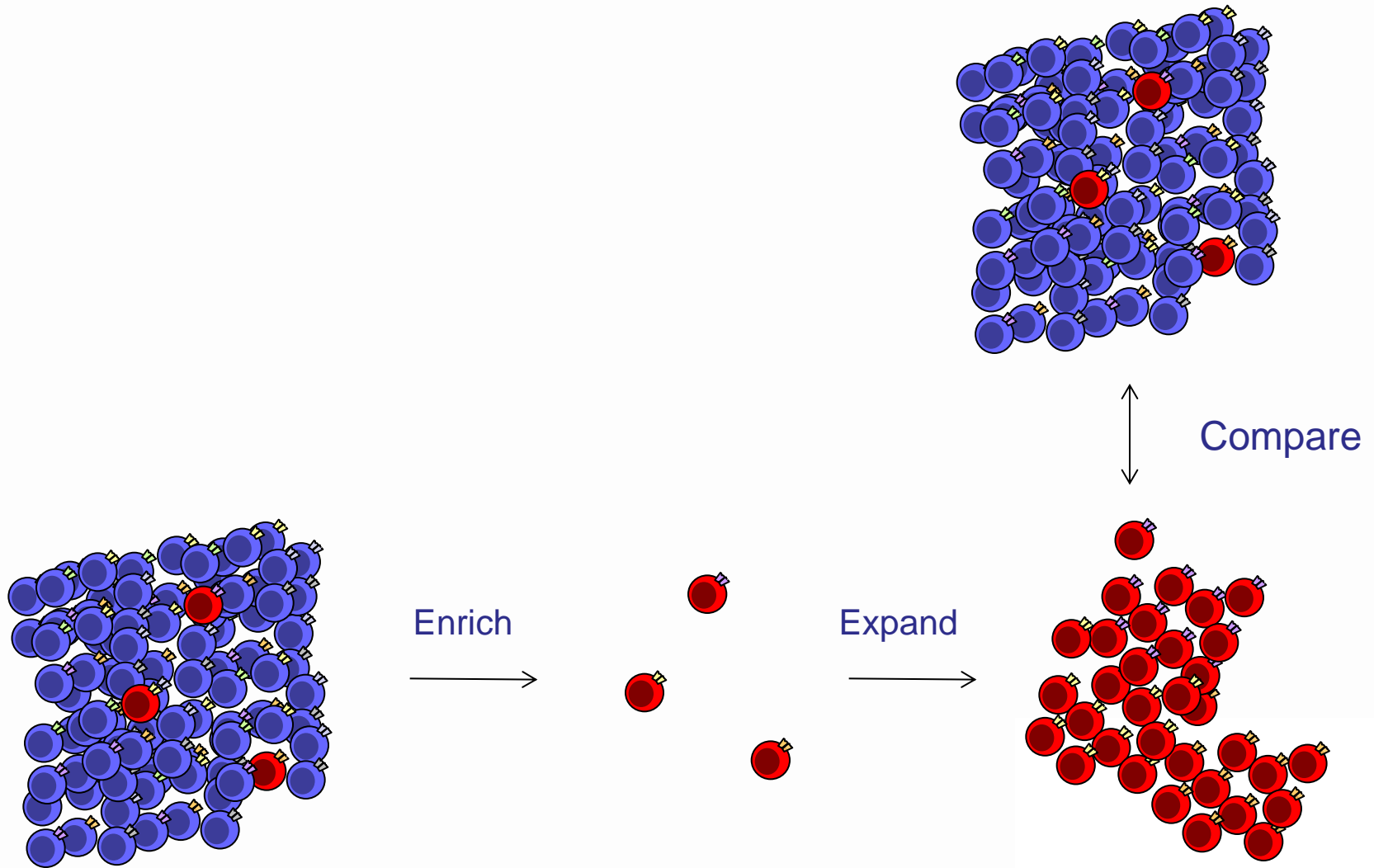


**DNAH17<sub>H>Y</sub> (0.003%)**  
**VLFEDAVAHH > VLFEDAVAYY**

**CDK4<sub>R>L</sub> (1.604%)**  
**ARDPHSGHFV > ALDPHSGHFV**

**GCN1L1<sub>L>P</sub> (0.407%)**  
**ALLETLSLLL > ALLETPSLLL**

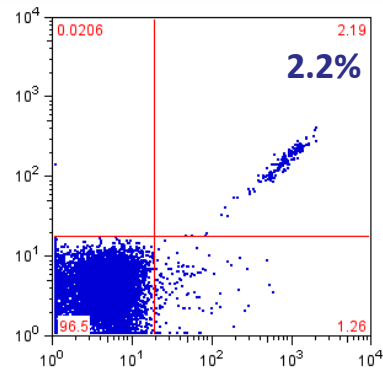
# Are neo-antigens superior cancer rejection antigens?



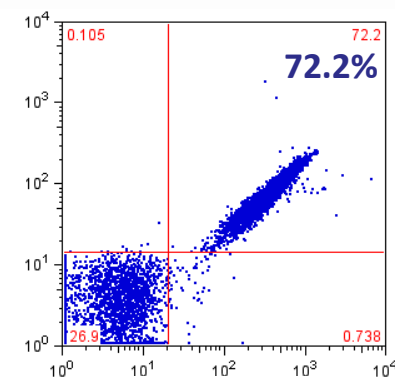
# Are neo-antigens superior cancer rejection antigens?

**CDK4<sub>R>L</sub>**

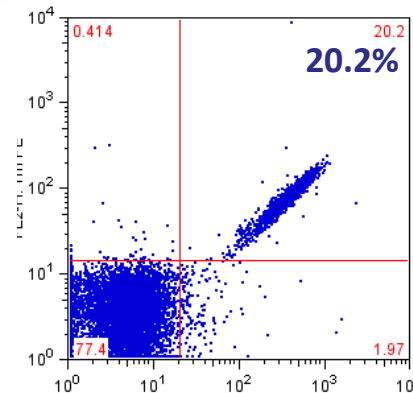
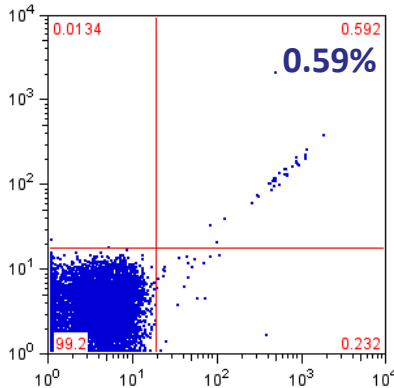
**Pre-enrichment**



**Post-joint enrichment**



**GCN1L1<sub>L>P</sub>**

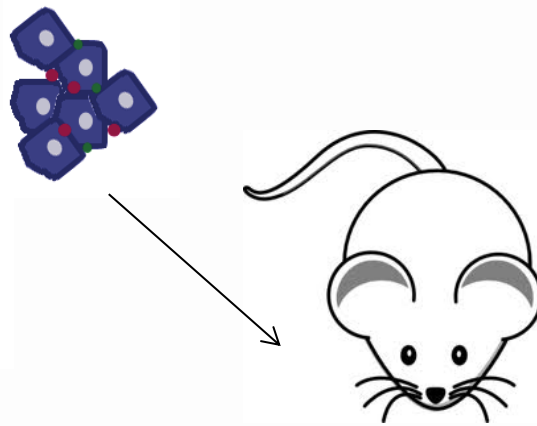


**Combined**

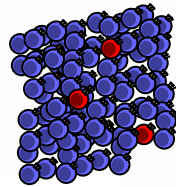
**2.8%**

**92.4%**

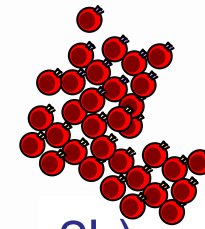
1)  
Inject human  
melanoma  
(NSG-mice)



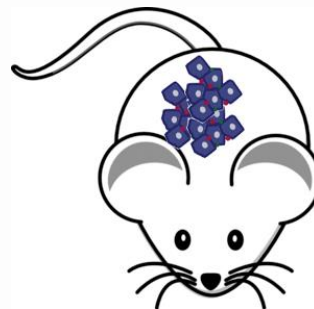
2a)  
Inject autologous  
bulk T-cell product



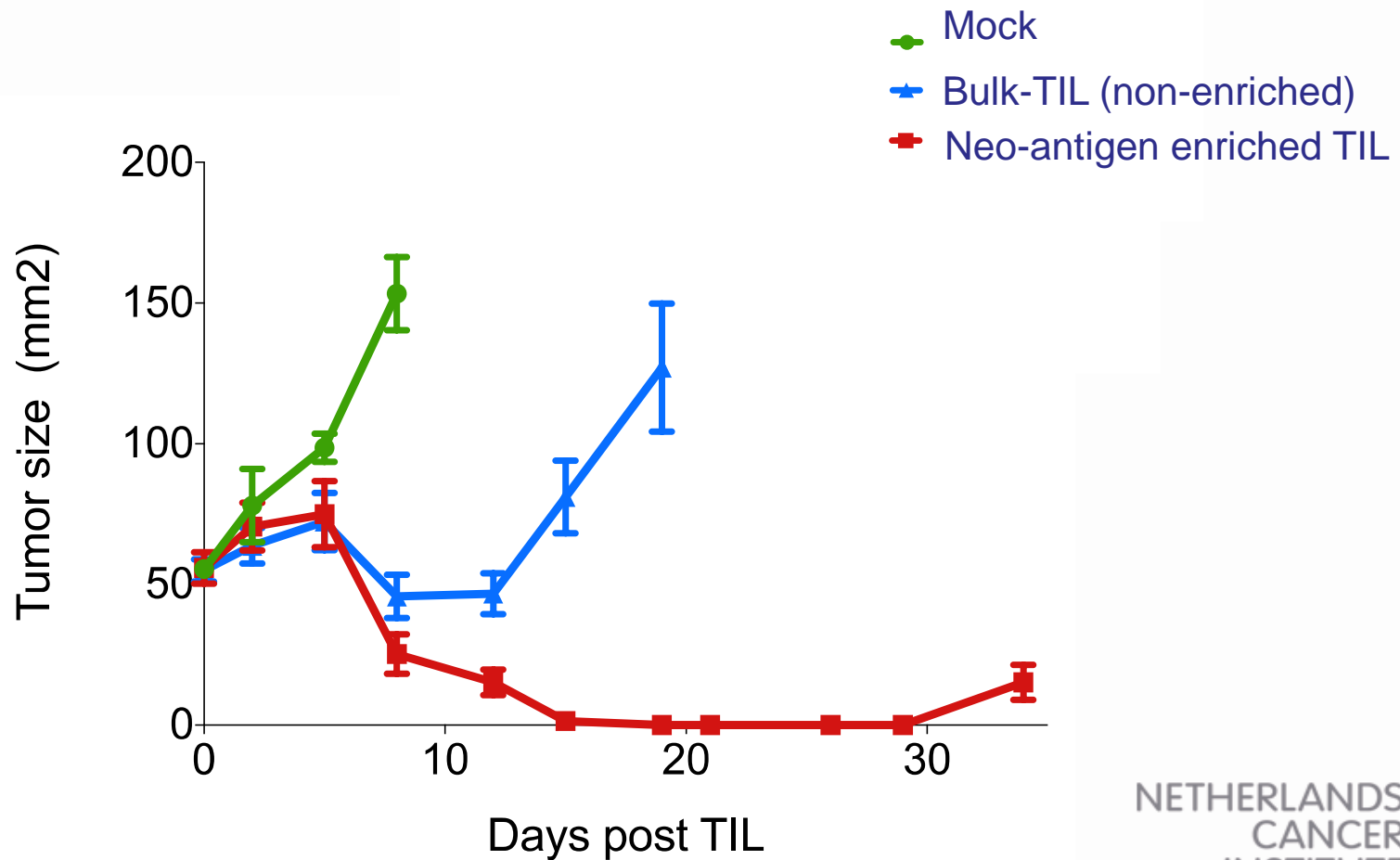
2b)  
Inject autologous  
neo-Ag enriched  
T-cell product



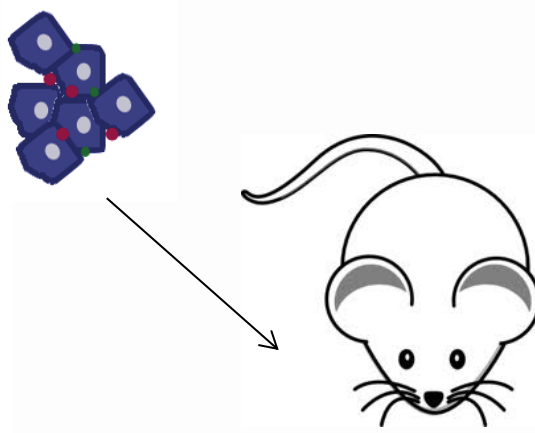
3)  
Monitor tumor growth



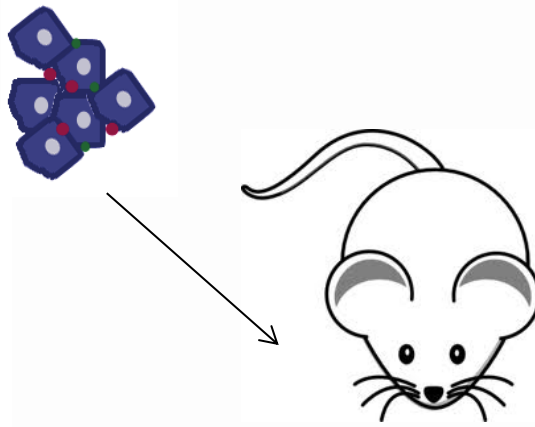
# Neo-antigen enriched TIL can mediate superior tumor control



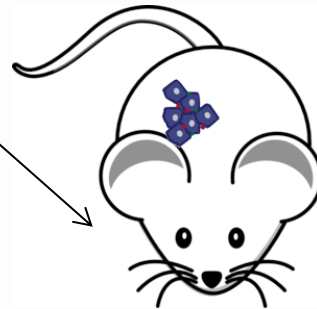
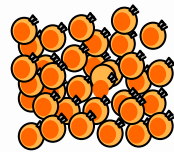
**1)**  
**Create human**  
**melanoma**  
**PDX model**  
**(NSG-mice)**



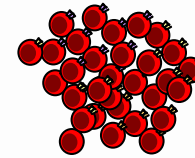
**1)  
Create human  
melanoma  
PDX model  
(NSG-mice)**



**2a)  
treat with T cells  
transduced with  
autologous C/T Ag  
specific TCRs**



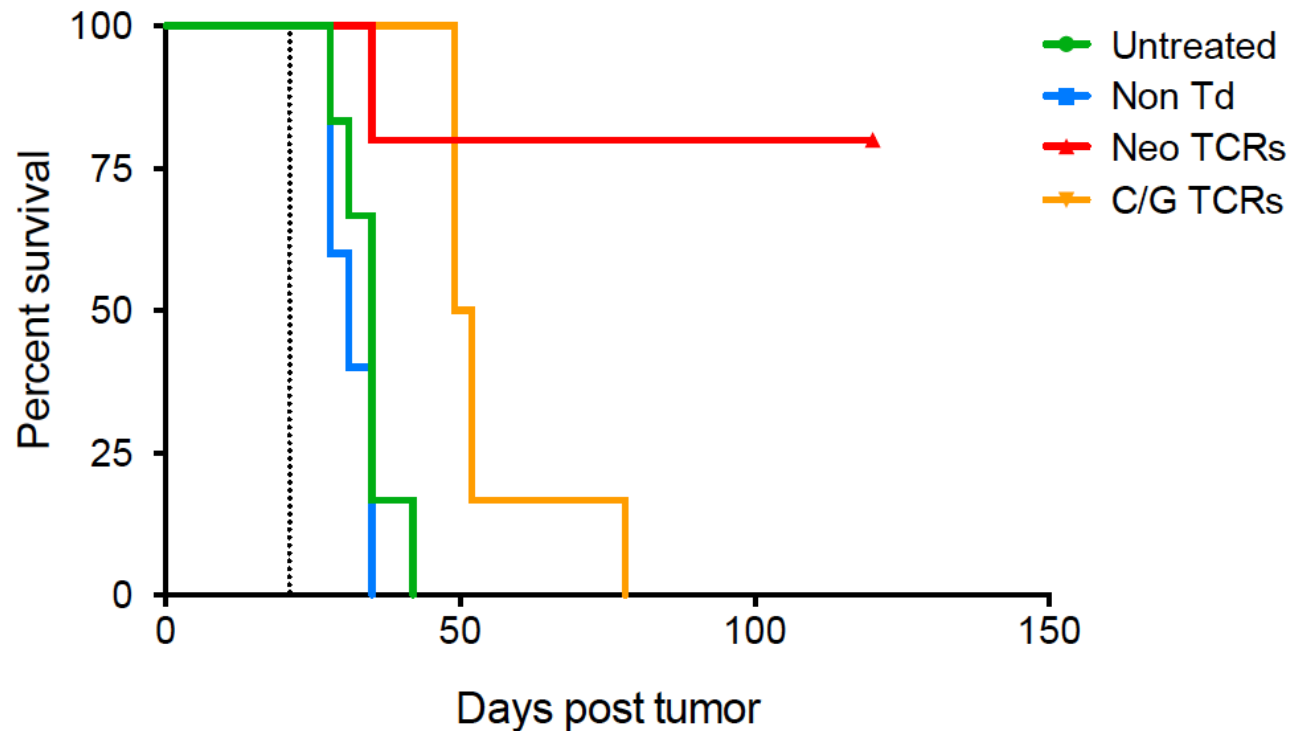
**2b)  
treat with T cells  
transduced with  
autologous Neo Ag  
specific TCRs**



Assess whether neo-antigen specific TCRs outperform C/G specific TCRs

# Do neo-antigen specific TCRs\* outperform C/G antigen specific TCRs\*\*?

*caution: n=1 expt, repeat ongoing*



\* 2 TCRs, against CDK4 and GCN1L1 neo-antigens

\*\* 4 TCRs, against 3 MAGE-C2 epitopes, 1 MAGE-A10 epitope

# What have we learned from TIL therapy?

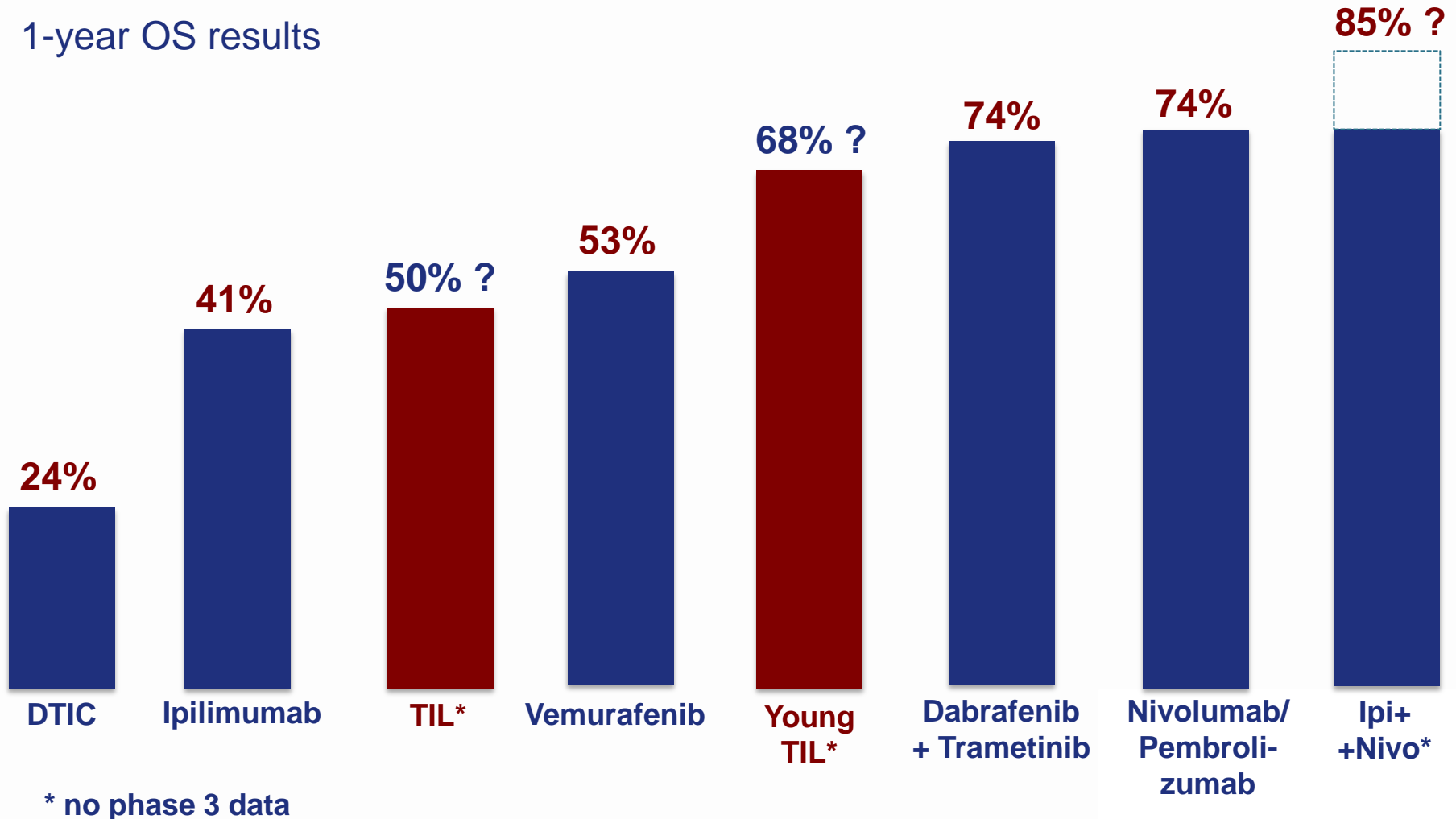
- TIL contain oftentimes many melanoma-specific CD8 and CD4 T cell populations
  - Against shared antigens (MDA, C/T, overexpressed)
  - Against neo-antigens
- Upon infusion of TIL, the tumor-reactive CD8 and CD4 T cell compartment is broadened in melanoma patients

# What have we learned from TIL therapy?

- Objective clinical response rates vary between 38% and 72% of treated melanoma patients in phase II clinical trials (mostly heavily pretreated pts)
- Median OS in this group 16 months
- Patients with CR upon TIL have an excellent prognosis

# How does TIL compare to other therapies?

1-year OS results



*McArthur et al. Lancet Oncol 2014; Hodi et al. NEJM 2010; based on Rosenberg and Dudley Curr Opin Immunol 2009; McArthur et al. Lancet Oncol 2014; based on Dudley et al. JCO 2013; Long et al. Lancet 2015; based on Robert et al. NEJM 2015; prediction based on Larkin et al. NEJM 2015 and Sznol et al. ASCO 2014*

# Comparison between TIL and checkpoint inhibitors

- TIL: one treatment
- Surgery is required
- Complex GMP and patient specific production process
- Drop-out rate up to 25% of pts
- In hospital (2-3 weeks)
- Predictable and manageable side-effects
- High treatment costs
- Ipilimumab: 4 infusions
- Anti-PD1: >>4 infusions
- Off-the-shelf product
- No ipilimumab in LDH > 2x ULN
- Outpatient clinic
- Unpredictable, but manageable side effects
- Even higher treatment costs

# How to further develop TIL therapy

1. Approval of TIL therapy as treatment option for MM
  - RCT
  - A large phase II trial in checkpoint inh failing pts
2. Enrichment for tumor-reactive TIL
3. Generate a personalized TIL product
4. Expand TIL therapy beyond melanoma

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# Taking the next step for TIL based ACT

Randomized phase III study comparing TIL based ACT to standard ipilimumab treatment in metastatic melanoma

To obtain EMA approval of 'classical' TIL therapy as an ATMP





- **NL:**
  - **John Haanen:** NKI-AVL, Amsterdam, The Netherlands
- **DK:**
  - **Inge Marie Svane:** Herlev Hospital, Copenhagen,
- **UK:**
  - **Robert Hawkins:** University of Manchester and the Christie NHS Foundation Trust, UK

# Phase II trial in refractory MM patients

- Lion Biotechnology



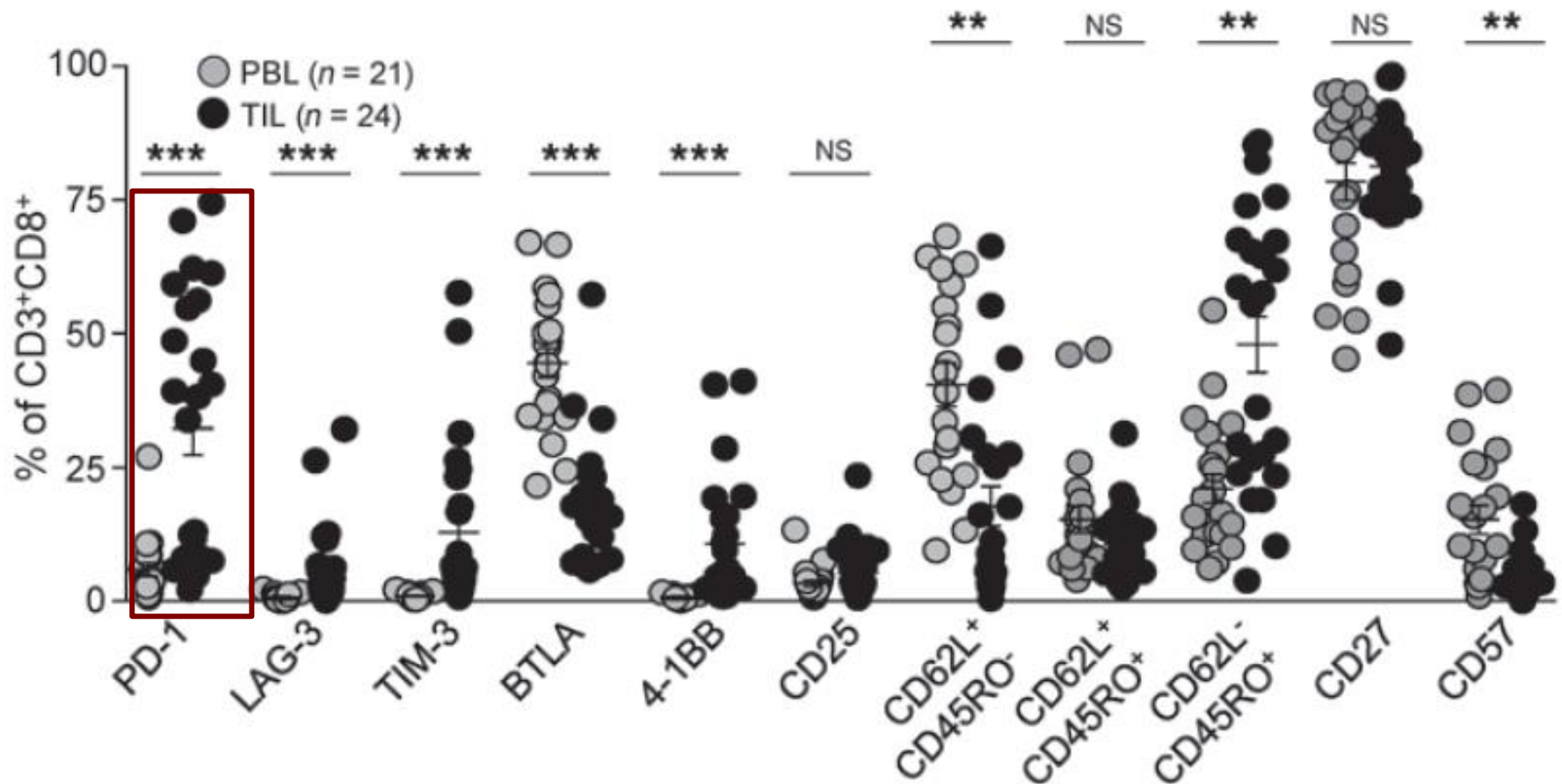
## Pipeline

Sponsor	Indication	Phase 1	Phase 2	Phase 3
NCI/Lion Bio	<a href="#">2nd line metastatic melanoma</a>	 Phase 2 Complete		
NCI/Lion Bio	<a href="#">1st line metastatic melanoma in combination with vemurafenib</a>	 Phase 1 in progress		
Moffitt Cancer Center	<a href="#">1st line metastatic melanoma in combination with ipilimumab</a>	 Pilot trials in progress		
	1st line metastatic melanoma in combination with nivolumab	 Pilot trials in progress		

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  - RCT
  - A large phase II trial in checkpoint inh failing pts
2. Enrichment for tumor-reactive TIL
3. Generate a personalized TIL product
4. Expand TIL therapy beyond melanoma

# PD1 identifies patient-specific tumor-reactive TIL



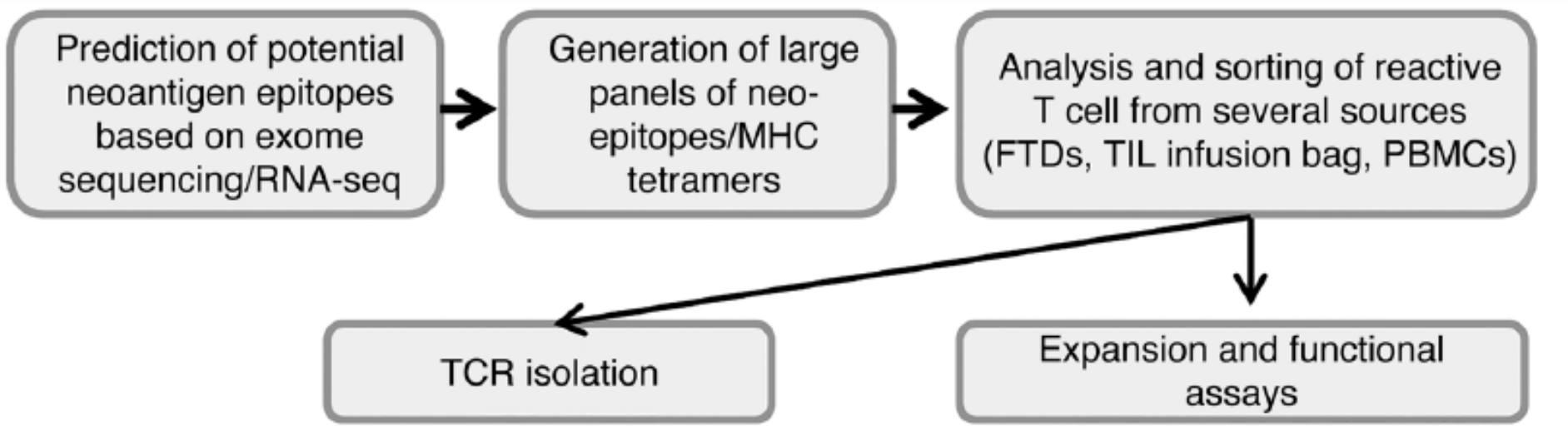
# How to further develop TIL therapy

1. Approval of TIL therapy as treatment option for MM
  - RCT
  - A large phase II trial in checkpoint inh failing pts
2. Enrichment for tumor-reactive TIL
3. Generate a personalized TIL product
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# Isolation of neoantigen-specific T cells from tumor and peripheral lymphocytes

Cyrille J. Cohen,<sup>1,2</sup> Jared J. Gartner,<sup>2</sup> Miryam Horovitz-Fried,<sup>1</sup> Katerina Shamalov,<sup>1</sup> Kasia Trebska-McGowan,<sup>2</sup> Valery V. Bliskovsky,<sup>3</sup> Maria R. Parkhurst,<sup>2</sup> Chen Ankri,<sup>1</sup> Todd D. Prickett,<sup>2</sup> Jessica S. Crystal,<sup>2</sup> Yong F. Li,<sup>2</sup> Mona El-Gamil,<sup>2</sup> Steven A. Rosenberg,<sup>2</sup> and Paul F. Robbins<sup>2</sup>

<sup>1</sup>Laboratory of Tumor Immunology and Immunotherapy, Goodman Faculty of Life Sciences, Bar-Ilan University, Ramat Gan, Israel. <sup>2</sup>Surgery Branch and <sup>3</sup>Laboratory of Cancer Biology and Genetics, National Cancer Institute (NCI), NIH, Bethesda, Maryland, USA.



# How to further develop TIL therapy

1. Approval of TIL therapy as treatment option for MM
  - RCT
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2. Enrichment for tumor-reactive TIL
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4. Expand TIL therapy beyond melanoma

# Cancer Immunotherapy Based on Mutation-Specific CD4+ T Cells in a Patient with Epithelial Cancer

Eric Tran,<sup>1</sup> Simon Turcotte,<sup>1\*</sup> Alena Gros,<sup>1</sup> Paul F. Robbins,<sup>1</sup> Yong-Chen Lu,<sup>1</sup> Mark E. Dudley,<sup>1†</sup> John R. Wunderlich,<sup>1</sup> Robert P. Somerville,<sup>1</sup> Katherine Hogan,<sup>1</sup> Christian S. Hinrichs,<sup>1</sup> Maria R. Parkhurst,<sup>1</sup> James C. Yang,<sup>1</sup> Steven A. Rosenberg<sup>1‡</sup>

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JOURNAL OF CLINICAL ONCOLOGY

ORIGINAL REPORT

## Complete Regression of Metastatic Cervical Cancer After Treatment With Human Papillomavirus–Targeted Tumor-Infiltrating T Cells

*Sanja Stevanović, Lindsey M. Draper, Michelle M. Langhan, Tracy E. Campbell, Mei Li Kwong, John R. Wunderlich, Mark E. Dudley, James C. Yang, Richard M. Sherry, Udai S. Kammula, Nicholas P. Restifo, Steven A. Rosenberg, and Christian S. Hinrichs*

## Immunogenicity of somatic mutations in human gastrointestinal cancers

**Eric Tran, Mojgan Ahmadzadeh, Yong-Chen Lu, Alena Gros, Simon Turcotte,\* Paul F. Robbins, Jared J. Gartner, Zhili Zheng, Yong F. Li, Satyajit Ray, John R. Wunderlich, Robert P. Somerville, Steven A. Rosenberg†**

Surgery Branch, National Cancer Institute, National Institutes of Health, Bethesda, MD 20892, USA.

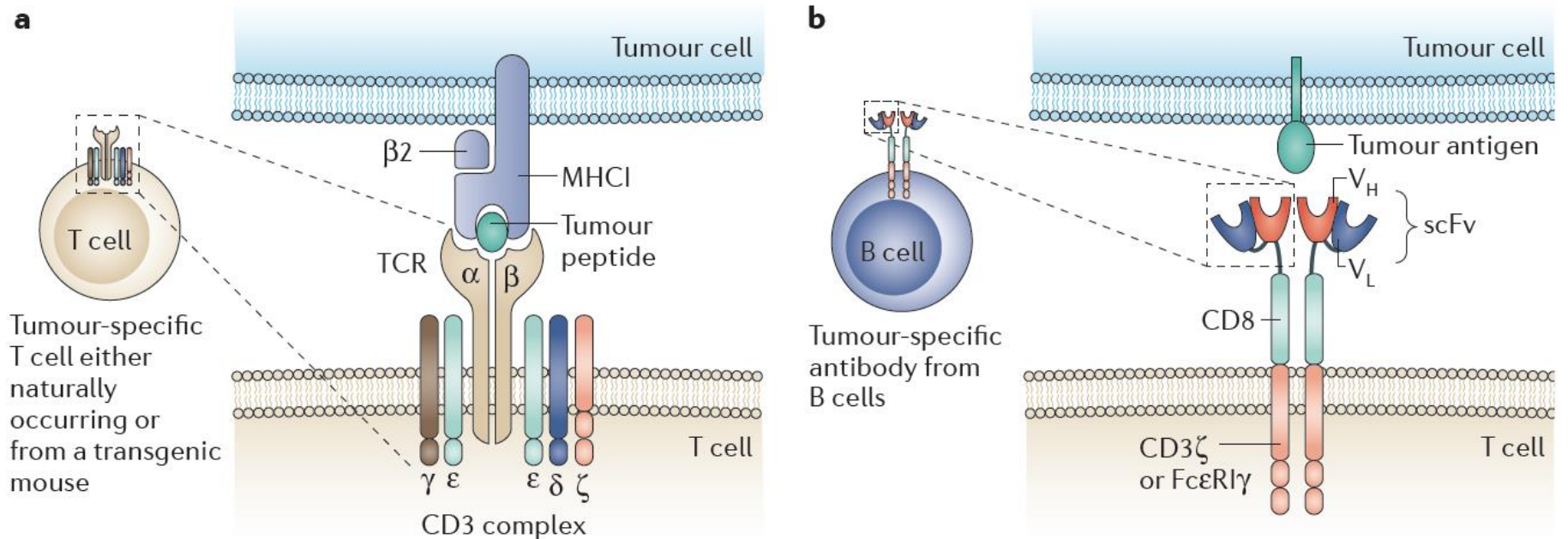
\*Present address: Department of Surgery, Université de Montréal, and Institut du Cancer de Montréal, Centre de Recherche du Centre Hospitalier de l'Université de Montréal, Montréal, QC H2X 0A9, Canada.

# Mutation-specific TIL in GI cancers harboring a low mutational load

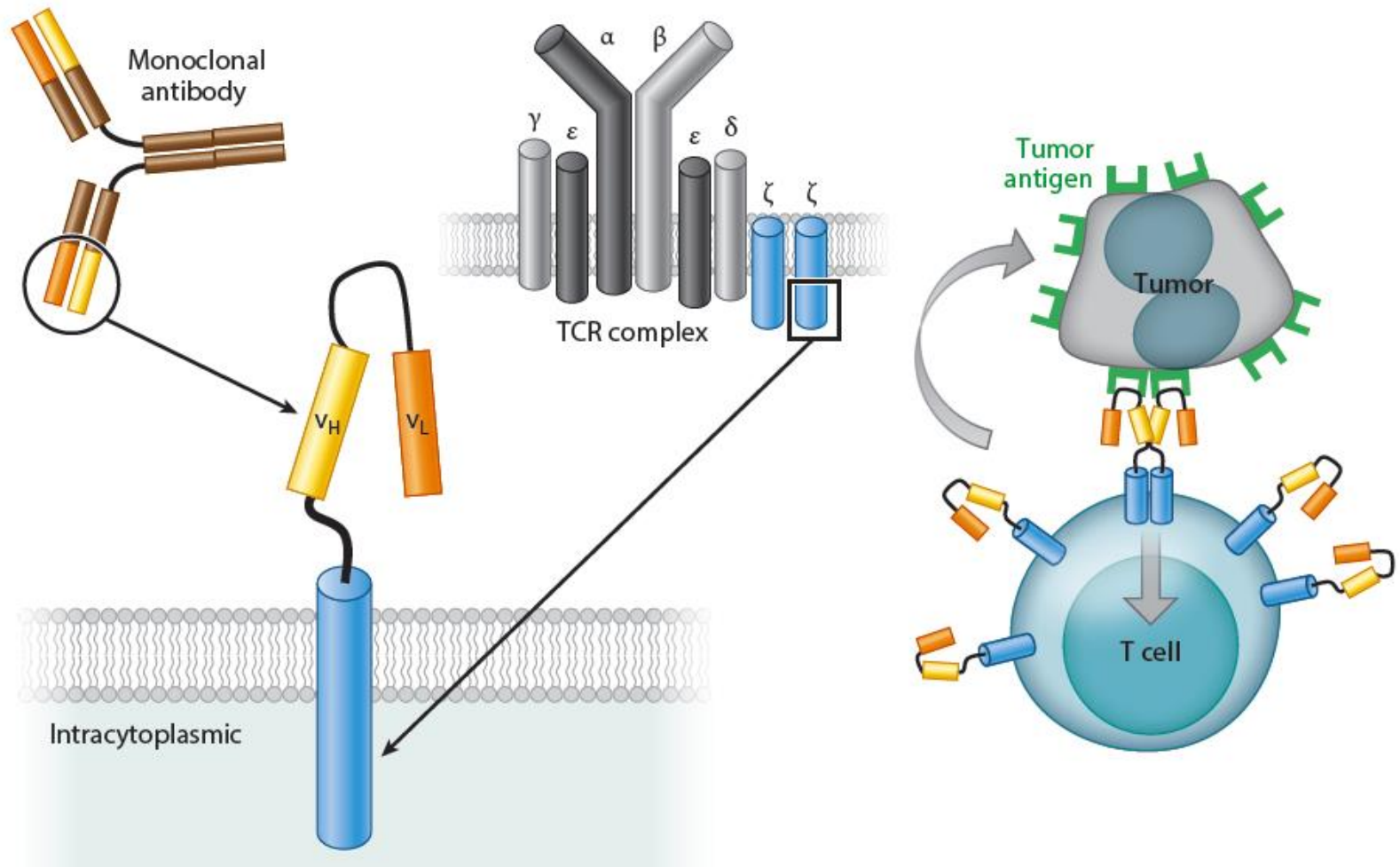
**Table 1. Mutation-reactive T cells in metastatic GI cancers.** NE, not evaluated.

Patient ID	Age/sex	Tumor type	Number of mutations <sup>†</sup>	Number of mutations assessed <sup>‡</sup>	Number of TIL cultures assessed	Number TIL cultures with mutation reactivity <sup>§</sup>	Mutated protein recognized	Amino acid change	T cell type	Frequency of mutation-reactive TCR in tumor (%)
3737*	45/F	Bile duct	26	25	5	5	ERBB2IP	E805G E805G	CD4 CD4	0.009 0.375
3812	44/M	Bile duct	48	179	5	0	—	—	—	—
3942	46/F	Rectal	155	144	6	2 4 3	NUP98 KARS GPD2	A359D D356H E426K	CD8 CD8 CD4	0.67 0.020 0.037
3948	48/M	Esophageal	84	211	5	2 2 2	PLEC XPO7 AKAP2	E1179K P274S Q418K	CD4 CD4 CD4	NE NE NE
3971	49/M	Colon	118	118	23	11	CASP8	F67V	CD8	1.25
3978	46/F	Bile duct	39	38	9	1	ITGB4	S1002I	CD4	NE
3995	50/M	Colon	58	154	19	2 15 2	TUBGCP2 RNF213 KRAS	P293L N1702S G12D	CD8 CD8 CD8	0.023 0.60 0.055
4007	52/M	Colon	134	264	23	4 5	SKIV2L H3F3B	R653H R653H A48T	CD8 CD8 CD8	0.090 0.014 1.19
4032	46/M	Colon	101	222	24	12 1 7	API5 RNF10 PHLPP1	R243Q R243Q E572K G566E	CD8 CD8 CD8 CD8	0.083 0.059 0.030 0.081
4069	57/M	Pancreatic	10	97	15	1	ZFYVE27	R6H	CD8	0.088

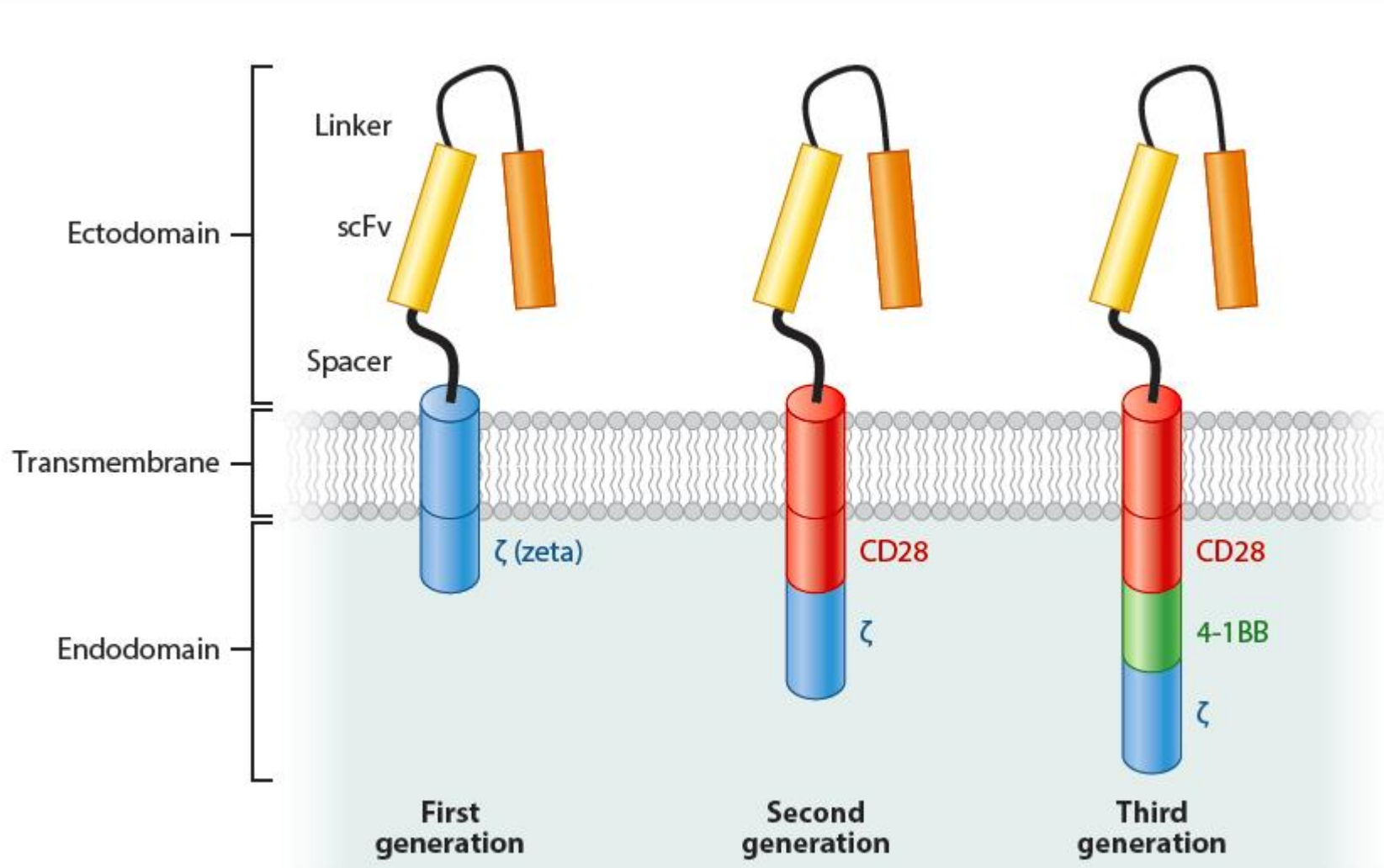
# Infusion of gene-modified T cells



# CAR T cell concept



# Further development of CARs



# Success of CD19 CAR T cell therapy

Study	CAR design	Trial design	Malignancy	Outcome <sup>a</sup>	Refs
Kochenderfer <i>et al.</i> 2010	Murine CD19scFv–CD28/CD3 $\zeta$	Case report	Follicular lymphoma	PR	[3]
Brentjens <i>et al.</i> 2011	Murine CD19 scFv–CD28/CD3 $\zeta$	Pilot (nine patients)	CLL ( $n=8$ ), acute lymphoblastic leukemia ( $n=1$ )	One PR	[4]
Kalos <i>et al.</i> 2011 Porter <i>et al.</i> 2011	Murine CD19 scFv–4-1BB/CD3 $\zeta$	Pilot (three patients)	CLL ( $n=3$ )	Two CR, one PR	[5,6]
Savoldo <i>et al.</i> 2011	Murine CD19 scFv–CD3 $\zeta$ and CD28/CD3 $\zeta$	Pilot (six patients)	Non-Hodgkin lymphoma ( $n=6$ )	Two SD	[7]
Kochenderfer <i>et al.</i> 2012	Murine CD19scFv–CD28/CD3 $\zeta$	Pilot (eight patients)	Non-Hodgkin lymphoma ( $n=4$ ) CLL ( $n=4$ )	One CR, five PR	[8]
Brentjens <i>et al.</i> 2013 Davila <i>et al.</i> 2014	Murine CD19 scFv–CD28/CD3 $\zeta$	Phase I (16 patients)	Acute lymphoblastic leukemia	CR 88%	[9,10]
Grupp <i>et al.</i> 2013	Murine CD19 scFv–4-1BB/CD3 $\zeta$	Pilot (two patients)	Acute lymphoblastic leukemia	Two CR	[11]
Maude <i>et al.</i> 2014	Murine CD19 scFv–4-1BB/CD3 $\zeta$	Phase I/II (30 patients)	Acute lymphoblastic leukemia	CR 90%	[12]
Lee <i>et al.</i> 2015	Murine CD19scFv–CD28/CD3 $\zeta$	Phase I (21 patients)	Acute lymphoblastic leukemia	CR 70%	[13]
Kochenderfer <i>et al.</i> 2015	Murine CD19scFv–CD28/CD3 $\zeta$	Phase I (15 patients)	Non-Hodgkin lymphoma ( $n=11$ ) CLL ( $n=4$ )	CR 53% PR 26%	[14]

## 8. Toxicity management

Anti-cytokine therapy

Steroids

## 7. Safety systems

Suicide systems

Targets for clearance  
by exogenous antibody

Transient expression  
or persistence

## 6. Host conditioning

Lymphodepleting chemotherapy

Lymphodepleting antibodies

Allogeneic transplant

## 5. Ex vivo preparation

Artificial APCs

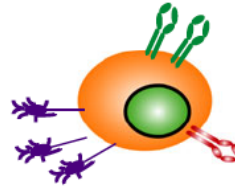
Beads

Culture time

Cell dose and character

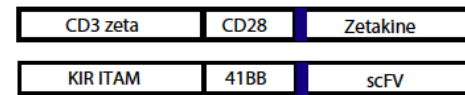
Cytokine support

## 1. Target choice

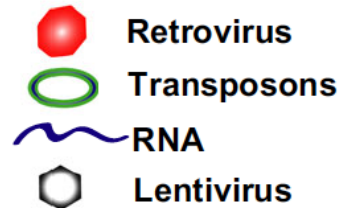


Tissue distribution  
Surface density  
Oncogene addition

## 2. CAR structure



## 3. Vector choice

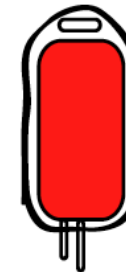


## 4. Effector cells

Third party

Patient derived

Cryopreserved  
Gene edited  
Viral specific



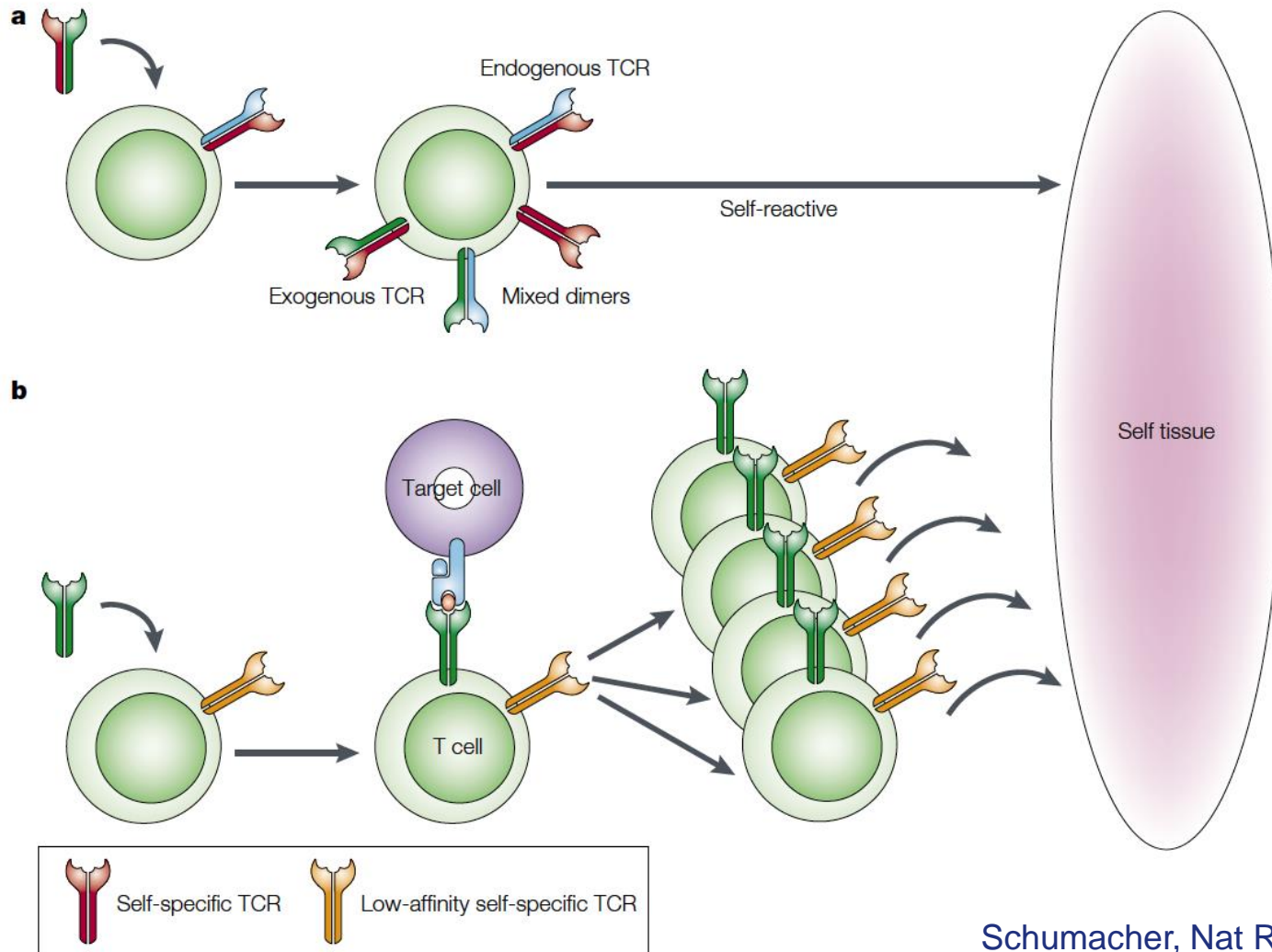
Flow sorting  
Column separation  
Composition  
NK cells  
T cells  
PBMCs



# TCR gene therapy

- 2006: MART-1 TCR gene therapy
  - RR 13% (n=15)  
(Morgan et al., Science 2006)
- 2008: MART-1 and gp100 TCR gene therapy
  - RR 30% (MART-1 TCR; n=20)
  - RR 19% (murine gp100 TCR; n=16)  
(Johnson et al., Blood 2009)
- 2011: NY-eso-1 TCR gene therapy
  - RR 45% (n=11)  
(Robbins et al., J Clin Oncol 2011)
- 2012 MAGE-A3 TCR gene therapy
  - Aborted due to unexpected toxicity

# Potential toxicities of TCR gene therapy (I)



Schumacher, Nat Rev Immunol 2002  
Bendle et al., Nat Med 2010

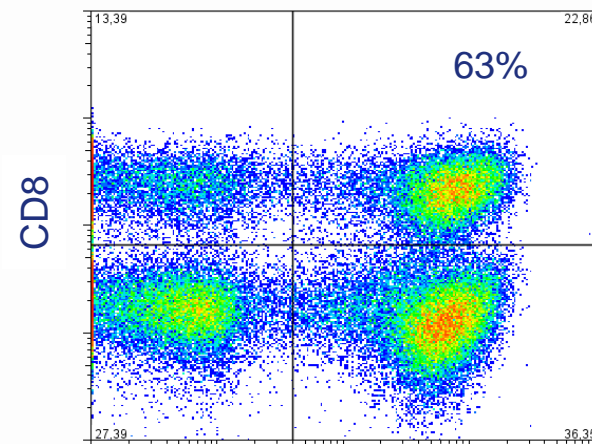
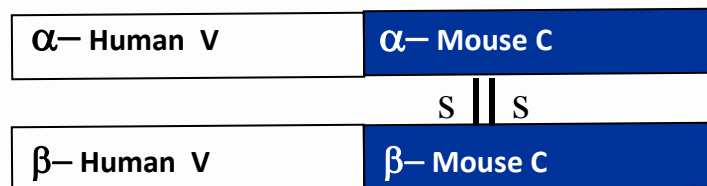
# Selection of TCR and modification of vector

## 1D3<sub>opt</sub>HMCys:

- 1D3 TCR recognizes MART-I epitope (from vaccinated patient)
- codon-optimized for optimal expression
- additional disulfide bond to prevent mispairing with endogenous TCR chains\*
- murine instead of human constant region to further reduce mispairing
- 2A peptide between alpha and beta chain for equimolar production

## Retrovirus:

- MP71 retroviral vector (good expression and used elsewhere in clinical trials)



MART-1 Tm

\* (Kuball Blood 2007, Cohen Cancer Res 2006&2007)

# Potential toxicities of TCR gene therapy

- On and off-target toxicity
  - Melanocyte differentiation antigens
    - Vitiligo; uveitis; Vogt-Koyanagi-Harada syndr.
  - MAGE-A3
    - CNS toxicity (cross-reactivity with MAGE-A9 peptide)
    - Cardiac toxicity (cross-reactivity with myocardiac peptide)
  - CEA
    - colitis

# Take home message

- TIL research and therapy has contributed extensively to our understanding of cancer immunity
- TIL therapy and adoptive therapy of gene modified T cells will be developed further to become a (personalized) drug treatment
- These therapies may be combined with other (non)-immunotherapies in the future
- Use of gene modified T cells:
  - Extremely promising in hemato-oncology
  - Promising in solid tumors (especially TCR modified T cells)



## Cancer exome-guided immunomonitoring

### MHC-based technologies

#### **Chemical Biology**

Boris Rodenko

Huib Ovaa

#### **CCIT, Copenhagen**

Sine Hadrup

#### **STAGE Therapeutics**

Lothar Germeroth

#### **SB, NIH**

Marc Dudley

Steven Rosenberg

#### **ELLA institute, Israel**

Mchal Besser

Jakob Schachter

Nienke van Rooij

Marit van Buuren

Daisy Philips

Mireille Toebes

Laura van Dijk

Pia Kvistborg

**Ton Schumacher**

#### **PDX models**

Kristel Kemper

Daniel Peeper

#### **Sanger Institute**

Sam Behjati

Mike Stratton

#### **Utrecht University**

Can Kesmir



Cancer Immunotherapy  
Dream Team

### Clinical translation

Sander Kelderman

Raquel Gomez

**Joost van den Berg**

Maaïke van Zon

Noor Bakker

Renate de Boer

Bastiaan Nuijen

Christian Blank

Hans van Thienen

Marnix Geukes

Henk Mallo