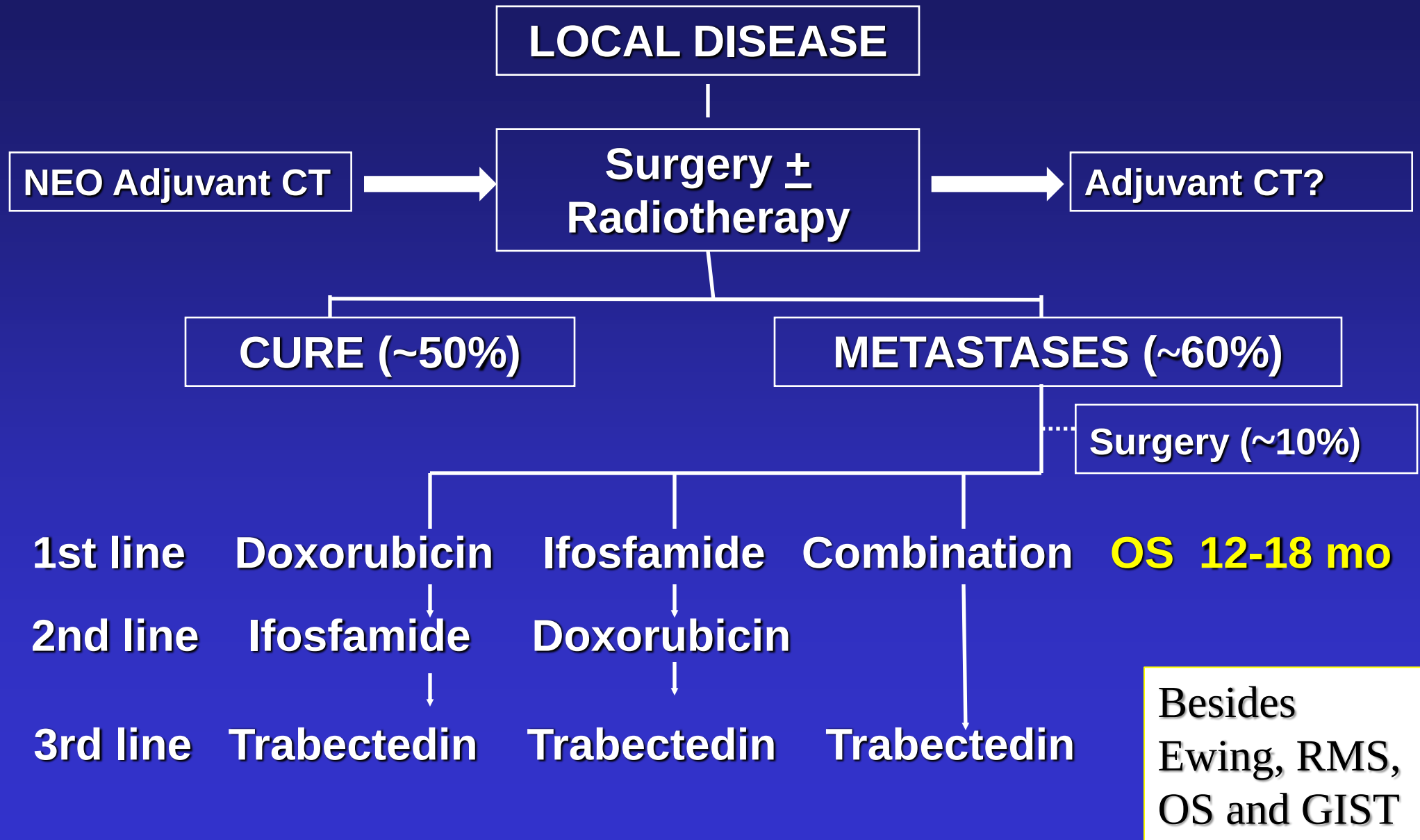


Treating sarcoma subtypes based on molecular characteristics

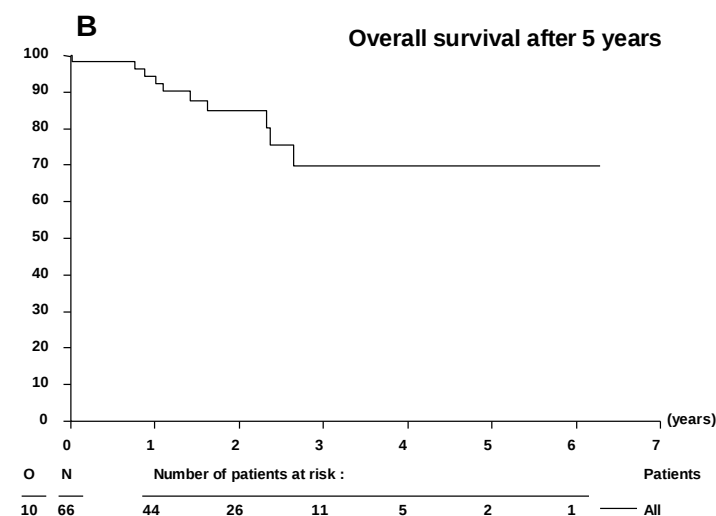
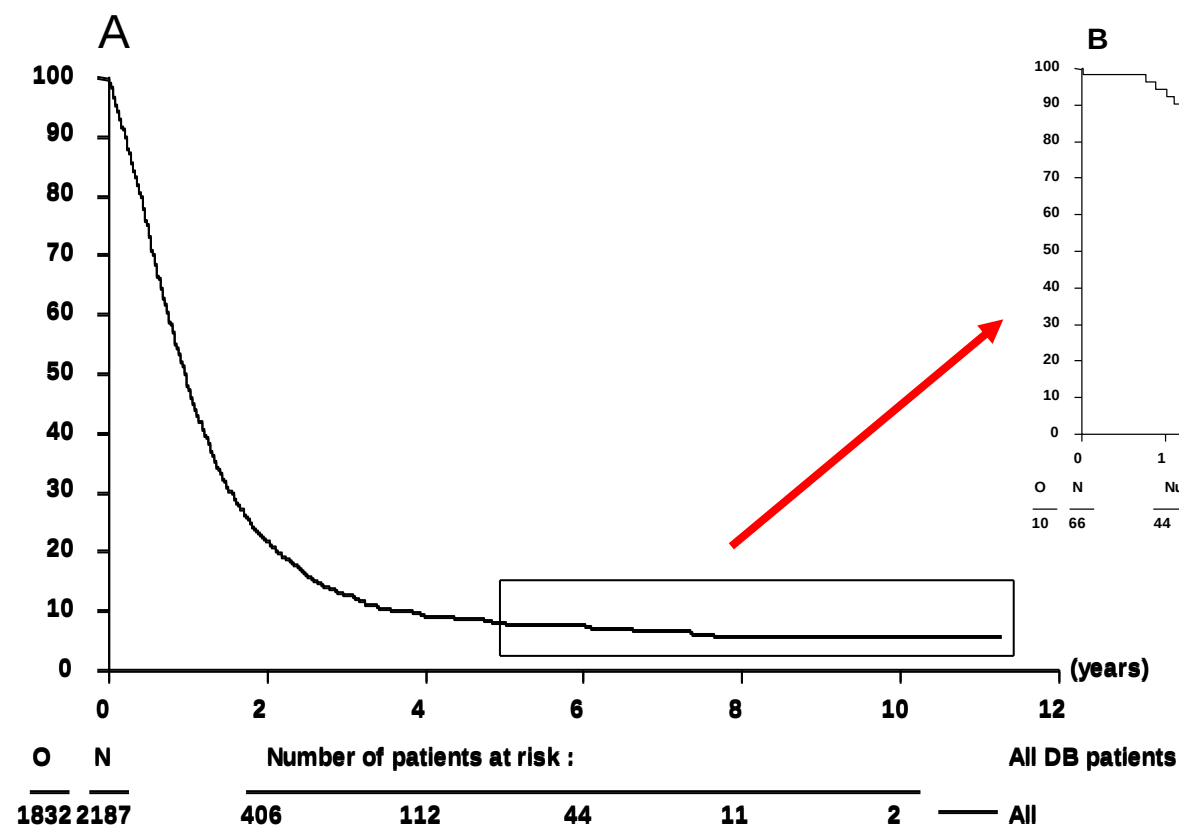
JY Blay
Lyon, France
FSG, EORTC

A simple algorithm?



After local or metastatic relapse

Overall survival



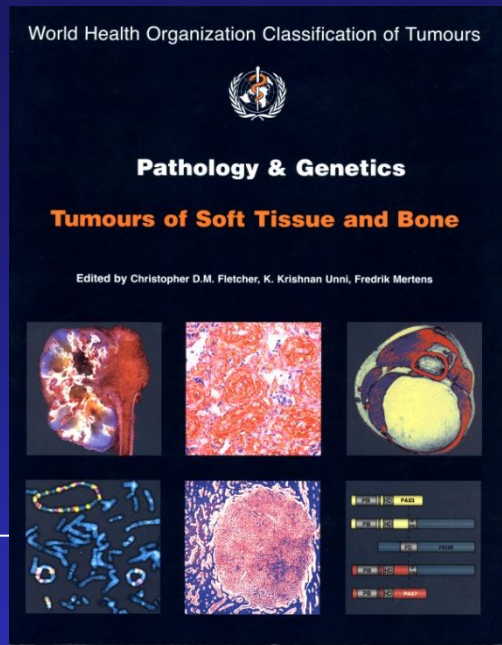
Long term survivors

Response to first line doxorubicin-containing regimens.

Response to 1st line ¶	5-year survivor (n=66) N (%)	Others* (n=1822) N (%)	p	% of 5-year survivors among response subgroup
CR	17 (31%)	64 (4%)		17/81 (21%)
PR	17 (31%)	306 (19%)		17/323 (5%)
SD	17 (31%)	641 (39%)		17/658 (3%)
PD	3 (6%)	627 (38%)	0.00001	3/630 (0.5%)

*: patients with a minimum follow-up of 5 years after inclusion who died within the 5 years of inclusion.

¶: Response was not documented in 196 (10%) patients



Fragmentation

>50 different histotypes
AND molecular subtypes
2013 classification

Adipocytic tumours

Well differentiated / dedifferentiated liposarcoma
Myxoid / round cell liposarcoma
Pleomorphic liposarcoma

.....

Fibroblastic / myofibroblastic tumours

Fibromatosis (desmoid)
Solitary fibrous tumour / haemangiopericytoma
Low grade myofibroblastic tumour
Infantile fibrosarcoma
Adult fibrosarcoma
Mixofibrosarcoma

.....

So-called fibrohistiocytic tumours

Pleomorphic MFH / Undifferentiated pleomorphic sarcoma

.....

Smooth muscle tumours

Leiomyosarcoma

.....

Skeletal muscle tumours

Embryonal rhabdomyosarcoma
Alveolar rhabdomyosarcoma
Pleomorphic rhabdomyosarcoma

Vascular tumours

Epithelioid haemangioendothelioma
Angiosarcoma of soft tissue

.....

Chondro-osseous tumours

Mesenchymal chondrosarcoma
Extraskeletal osteosarcoma

Tumours of uncertain differentiation

Synovial sarcoma
Epithelioid sarcoma
Alveolar soft part sarcoma
Clear cell sarcoma of soft tissue
Extraskeletal myxoid chondrosarcoma
Extraskeletal Ewing tumour
Desmoplastic small round cell tumour
Extra-renal rhabdoid tumour
Malignant mesenchymoma
Neoplasms with perivascular epithelioid cell differentiation

(PEComa)

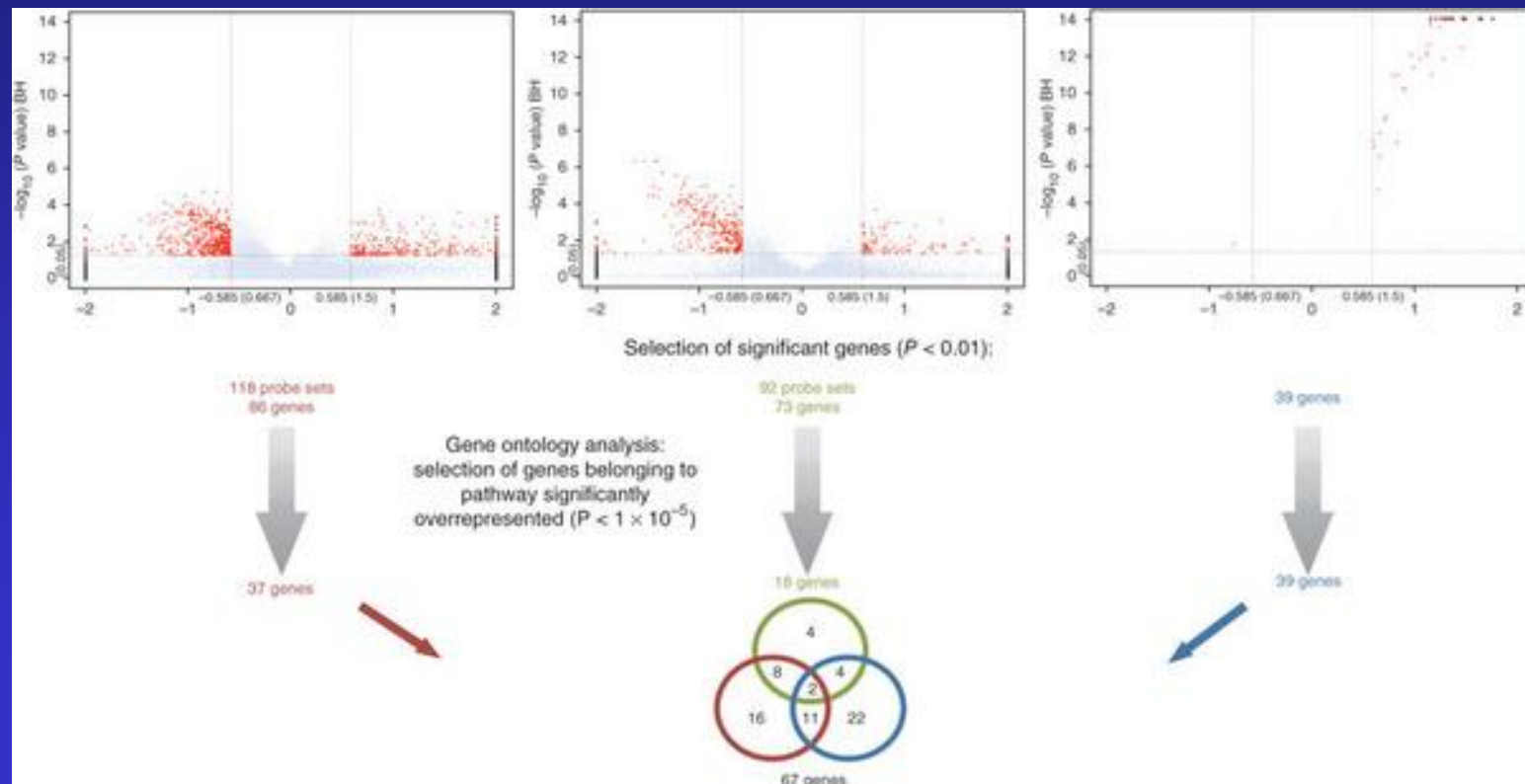
Intimal sarcoma

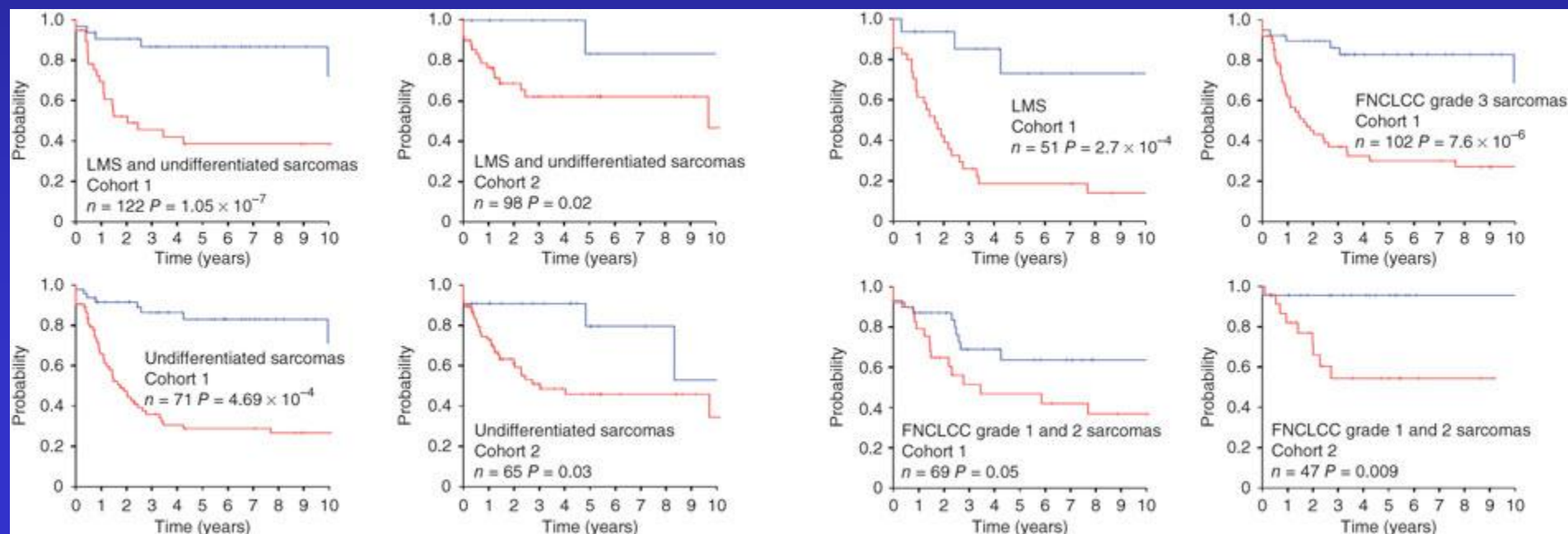
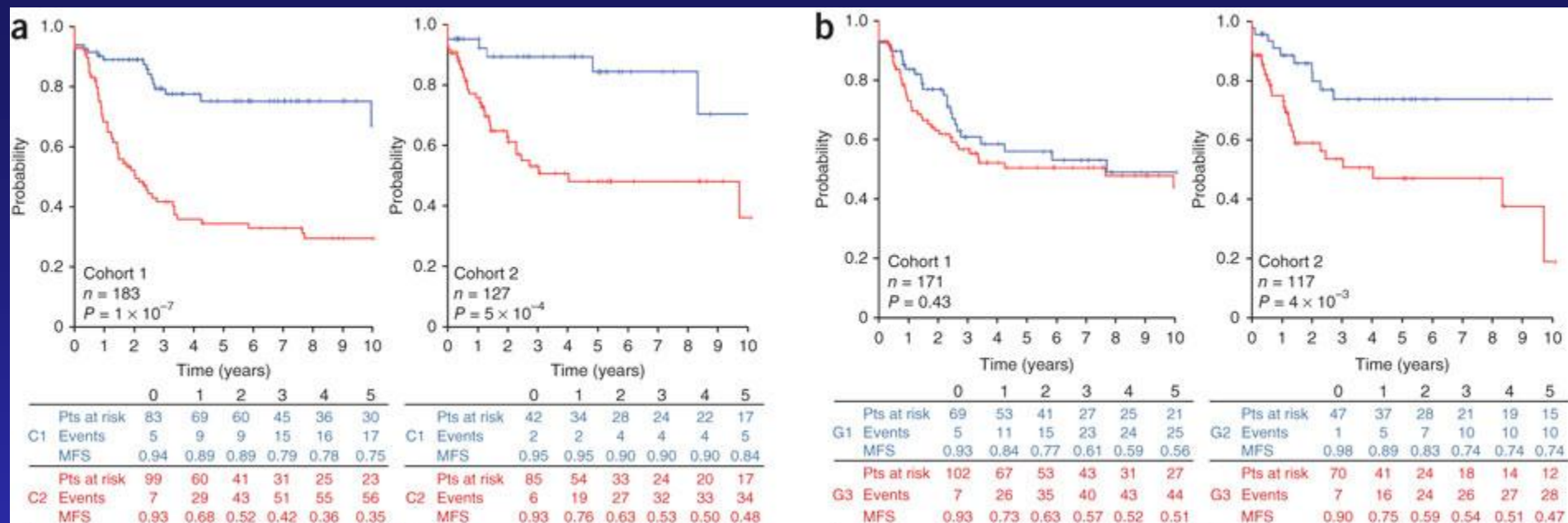
Connective tissue tumours

- Sarcoma with translocations ~15%
 - Ewing, DFSP, Synovial sarcomas,...
- Sarcoma with kinase mutations ~15%
 - GIST, few Angiosarcomas
- Sarcoma with tumor suppressor gene inactivation ~10%
 - MPNST NF1, Rhabdoid tumors- INI1, PEComas TSC...
- Sarcomas with chromosome 12q14-15 amplification ~15%
 - WD/DDLPS, intimal sarcomas, LG OS...
- Sarcomas with complex genetic alterations ~50%
 - Pleomorphic sarcomas, LMS, ...
- Low grade or locally aggressive
 - Desmoid tumors beta catenin or APC mutation
 - Giant cell tumor of the bone ? (RANK involved)
 - Giant cell tumor of the soft part (PVNS) translocation

Sarcomas with complex genomics

- Molecular signature for sarcoma
- Beyond FNCLCC grading?





Systemic treatment of sarcomas 2000-2012 ESMO 2012

2000:

- All sarcomas
 - Doxorubicin
 - Ifosfamide
 - DTIC
- Ewing , RMS, OS
 - Dactinomycin
 - CDDP
 - Vincaalcaloids
 - Cyclophosphamide
 - HDMTX

2012

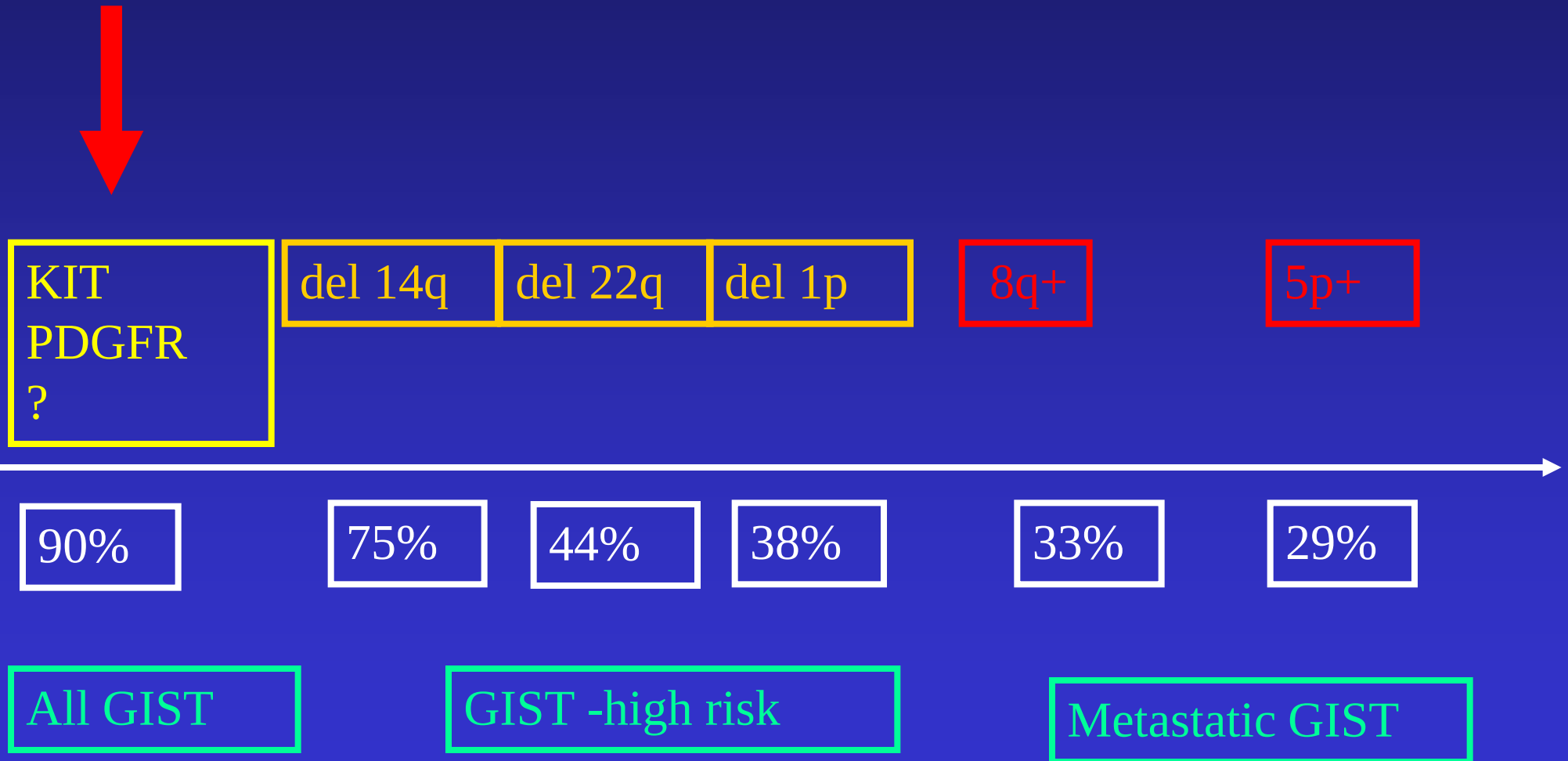
- All subtypes : Trabectedin
- GIST: Imatinib, sunitinib,
- DFSP : Imatinib
- Osteosarcomas: MTPPE
- LMS: Gemcitabine (+/- DTIC or Tax)
- EWS: TopoI inh., IGF1R
- A/E RMS: Topo inh
- Angio: Dox, Paclitaxel, GemTax
- ESS : Aromatase inh.
- All but LPS: pazopanib
- PVNS :Imatinib
- Desmoid Tumors : HT, imatinib
- GCTB : denosumab
- PECOMAs: mTOR
- WD/DD: LPS:CDK4, MDM2
- GIST: regorafenib
- ASPS: cediranib



Three situations

- Initial molecular event
 - KIT in GIST
 - Loss NF1, TSC
 - Translocations
 - Mdm2 amplification
 - ...
- Secondary event
 - VEGF production
 - Activation of mTOR pathway
 - ER expression in ESS
- Simple bystander
 - PDGFR expression in normal (and malignant) cells of connective tissue

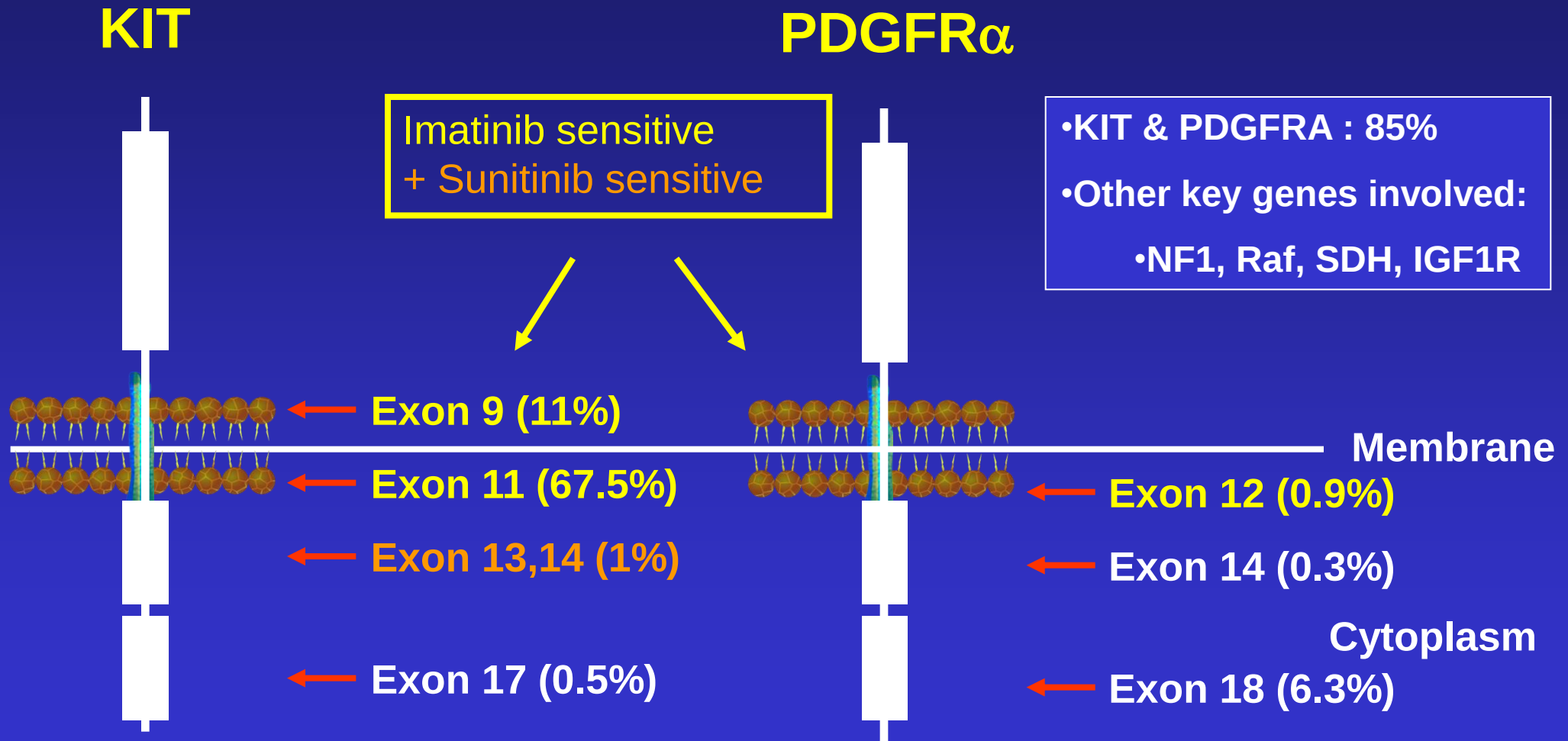
1.Targeting an initial event



Connective tissue tumours

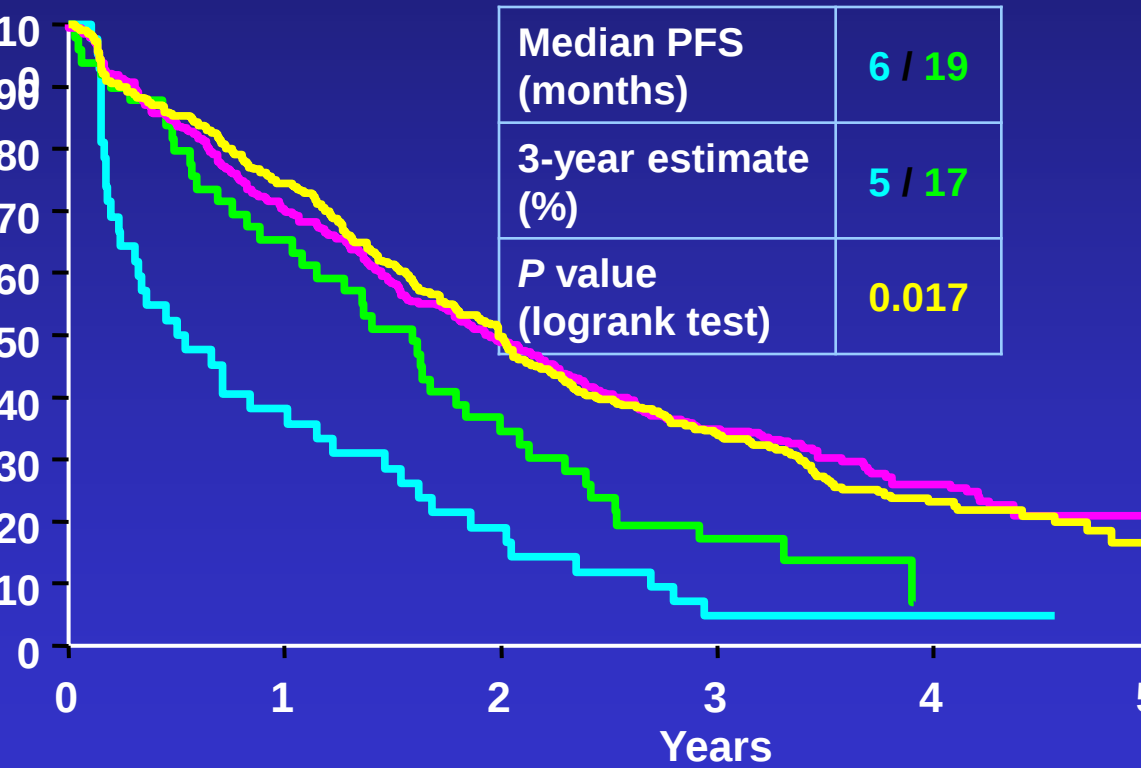
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KIT and PDGFR α mutations in GIST



GIST are at least 10 diseases

KIT exon 9 mutants (10% of patients)



KIT exon 9 mutants: 400 mg / 800 mg
Other patients: 400 mg / 800 mg

	Dose	Adjuvant
KIT Exon 11	Im 400	+
KIT exon 9	Im 800	+
PDGFRA		
Non D842V	Im 400	+
D842V:	0	0
KIT/PDGFR WT	Im 400	+/?
NF1	?/Im 400	+/?
SDHB	?/Im 400	+/?
Raf	?	?
Pediatric	?	?

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DFSP and imatinib

DFSP /giant cell fibroblastoma

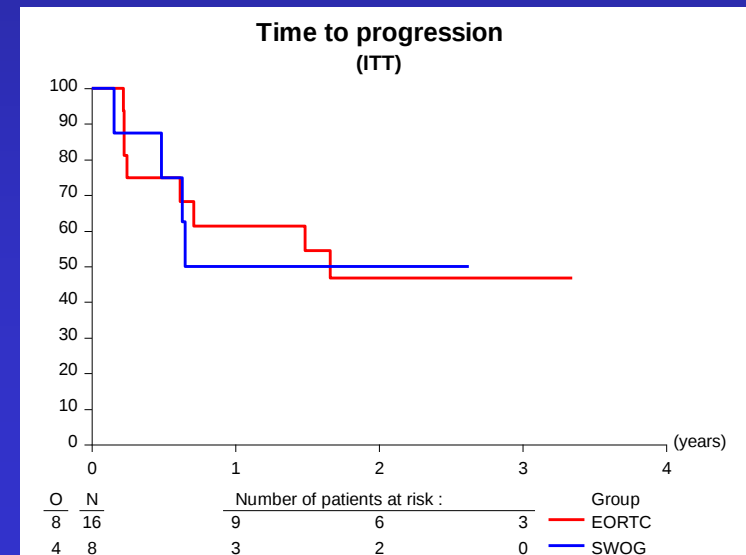
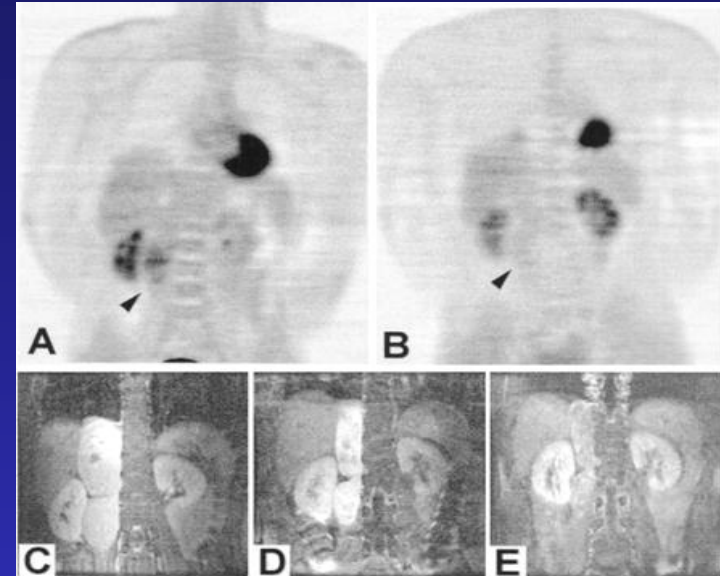
• **t(17,22)** :17q22 and 22q13
(*COL1A1* et *PDGFB*)

• Autocrine loop with PDGF β

Maki et al 2002

Mc Arthur et al 2005

Rutkowski et al 2010





FONDAZIONE IRCCS
ISTITUTO NAZIONALE
DEI TUMORI



LEIDEN UNIVERSITY MEDICAL CENTER

ESMO 2012

Imatinib mesylate (IM) for the treatment of locally advanced and/or metastatic pigmented villonodular synovitis/tenosynovial giant cell tumor (PVNS/TGCT).

Philippe A. Cassier, Silvia Stacchiotti, Hans Gelderblom, David Thomas, Winette van der Graaf, Beatrice Seddon, Julien Domont, Andrew J. Wagner, Jean-Yves Blay.

Peter Mac

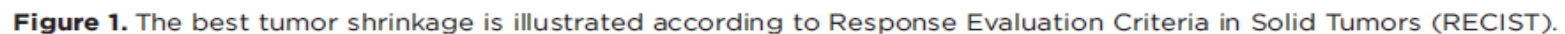
UMC  St Radboud

University College London Hospitals 
NHS Foundation Trust

 Institut de cancérologie
GUSTAVE ROUSSY
VILLEJUIF - www.igr.fr

 DANA-FARBER
CANCER INSTITUTE

Philippe A. Cassier, MD¹; Hans Gelderblom, MD²; Silvia Stacchiotti, MD³; David Thomas, MD⁴; Robert G. Maki, MD⁵; Judith R. Kroep, MD²; Winette T. van der Graaf, MD⁶; Antoine Italiano, MD⁷; Beatrice Seddon, MD⁸; Julien Dômont, MD⁹; Emanuelle Bompas, MD¹⁰; Andrew J. Wagner, MD¹¹; and Jean-Yves Blay, MD^{1,12}



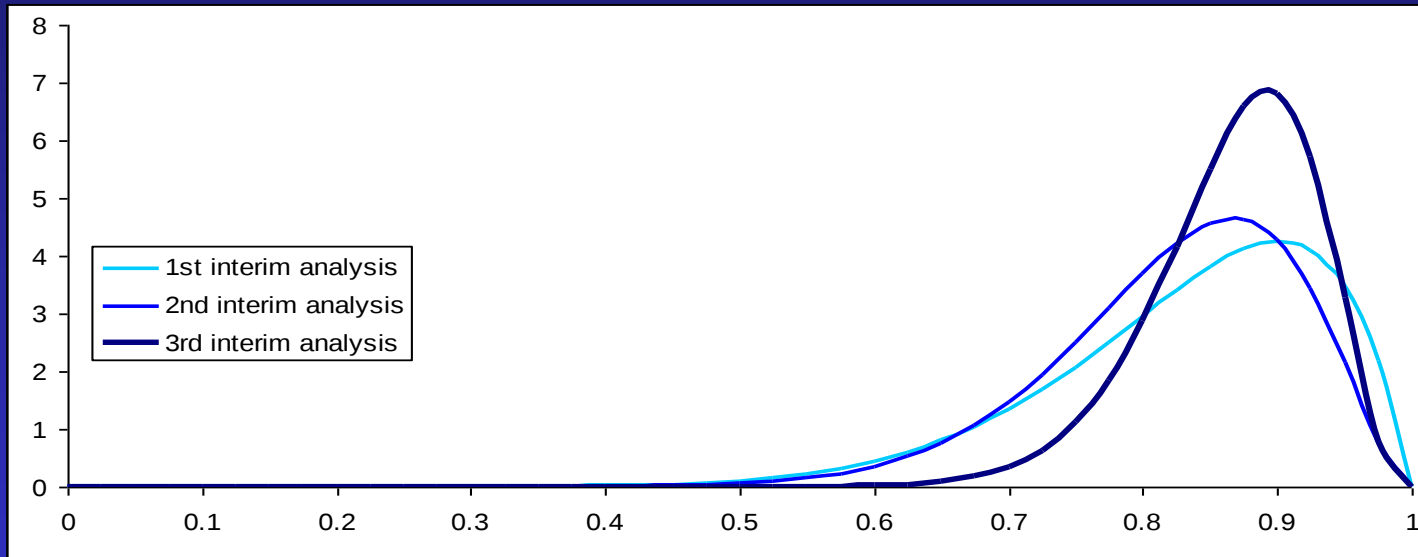
PVNS study: An open-label international multicentric phase II study of nilotinib in progressive pigmented villonodular synovitis not amenable to a conservative surgical treatment

Isabelle RAY-COQUARD, Hans GELDERBLOM, Christine CHEVREAU, Judith R. KROEP, Antoine ITALIANO, Silvia STACCHIOTTI, Axel LE CESNE, Sophie PIPERNO-NEUMANN, Virginia FERRARESI, Florence DUFFAUD, Nicolas PENEL, Philippe A. CASSIER, Martin TATTERSALL, Andrew Bassim HASSAN, Binh BUI NGUYEN, Valérie BOURNE-BRANCHU, Séverine GUILLEMAUT, Claire CROPET, David PEROL, Jean-Yves BLAY

for the World Sarcoma Network

Phase II nilotinib in PVNS

- Non progression rate at 12 weeks: **88.9% (95% CI: 70.8-97.6)**
- Updated distributions of the NPR (Bayesian estimation)



➔ Based on the updated distribution (3rd interim analysis):
 $\text{Pr.}(\text{NPR} \leq 30\%) = 3.34 \times 10^{-11}$: **the study has not to be stopped at this stage**

Crizotinib in *ALK*-Rearranged Inflammatory Myofibroblastic Tumor

James E. Butrynski, M.D., David R. D'Adamo, M.D., Ph.D.,
Jason L. Hornick, M.D., Ph.D., Paola Dal Cin, Ph.D., Cristina R. Antonescu, M.D.,
Suresh C. Jhanwar, Ph.D., Marc Ladanyi, M.D., Marzia Capelletti, Ph.D.,
Scott J. Rodig, M.D., Ph.D., Nikhil Ramaiya, M.D., Eunice L. Kwak, M.D.,
Jeffrey W. Clark, M.D., Keith D. Wilner, Ph.D., James G. Christensen, Ph.D.,
Pasi A. Jänne, M.D., Ph.D., Robert G. Maki, M.D., Ph.D.,
George D. Demetri, M.D., and Geoffrey I. Shapiro, M.D., Ph.D.

SUMMARY

Inflammatory myofibroblastic tumor (IMT) is a distinctive mesenchymal neoplasm characterized by a spindle-cell proliferation with an inflammatory infiltrate. Approximately half of IMTs carry rearrangements of the anaplastic lymphoma kinase (*ALK*) locus on chromosome 2p23, causing aberrant *ALK* expression. We report a sustained partial response to the *ALK* inhibitor crizotinib (PF-02341066, Pfizer) in a patient with *ALK*-translocated IMT, as compared with no observed activity in another patient without the *ALK* translocation. These results support the dependence of *ALK*-rearranged tumors on *ALK*-mediated signaling and suggest a therapeutic strategy for genomically identified patients with the aggressive form of this soft-tissue tumor. (Funded by Pfizer and others; ClinicalTrials.gov number, NCT00585195.)

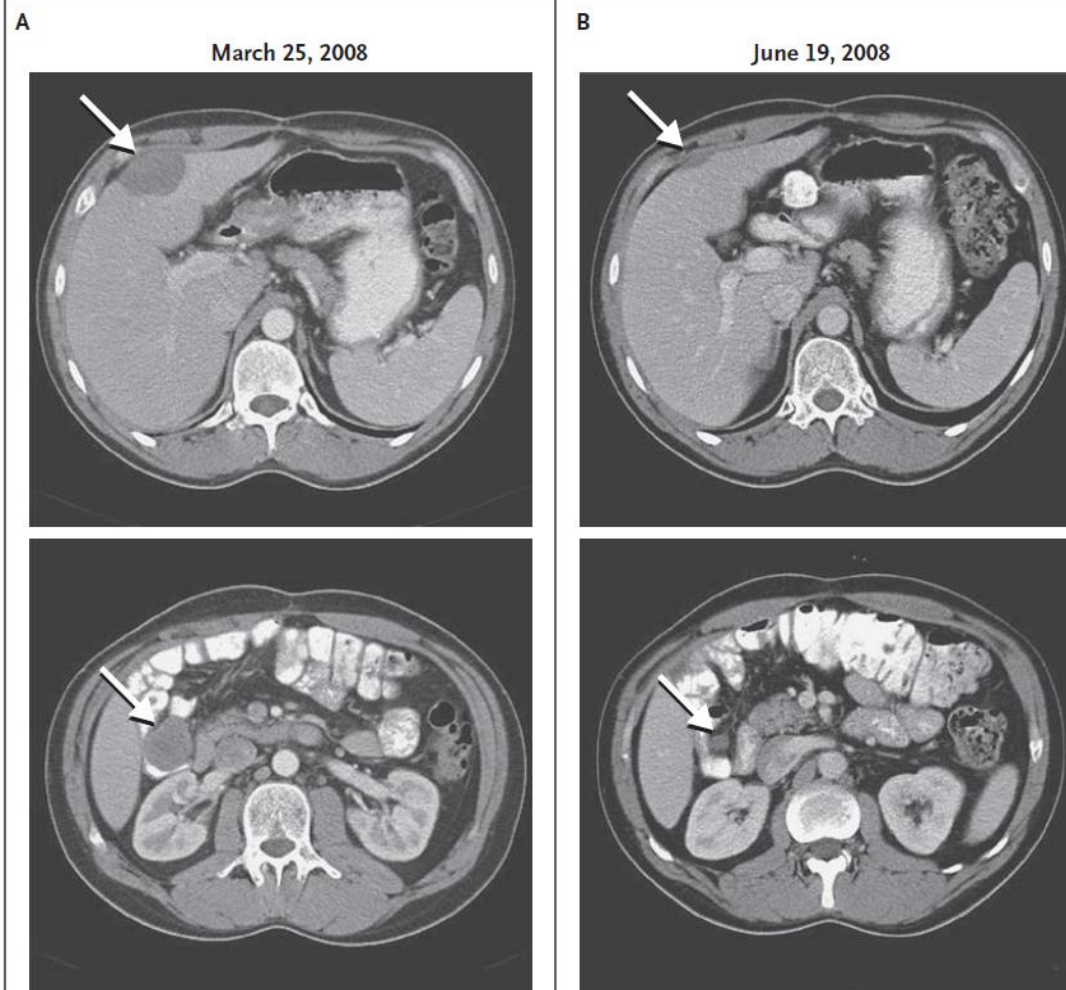
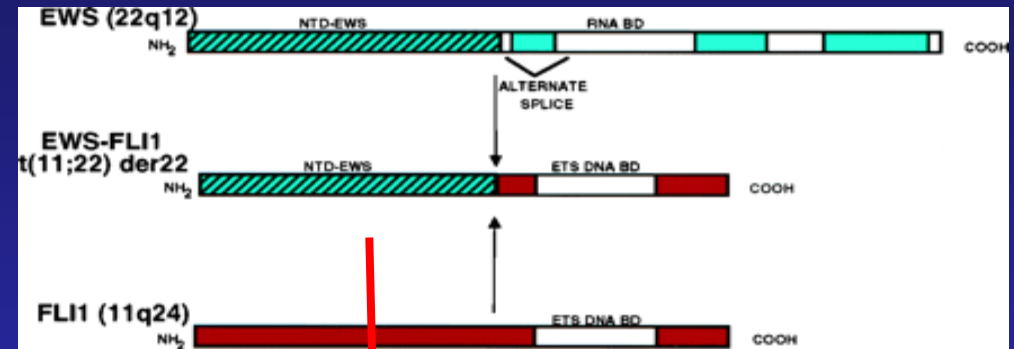
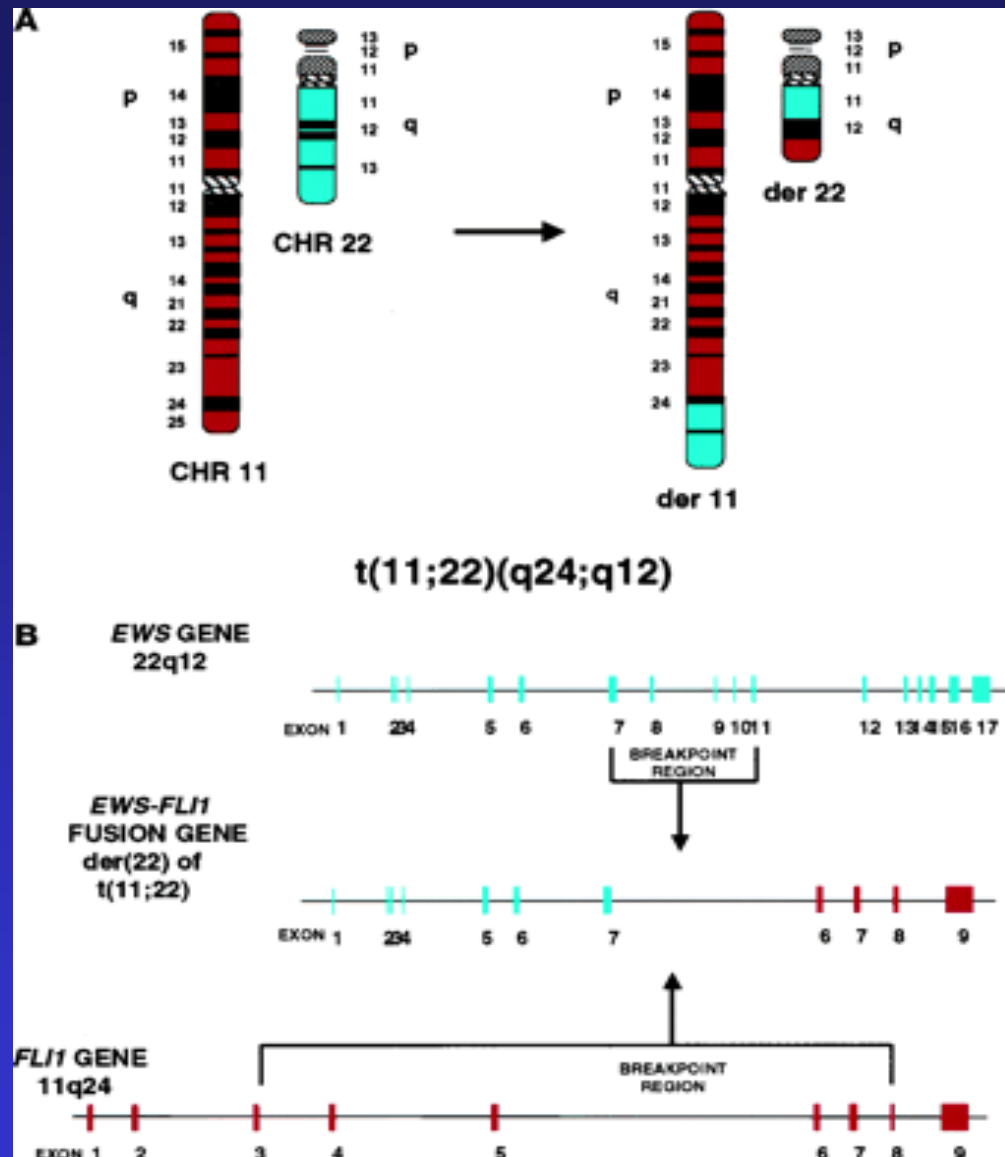


Figure 2. CT Scans Showing the Response to Crizotinib in Patient 1.

The baseline abdominal CT scan shows a hepatic mass measuring 4.8 by 3.3 cm (top) and one of several mesenteric masses measuring 3.8 by 3.3 cm (bottom) (Panel A, arrows). After 13 weeks of treatment with crizotinib, the hepatic and mesenteric masses measured 2.3 by 0.8 cm and 1.3 by 1.2 cm, respectively (Panel B, arrows). In October 2008, these masses measured 3.6 by 2.2 cm and 0.5 by 0.5 cm, respectively (not shown), indicating that the hepatic mass had regrown, despite a continued response, according to the Response Evaluation Criteria in Solid Tumors.

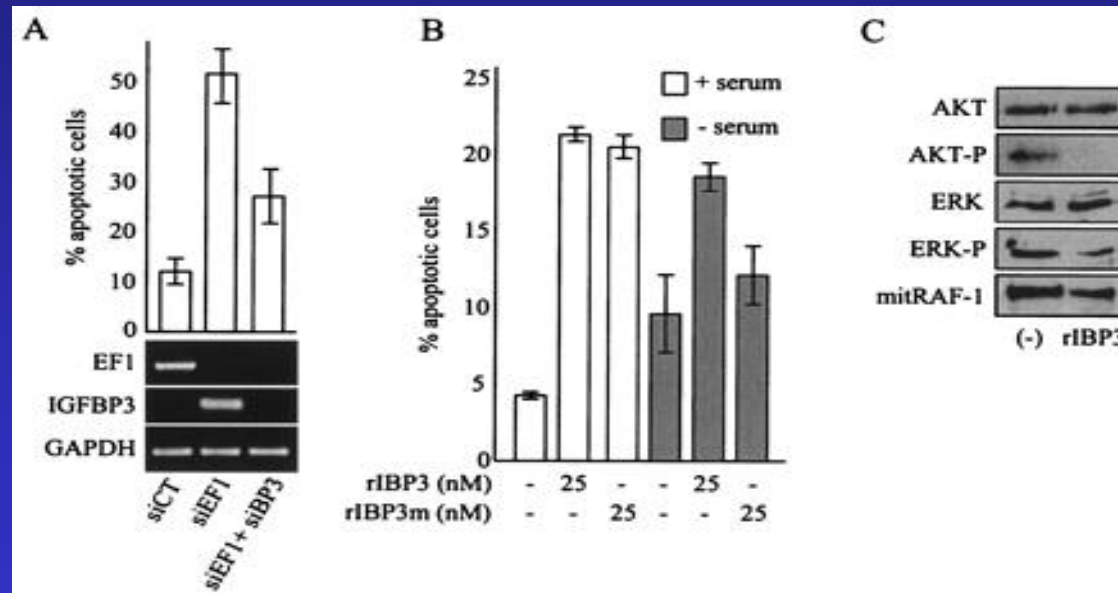


Abnormal gene transcription

Transformation

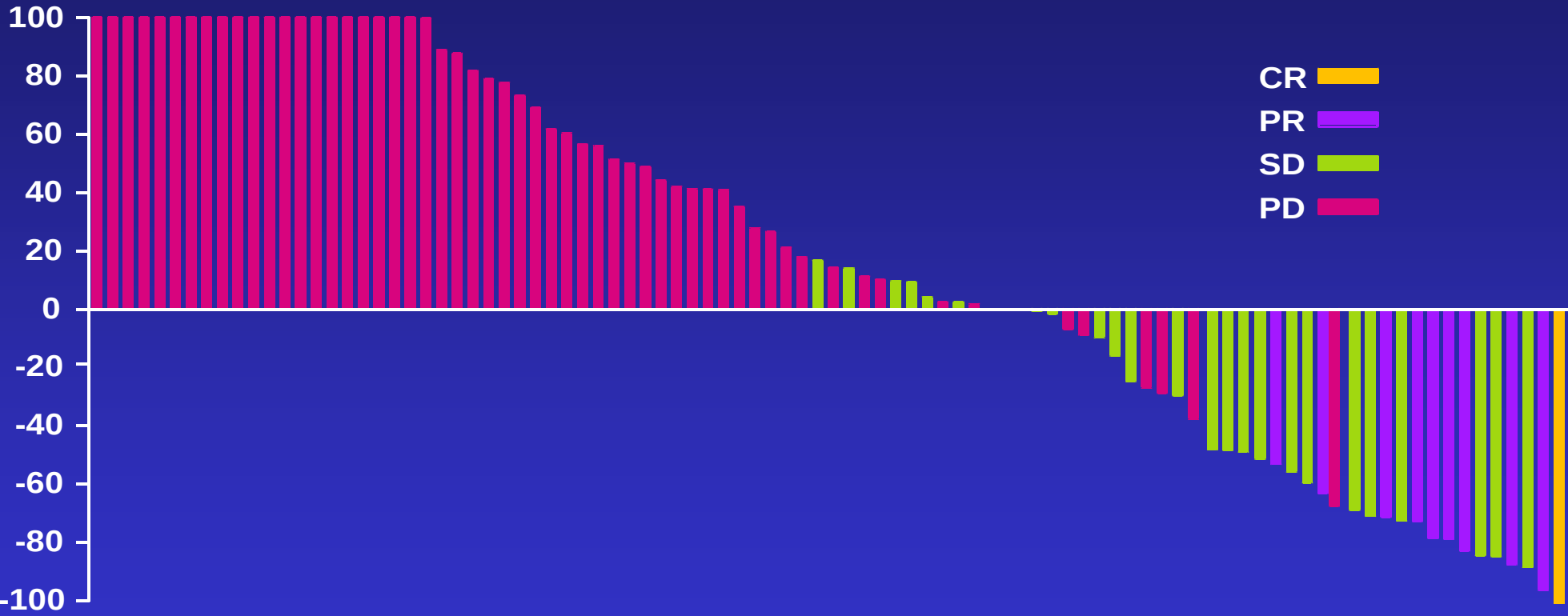
Target genes: include kinases and ligands (PDGF, IGFBP3...)

IGFBP3 induces Ewing tumor cells apoptosis

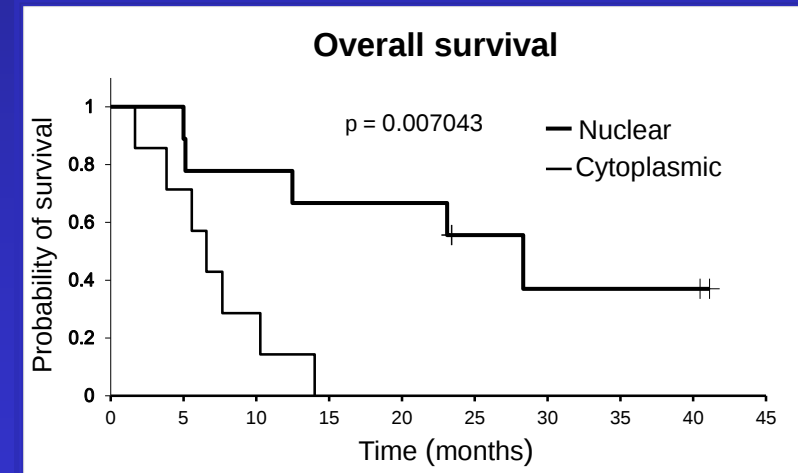
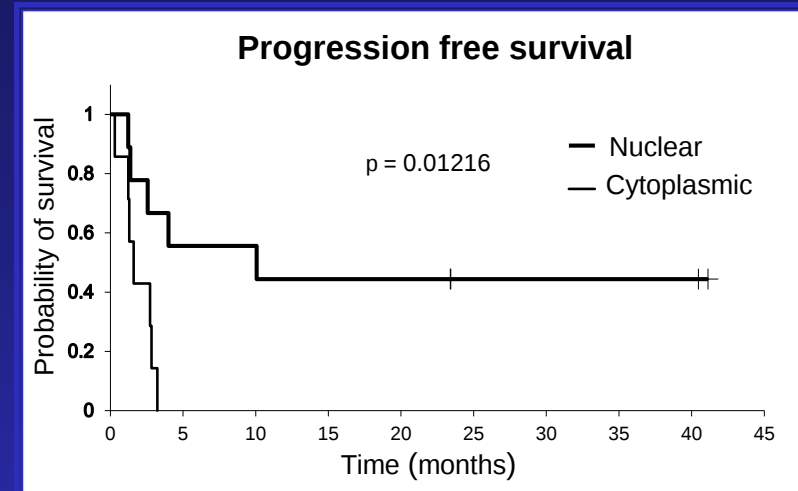
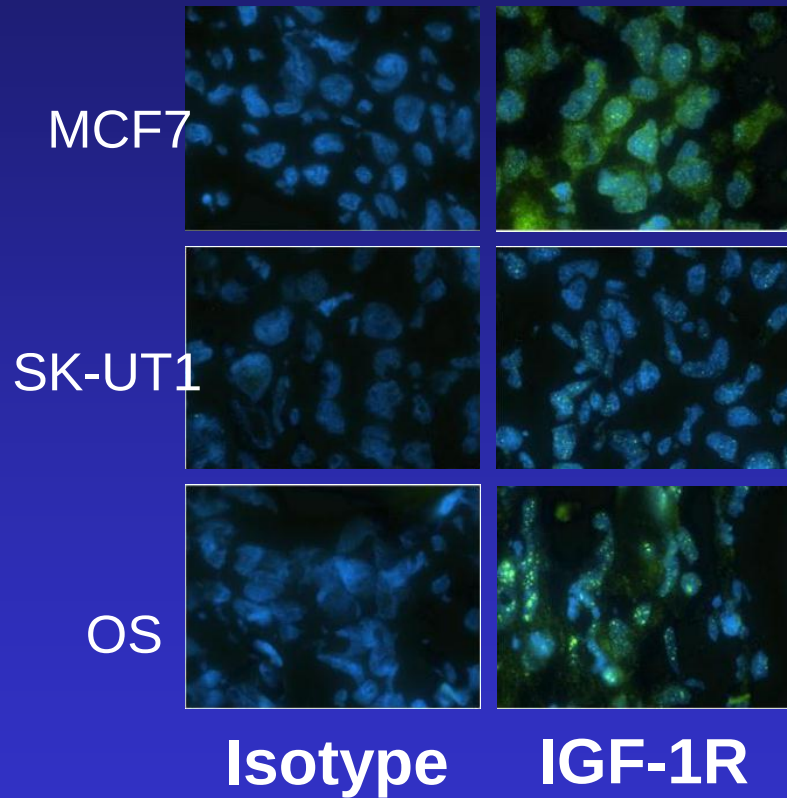


IGF1 inhibitors as potential targeted therapy in ES ?

Best Recorded Variation in Tumor Size: Ewing's Cohorts



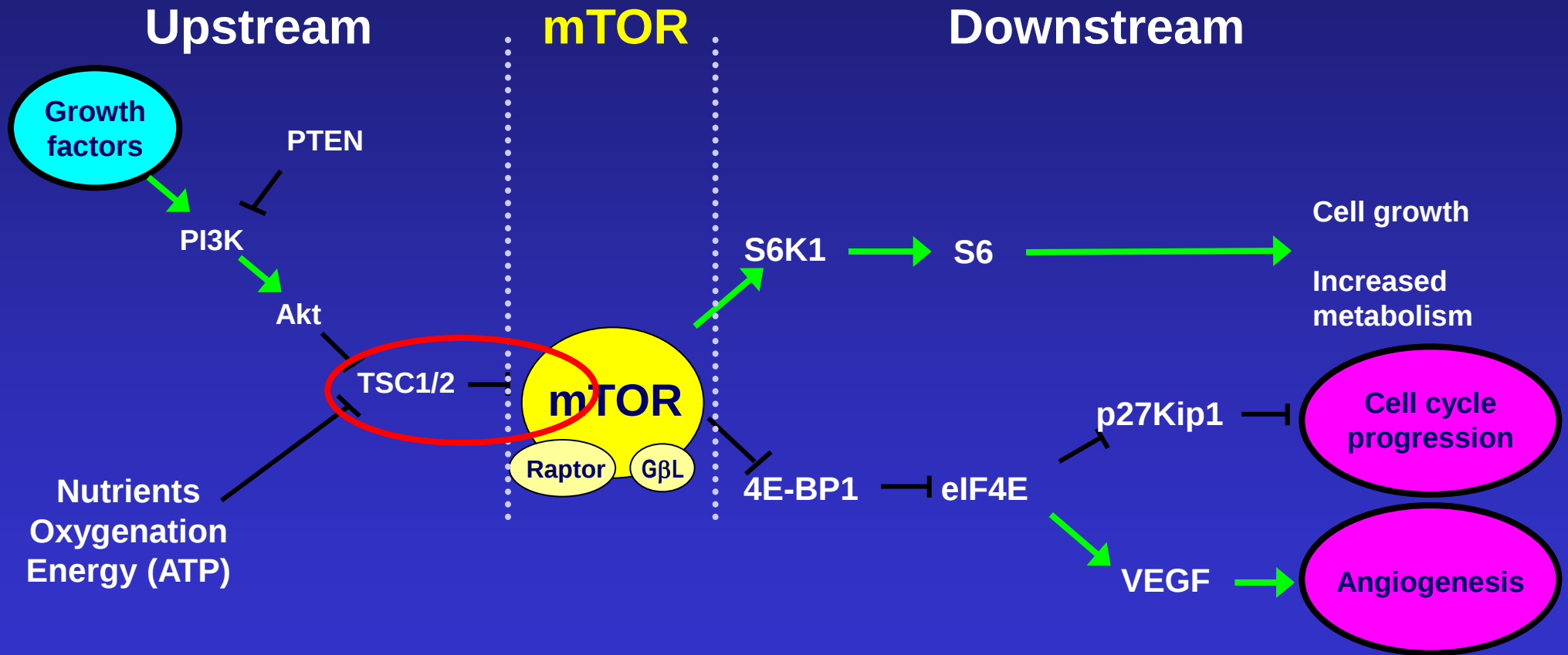
Nuclear staining for IGF1R: a biomarker for response in sarcoma?



Connective tissue tumours

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 - Giant cell tumor of the soft part (PVNS) translocation

mTOR signalling pathway



ORIGINAL ARTICLE

Sirolimus for Angiomyolipoma in Tuberous Sclerosis Complex or Lymphangioleiomyomatosis

John J. Bissler, M.D., Francis X. McCormack, M.D., Lisa R. Young, M.D.,
Jean M. Elwing, M.D., Gail Chuck, L.M.T., Jennifer M. Leonard, R.N.,
Vincent J. Schmithorst, Ph.D., Tal Laor, M.D., Alan S. Brody, M.D.,
Judy Bean, Ph.D., Shelia Salisbury, M.S., and David N. Franz, M.D.

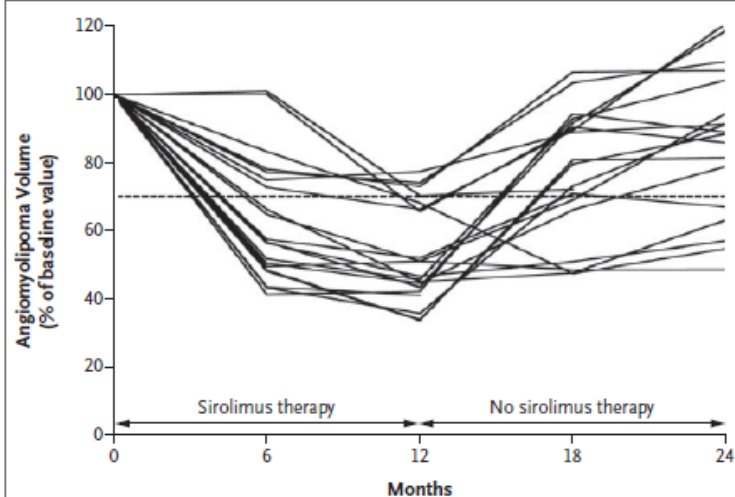


Figure 2. Angiomyolipoma Volume in the Patients with the Tuberous Sclerosis Complex or Sporadic Lymphangioleiomyomatosis during the Study.

Angiomyolipomas were visualized with the use of abdominal magnetic resonance imaging, and volumetric analysis was performed at baseline and at 2, 4, 6, 12, 18, and 24 months. The angiomyolipoma volume at each visit is expressed as a percentage of the baseline size. The dashed line represents 70% of the baseline value; data below the line indicate that the mean angiomyolipoma volume was reduced by 30% or more.

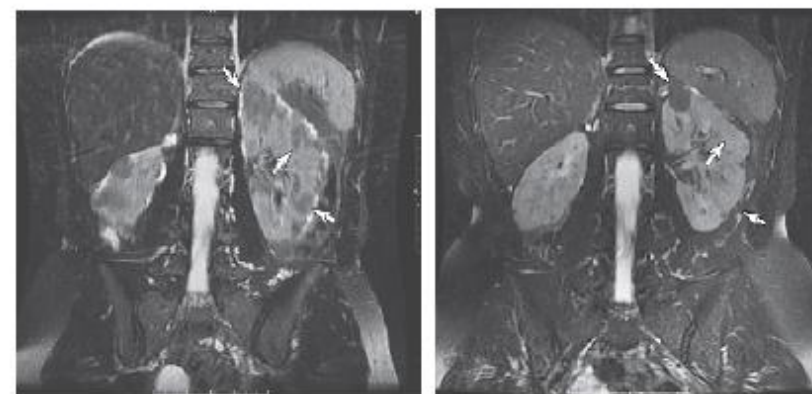


Figure 3. Renal Angiomyolipomas in the Abdomen of a Patient with the Tuberous Sclerosis Complex.

Bilateral angiomyolipomas are shown at baseline and after 12 months of sirolimus therapy. Three lesions in the left kidney are identified by arrows; at 12 months, the top lesion had become reduced in size and the bottom two had become imperceptible. The images were obtained with the use of fast spin-echo T₂-weighted magnetic resonance imaging with fat suppression.

Connective tissue tumours

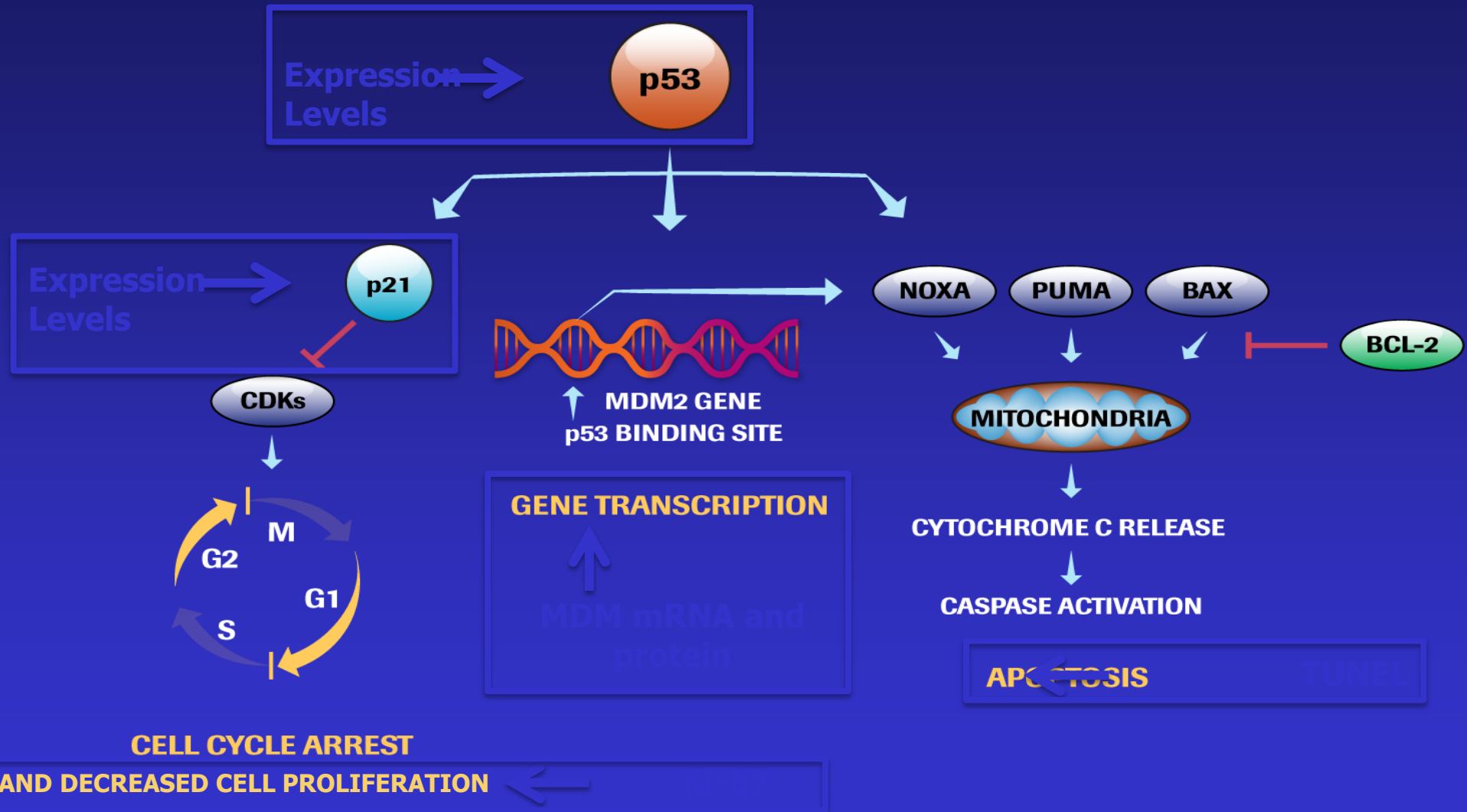
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Neoadjuvant MDM2 Antagonist RG7112 for Well-Differentiated and Dedifferentiated Liposarcomas (WD/DD LPS): A Pharmacodynamic (PD) Biomarker Study. (Abstract 10007b)

I. Ray-Coquard¹, J. Y. Blay¹, A. Italiano², A. Le Cesne³, N. Penel⁴, J. Zhi⁵, A. Beryozkina⁵, F. Heil⁶, R. Rueger⁶, G. L. Nichols⁵, B. Bui Nguyen²

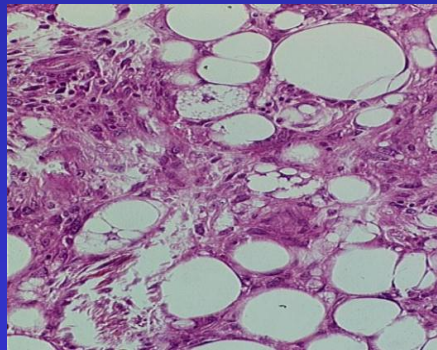
Centre Leon Bérard, Lyon, France¹; Institut Bergonie, Bordeaux, France²; Institut Gustave-Roussy, Villejuif, France³; Centre Oscar Lambret, Lille, France⁴; Hoffmann-La Roche, Nutley, NJ⁵; Roche Diagnostics GmbH, Penzberg, Germany⁶

Activated p53 Pathway Biomarkers

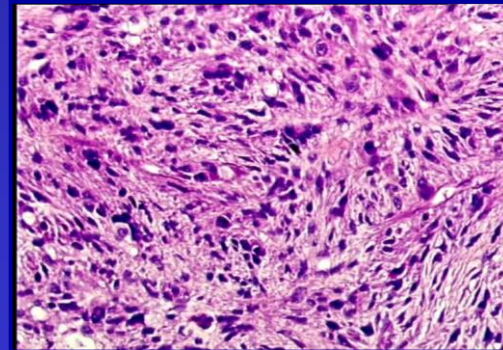


Liposarcoma: a Tumor Type Enriched for *mdm2* Amplification

- Well differentiated (WD) and de-differentiated (DD) morphologies are most common types of liposarcoma (LPS) (~80% of cases)
- Surgery is the main therapy for WD/DD LPS
 - WD LPS: low grade with low rate of recurrence and good prognosis after complete surgical resection
 - DD LPS: high grade, aggressive with higher rates of recurrence after surgery
- *mdm2* gene amplification common in LPS
 - *mdm2* gene resides on chromosome 12q13-15



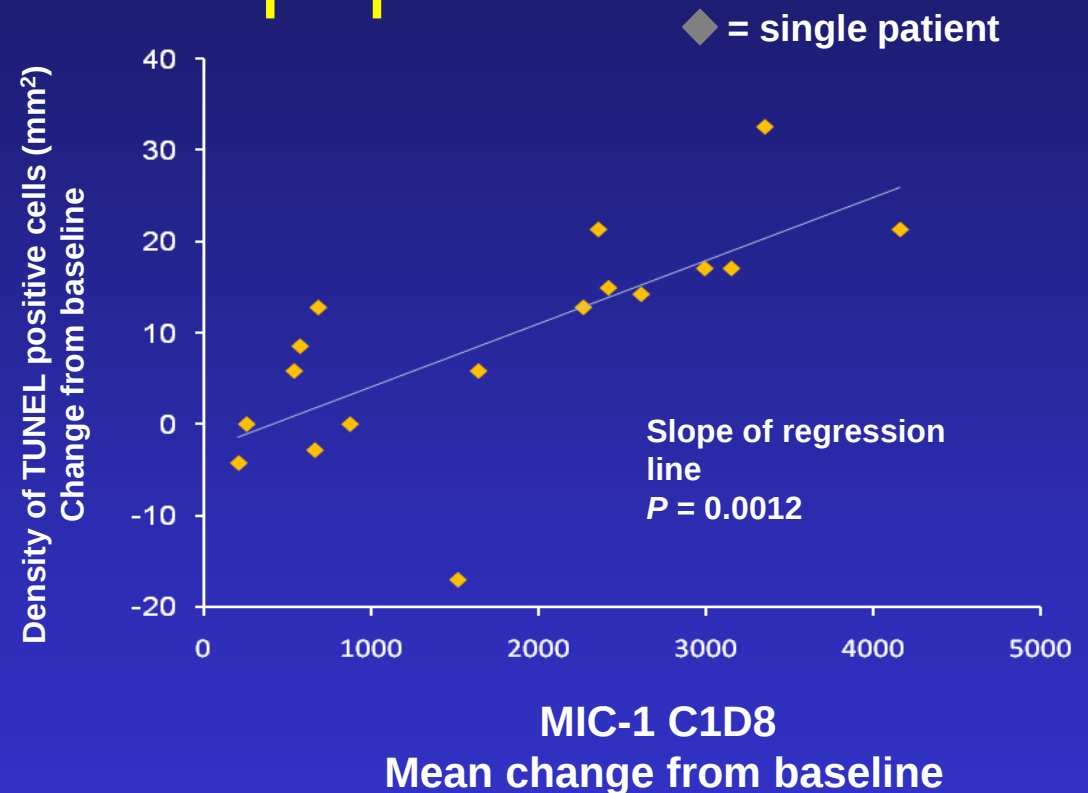
**Well
Differentiated**



**De-
Differentiated**

Increased MIC-1 Significantly Correlated With Increased Apoptosis

- Both MIC-1 and TUNEL are significantly positively correlated with drug exposure in this study
- In addition, MIC-1 blood levels and TUNEL changes in tumor biopsies were also well correlated
- Future studies will further evaluate MIC-1 blood levels as a quantitative PD marker



RG7112 in Liposarcoma

- MDM2 inhibition in human tumors activates p53, arrests cell proliferation, and induces apoptosis
- This proof of mechanism study in patients with LPS demonstrates:
 - Pharmacological p53 activation by an inhibitor of the p53-mdm2 interaction
 - Post-treatment Increases in p53, p21, and mdm2 levels
 - Exposure-related increases in MIC-1 levels
 - Post-treatment decreases proliferation as measured by change in Ki-67
 - Exposure-related induction of apoptotic signals
 - While not designed as an efficacy study, early signs of clinical activity included:
 - 1 PR after a single cycle
 - 13 SD
- This study also supports the feasibility of multiple biopsies in patients with liposarcomas eligible for surgery

Denosumab Treatment of Giant Cell Tumor of Bone: Interim Analysis of an Open-Label Phase 2 Study

David Thomas,¹ Sant Chawla,² Keith Skubitz,³ Arthur Staddon,⁴ Robert Henshaw,⁵ Armelle Dufresne⁶, Jean-Yves Blay,⁶ Judy Smith,⁷ Zhishen Ye,⁷ Winnie Sohn,⁷ Martine Roudier,⁸ Susie Jun⁷

¹Peter MacCallum Cancer Centre, East Melbourne, Victoria, Australia;

²Sarcoma Oncology Center, Los Angeles, CA, USA;

³University of Minnesota, Minneapolis, MN, USA;

⁴Pennsylvania Hematology/Oncology Associates, Philadelphia, PA, USA;

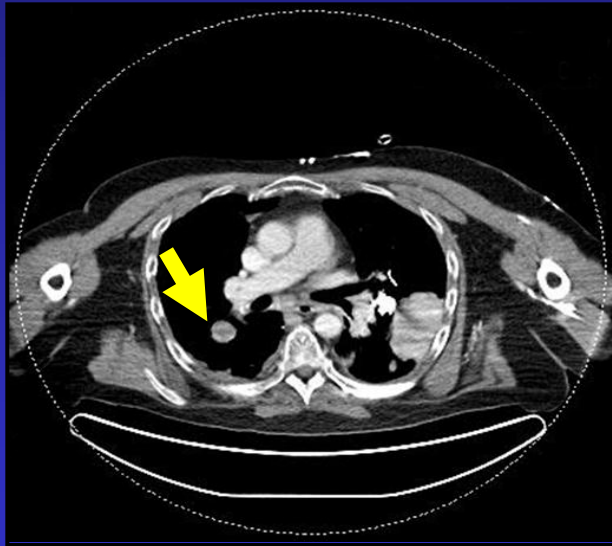
⁵Washington Cancer Institute, Georgetown University School of Medicine, Washington DC, USA;

⁶Hôpital Edouard Heriot, Lyon, France;

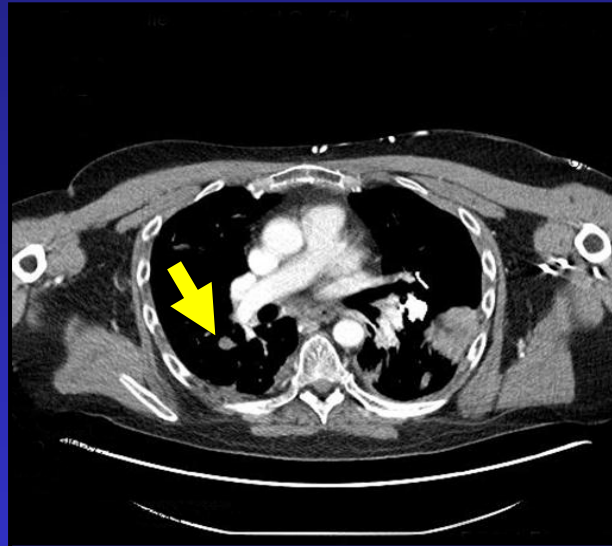
⁷Amgen Inc., Thousand Oaks, CA, USA;

⁸Amgen Inc., Seattle, WA, USA

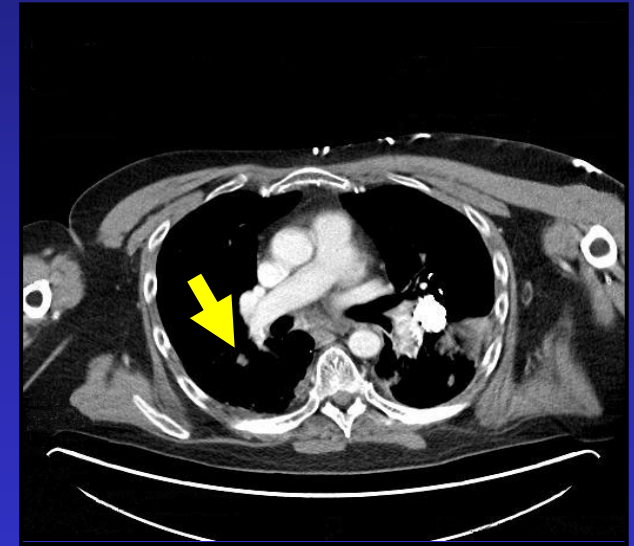
Radiologic Response to Denosumab



Baseline



Week 5



Week 37

Targeted treatments of sarcomas and connective tissue tumors

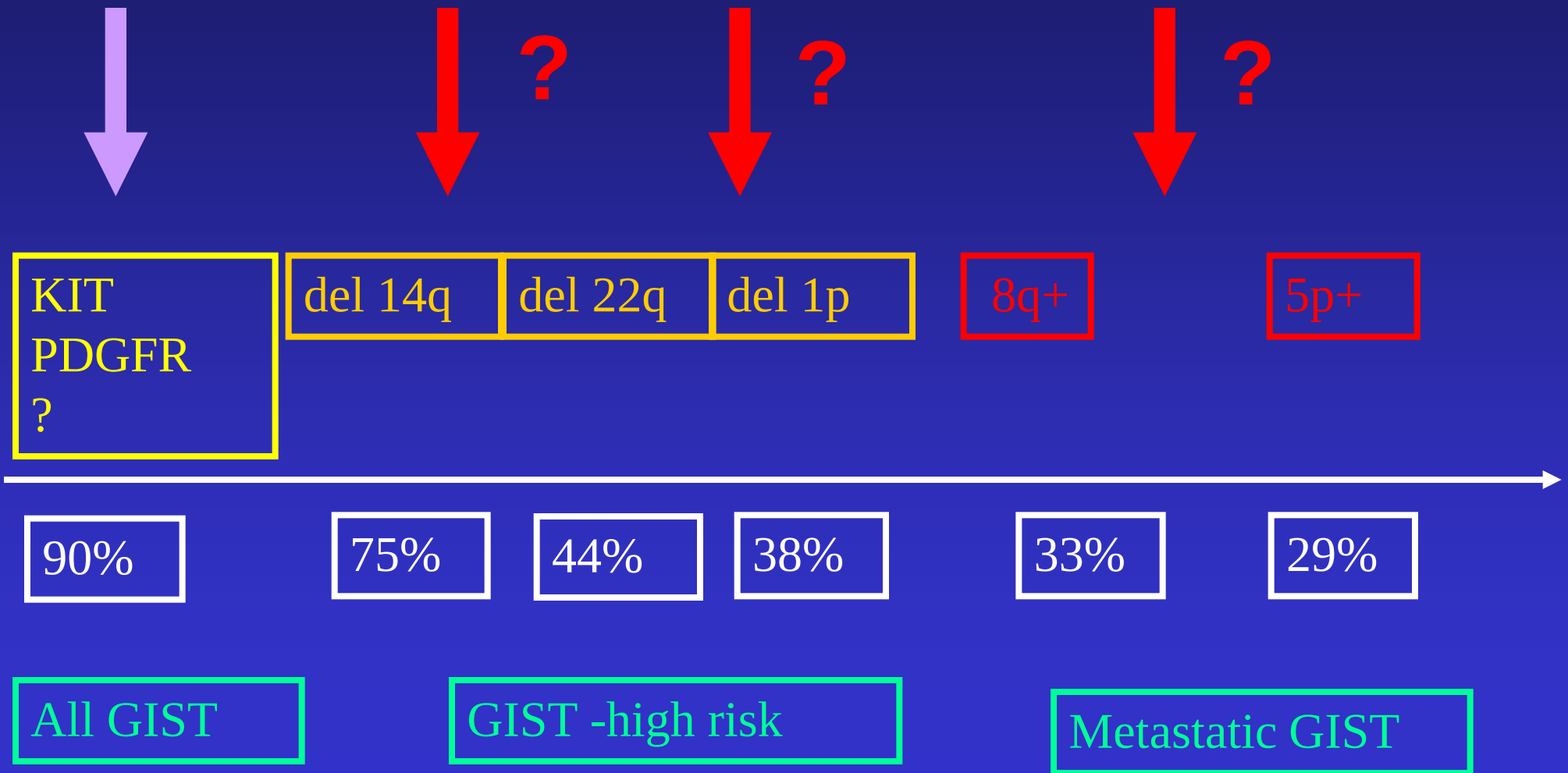
ESMO 2012

• GIST	Imatinib 400
• GIST KIT exon 9	Imatinib 800
• GIST PDGFR D842V	crenolanib?
• Other GIST	Imatinib 400
• GIST 2nd line	Sunitinib
• GIST 3rd line	Regorafenib
• DFSP	Imatinib
• GCT of the bone	Denosumab
• PEComas	mTOR inh.
• PVNS	CSF1R antagonists
• ASPS	Cediranib/VEGFR2 inhibitors
• Ewing/OS/RMS subsets	IGF1R Ab
• MLPS	trabectedine

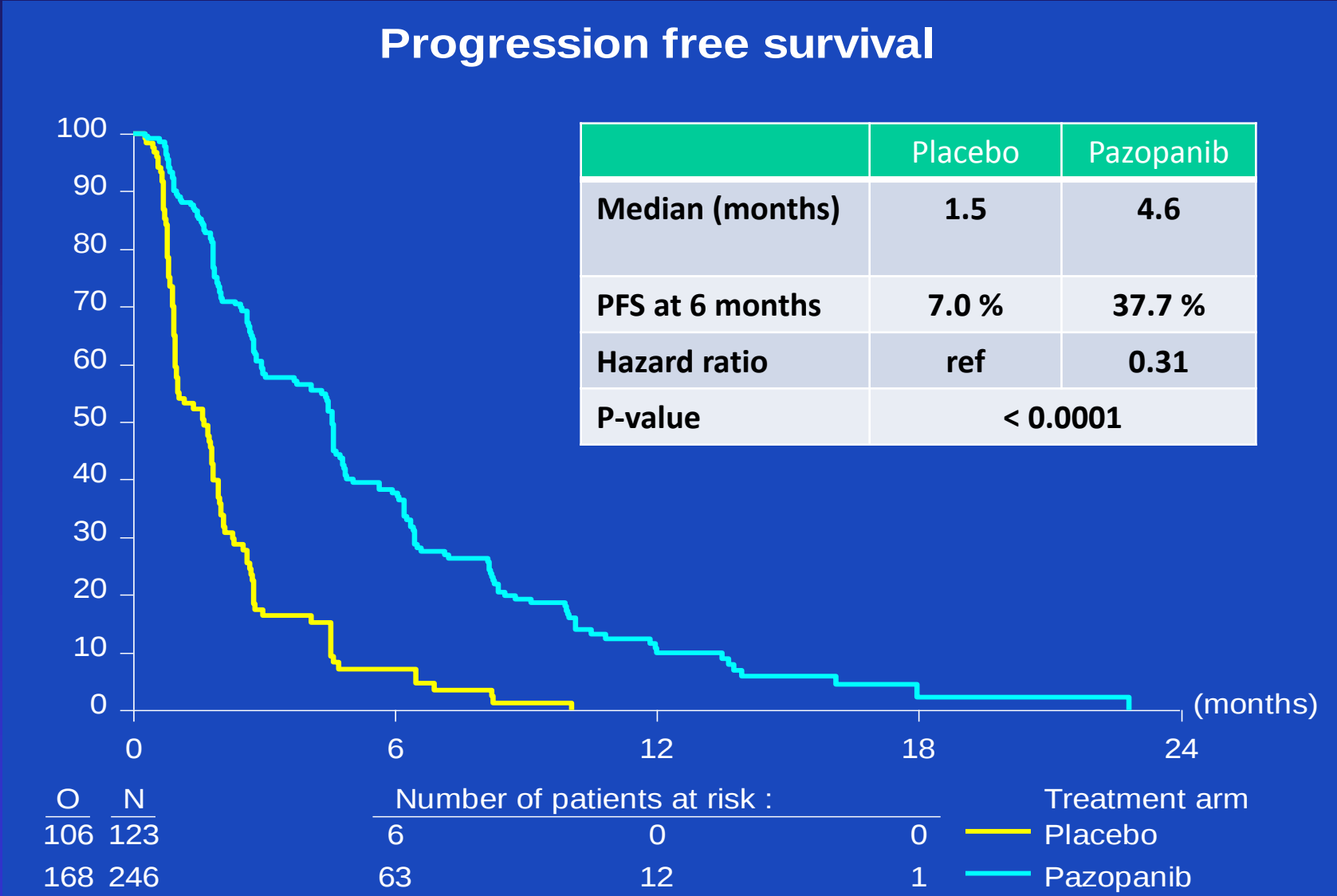
Three situations

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 - PDGFR expression in normal (and malignant) cells of connective tissue

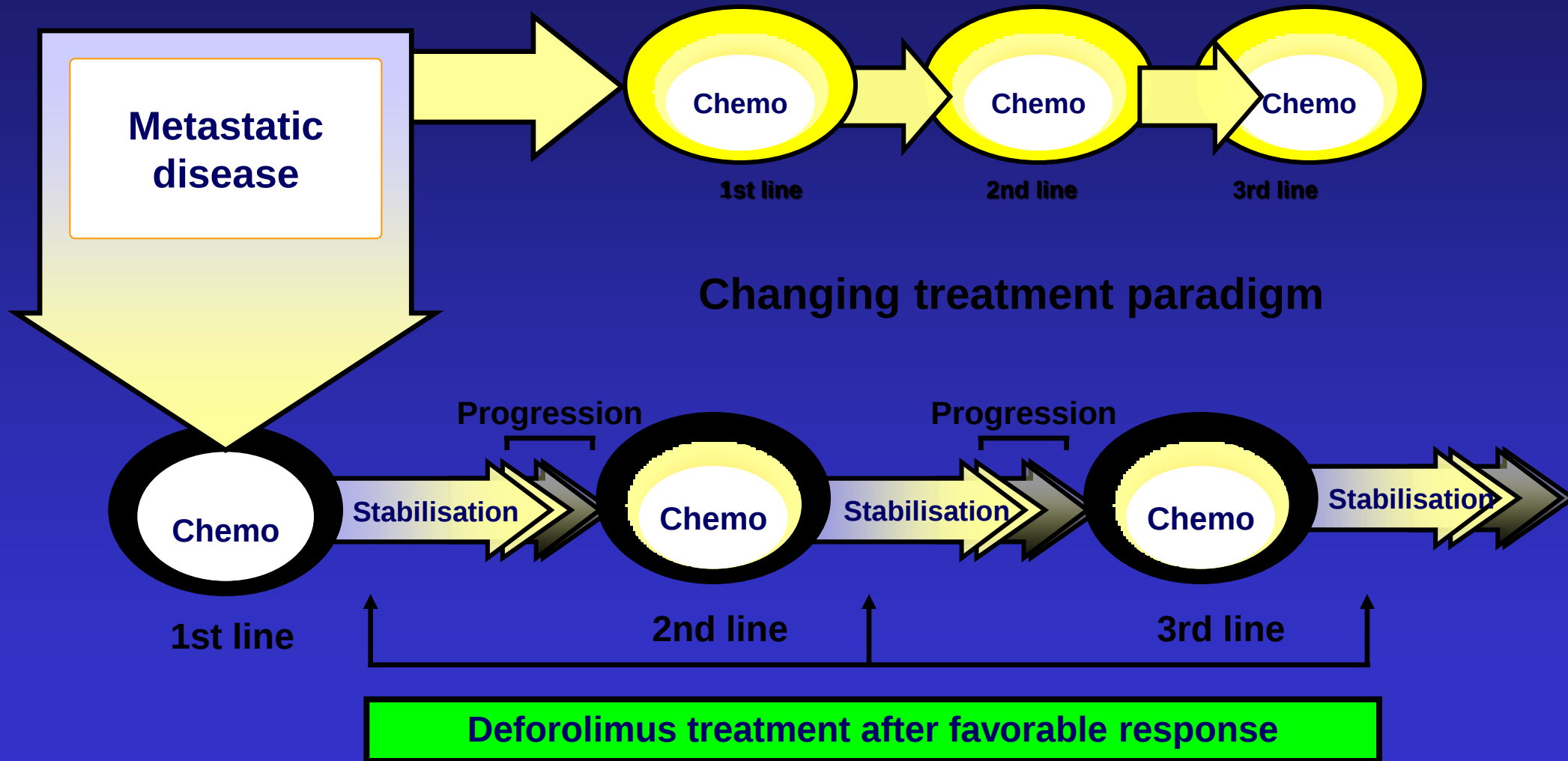
2.Targeting a « latter » event



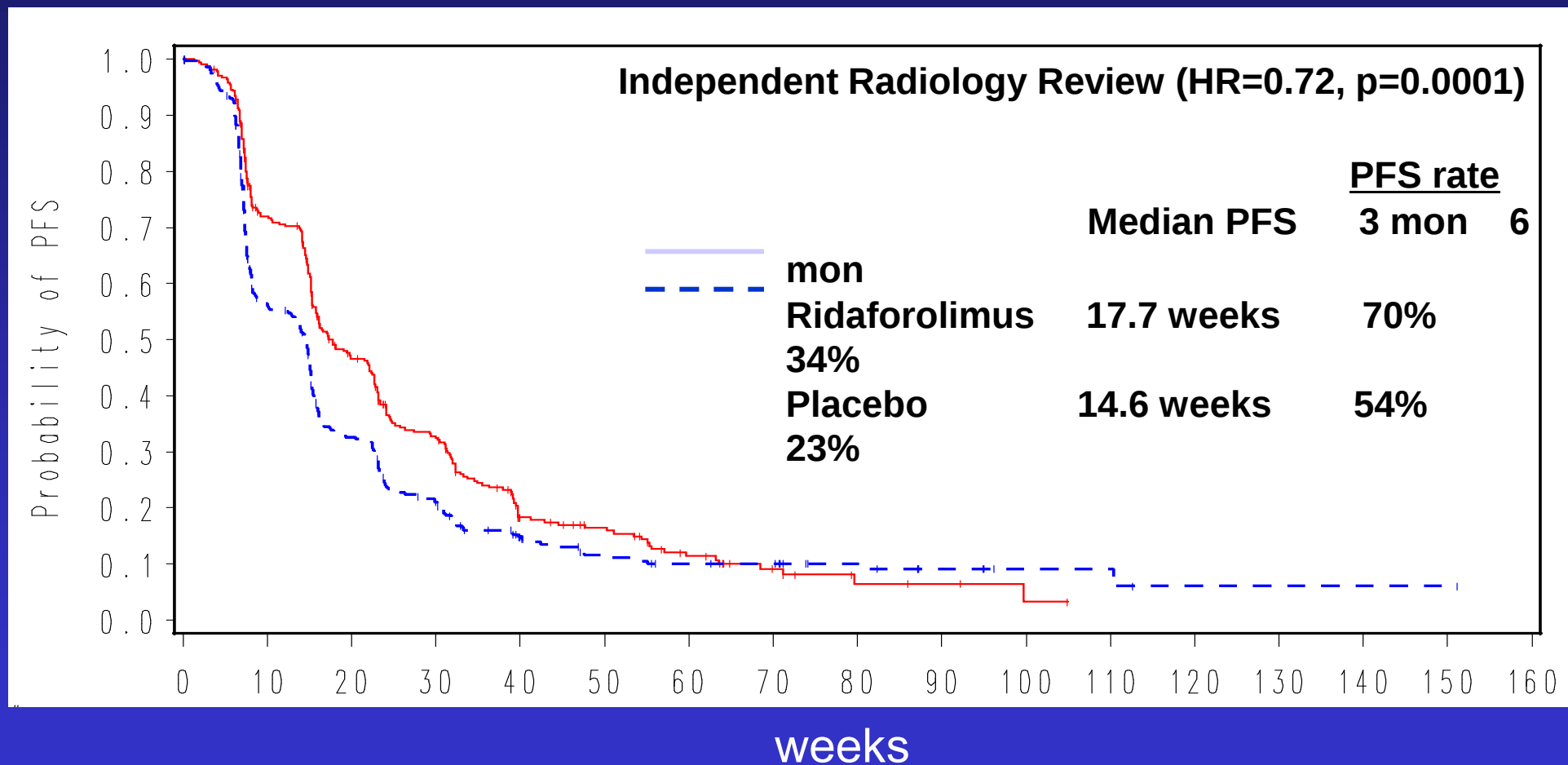
Palette: primary efficacy end-point



SUCCEED : Ridaforolimus in maintenance



SUCCEED: PFS



Ridaforolimus : a need for biomarkers?

Table 4: Summary of Potential Biomarkers Examined in Archival Tissue

Protein	Staining Index				p-value ^b
	CBR		Non-CBR		
	n ^a	Median	n ^a	Median	
Upstream of mTOR					
PTEN	28	70	33	90	0.96
p-AKT	30	75	31	90	0.73
FKBP12	15	200	12	230	0.52
IGF-1R	15	0	12	0	0.26
Downstream of mTOR					
p-S6	19	165	48	97.5	0.47
4E-BP1	18	180	46	200	0.30
eIF4E	19	135	47	130	0.52
p27kip1	26	25	58	12.5	0.48

CBR = Clinical benefit response

a: Number of patient samples analyzed

b: p-value for the difference in staining index between CBR and non-CBR; Kruskal-Wallis test

Three situations

- Initial molecular event
 - KIT in GIST
 - Loss NF1, TSC
 - Translocations
 - Mdm2 amplification
 - ...
- Secondary event
 - VEGF production
 - Activation of mTOR pathway
 - ER expression in ESS
- Simple bystander
 - PDGFR expression in normal (and malignant) cells of connective tissue

Overall and progression free survival

O	N	Number of patients at risk :												event
26	48	24	16	10	7	4	2	1	1	1	1	1		OS
47	48	10	4	2	1	0	0	0	0	0	0	0		PFS

event

— OS

PFS

Systemic treatment of sarcomas 2000-2012 ESMO 2012

2000:

- All sarcomas
 - Doxorubicin
 - Ifosfamide
 - DTIC
- Ewing , RMS, OS
 - Dactinomycin
 - CDDP
 - Vincaalcaloids
 - Cyclophosphamide
 - HDMTX

2012

- All subtypes : Trabectedin
- GIST: Imatinib, sunitinib,
- DFSP : Imatinib
- Osteosarcomas: MTPPE
- LMS: Gemcitabine (+/- DTIC or Tax)
- EWS: TopoI inh., IGF1R
- A/E RMS: Topo inh
- Angio: Dox, Paclitaxel, GemTax
- ESS : Aromatase inh.
- All but LPS: pazopanib
- PVNS :Imatinib
- Desmoid Tumors : HT, imatinib
- GCTB : denosumab
- PECOMAs: mTOR
- WD/DD: LPS:CDK4, MDM2
- GIST: regorafenib
- ASPS: cediranib



Subtype-specific genomic alterations define new targets for soft-tissue sarcoma therapy

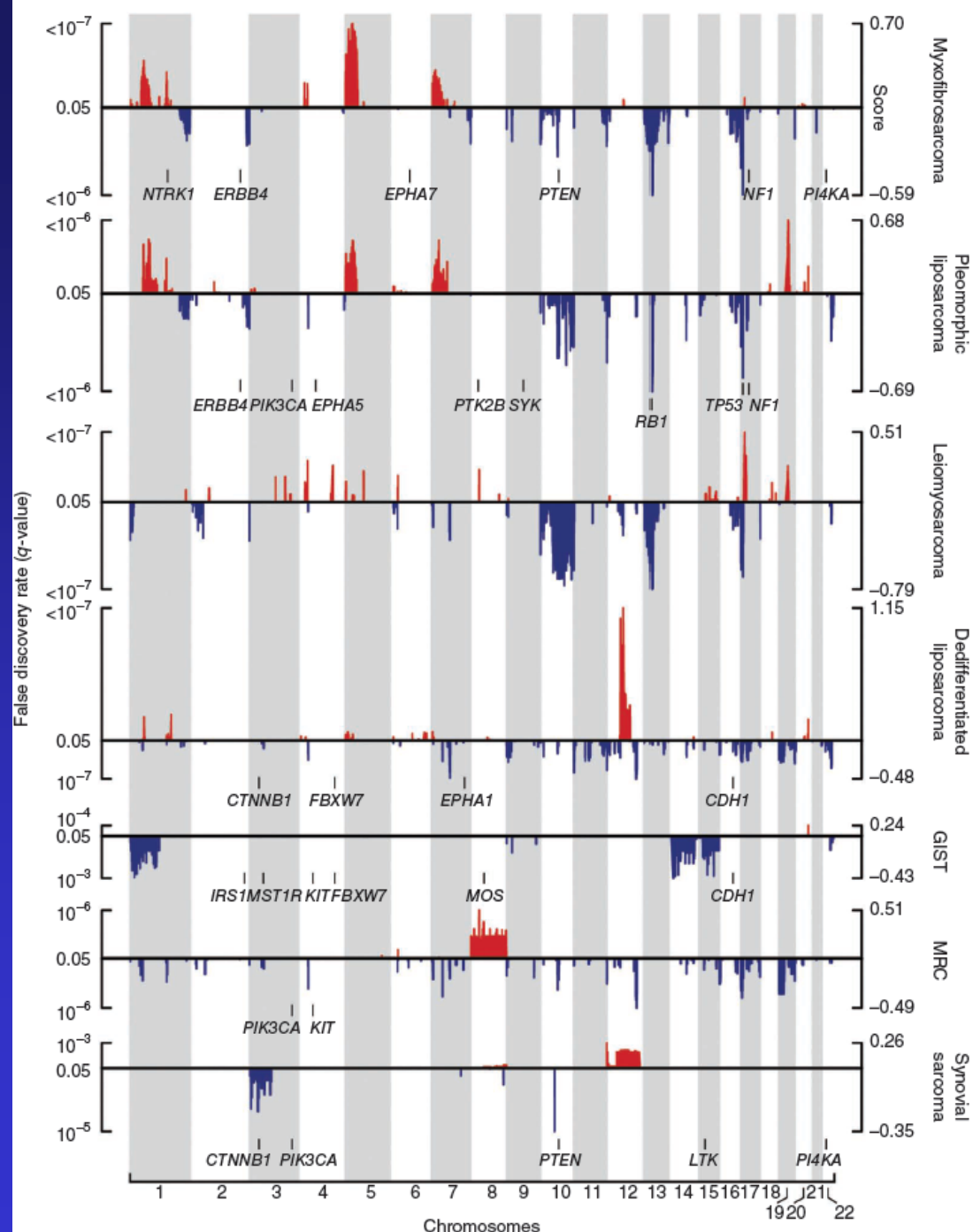
Jordi Barretina^{1-3,15}, Barry S Taylor^{4,5,15}, Shantanu Banerji¹⁻³, Alexis H Ramos¹⁻³, Mariana Lagos-Quintana⁶, Penelope L DeCarolis⁶, Kinjal Shah¹⁻³, Nicholas D Socci⁴, Barbara A Weir¹⁻³, Alan Ho⁷, Derek Y Chiang¹⁻³, Boris Reva⁴, Craig H Mermel¹⁻³, Gad Getz³, Yevgeniy Antipin⁴, Rameen Beroukhi¹⁻³, John E Major⁴, Charles Hatton^{1,2}, Richard Nicoletti^{1,2}, Megan Hanna^{1,2}, Ted Sharpe³, Tim J Fennell³, Kristian Cibulskis³, Robert C Onofrio³, Tsuyoshi Saito^{8,9}, Neerav Shukla^{8,9}, Christopher Lau^{8,9}, Sven Nelander⁴, Serena J Silver³, Carrie Sougnez³, Agnes Viale¹⁰, Wendy Winckler¹⁻³, Robert G Maki¹¹, Levi A Garraway¹⁻³, Alex Lash⁴, Heidi Greulich¹⁻³, David E Root³, William R Sellers¹², Gary K Schwartz⁷, Cristina R Antonescu⁸, Eric S Lander³, Harold E Varmus¹³, Marc Ladanyi^{8,9}, Chris Sander⁴, Matthew Meyerson^{1-3,14,16} & Samuel Singer^{6,16}

Table 2 Mutations identified in soft-tissue sarcoma

Gene	No. of mut. ^a	Subtype	Tumor ID	Cases affected (%) ^b	mRNA	Protein
<i>CDH1</i>	2	DDLPS	PT7DD	2.0	712A>AG	N238D
		GIST	PT61GT	4.5	1849G>AG	A617T ^c
<i>CTNNB1</i>	2	DDLPS	PT18DD	2.0	122C>CT	T41I ^d
		Synovial	PT195SYN	4.0	95A>AT	D32V ^d
<i>EPHA1</i>	1	DDLPS	PT10DD	2.0	634G>GA	A212T
<i>EPHA5</i>	1	Pleomorphic	PT182PL	4.2	2386A>AG	Y796H
<i>EPHA7</i>	1	MYXF	PT106MF	2.6	1649C>CT	S550N
<i>ERBB4</i>	2	MYXF	PT130MF	2.6	3437A>AT	D1146V
		Pleomorphic	PT167PL	4.2	1558A>AT	C520S
<i>FBXW7</i>	2	DDLPS	PT38DD	2.0	338_342delTCATC>TC	E113fs
		GIST	PT58GT	4.5	563G>GT	C188F
<i>IRS1</i>	1	GIST	PT61GT	4.5	3406C>CT	E1136K
<i>KIT</i>	6	GIST	PT57GT	23.0	1727T>CT	L576P ^d
		GIST	PT63GT		1961T>CT	V654A ^d
		GIST	PT61GT		1667_1674delAGTGGAAG>AG	Q556fs
		GIST	PT60GT		1667_1687del ^e	Q556_I563>Q
		GIST	PT59GT		1670_1675delGGAAGG	W557_V559>F ^c
		MRC	PT149MRC	4.8	2334G>CG	K778N
<i>LTK</i>	1	Synovial	PT190SYN	4.0	2243_2244delTT>T	C748fs
<i>MOS</i>	1	GIST	PT61GT	4.5	898A>AG	S300P
<i>MST1R</i>	1	GIST	PT60GT	4.5	1229G>AG	P410L
<i>NF1</i>	7	MYXF	PT104MF	10.5	7972C>CT	H2658Y
		MYXF	PT104MF		7790C>CT	S2597L
		MYXF	PT127MF		910C>T	R304* ^d
		MYXF	PT134MF		910C>T	R304* ^d
		MYXF	PT102MF		7010T>TG	L2337R
		Pleomorphic	PT176PL	8.3	1105C>CT	Q369* ^d
		Pleomorphic	PT179PL		4006C>CT	Q1336*
<i>NTRK1</i>	1	MYXF	PT101MF	2.6	2338C>CT	R780W
<i>PI4KA</i>	2	MYXF	PT137MF	2.6	4081_4088delTCTTATCT>TCT	1361fs
		Synovial	PT203SYN	4.0	4081_4088delTCTTATCT>TCT	1361fs
<i>PIK3CA</i>	6	MRC	PT143MRC	18.0	1633G>AG	E545K ^e
		MRC	PT149MRC		1633G>AG	E545K ^e
		MRC	PT138MRC		3140A>AG	H1047R ^e
		MRC	PT158MRC		3140A>AG	H1047R ^e
		Pleomorphic	PT173PL	4.2	1660delC	H554fs
		Synovial	PT195SYN	4.0	1659delT	S553fs
<i>PTEN</i>	2	MYXF	PT100MF	2.6	G>CG	Splice site
		Synovial	PT206SYN	4.0	106G>AA	G36R ^c
<i>PTK2B</i>	1	Pleomorphic	PT163PL	4.2	G>AG	Splice site
<i>RB1</i>	1	Pleomorphic	PT167PL	4.2	1818T>TA	Y606* ^c
<i>SYK</i>	1	Pleomorphic	PT163PL	4.2	52G>AA	G18S
<i>TP53</i>	4	Pleomorphic	PT163PL	16.7	404C>AA	C135F ^c
		Pleomorphic	PT169PL		464G>AA	T155I
		Pleomorphic	PT173PL		C>CT	Splice site
		Pleomorphic	PT164PL		C>TT	Splice site

DDLPS, dedifferentiated liposarcoma; MYXF, myxofibrosarcoma.

^aNumber of nonsynonymous or splice-site mutations detected in either primary sequencing or extended genotyping. ^bPercentage of cases by subtype. ^cMutations previously identified in any cancer type, as indicated in the Catalogue of Somatic Mutations in Cancer. ^dMutations previously identified in soft-tissue sarcoma. ^eReference allele: GTGGAAGGTTGTGAGGAGAT. Asterisks indicate nonsense mutations.



ARTICLES

The landscape of somatic copy-number alteration across human cancers

Rameen Beroukhi^{1,3,4,5*}, Craig H. Mermel^{1,3*}, Dale Porter⁸, Guo Wei¹, Soumya Raychaudhuri^{1,4}, Jerry Donovan⁸, Jordi Barretina^{1,3}, Jesse S. Boehm¹, Jennifer Dobson^{1,3}, Mitsuyoshi Urashima⁹, Kevin T. Mc Henry⁸, Reid M. Pinchback¹, Azra H. Ligon⁴, Yoon-Jae Cho⁶, Leila Haery^{1,3}, Heidi Greulich^{1,3,4,5}, Michael Reich¹, Wendy Winckler¹, Michael S. Lawrence¹, Barbara A. Weir^{1,3}, Kumiko E. Tanaka^{1,3}, Derek Y. Chiang^{1,3,13}, Adam J. Bass^{1,3,4}, Alice Loo⁸, Carter Hoffman^{1,3}, John Prensner^{1,3}, Ted Liefeld¹, Qing Gao¹, Derek Yecies³, Sabina Signoretti^{3,4}, Elizabeth Maher¹⁰, Frederic J. Kaye¹¹, Hidefumi Sasaki¹², Joel E. Tepper¹³, Jonathan A. Fletcher⁴, Josep Tabernero¹⁴, José Baselga¹⁴, Ming-Sound Tsao¹⁵, Francesca Demicheli¹⁶, Mark A. Rubin¹⁶, Pasi A. Janne^{3,4}, Mark J. Daly^{1,17}, Carmelo Nucera⁷, Ross L. Levine¹⁸, Benjamin L. Ebert^{1,4,5}, Stacey Gabriel¹, Anil K. Rustgi¹⁹, Cristina R. Antonescu¹⁸, Marc Ladanyi¹⁸, Anthony Letai³, Levi A. Garraway^{1,3}, Massimo Loda^{3,4}, David G. Beer²⁰, Lawrence D. True²¹, Aikou Okamoto²², Scott L. Pomeroy⁶, Samuel Singer¹⁸, Todd R. Golub^{1,3,23}, Eric S. Lander^{1,2,5}, Gad Getz¹, William R. Sellers⁸ & Matthew Meyerson^{1,3,5}

Sarcomas, as many rare tumors, are valuable models for « frequent » tumors on their way to fragmentation

LETTER

doi:10.1038/nature11003

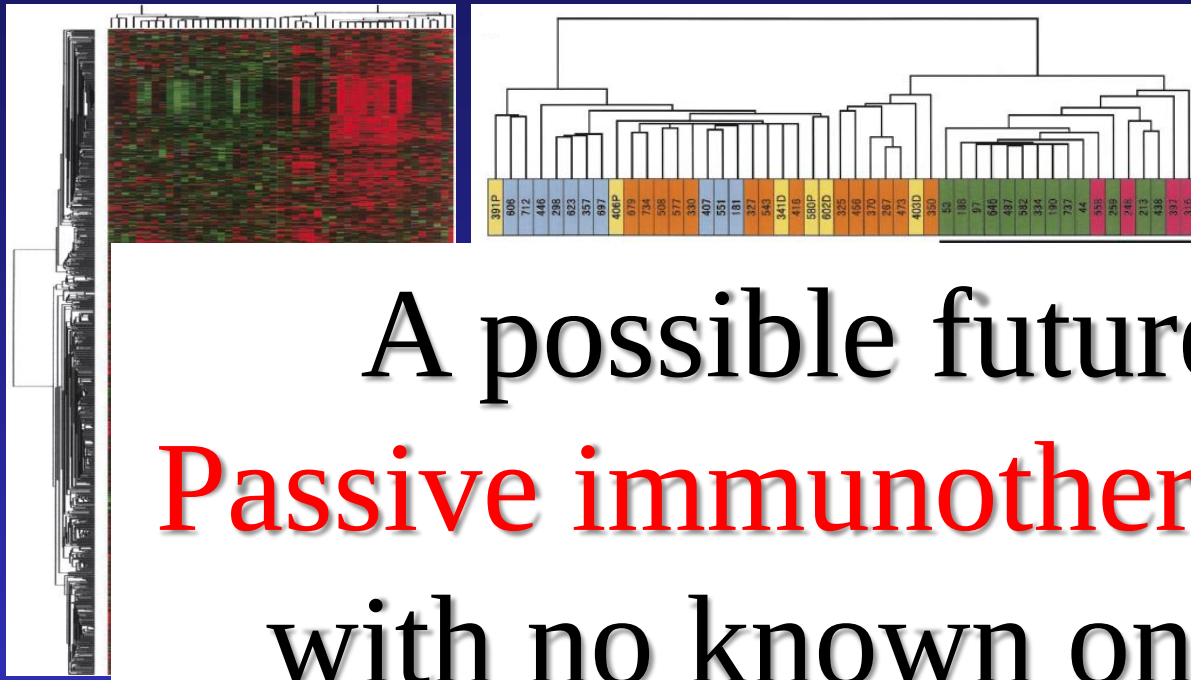
The Cancer Cell Line Encyclopedia enables predictive modelling of anticancer drug sensitivity

Jordi Barretina^{1,2,3†*}, Giordano Caponigro^{4*}, Nicolas Stransky^{1*}, Kavitha Venkatesan^{4*}, Adam A. Margolin^{1†*}, Sungjoon Kim⁵, Christopher J. Wilson⁴, Joseph Lehar⁴, Gregory V. Kryukov¹, Dmitriy Sonkin⁴, Anupama Reddy⁴, Manway Liu⁴, Lauren Murray¹, Michael F. Berger^{1†}, John E. Monahan⁴, Paula Morais¹, Jodi Meltzer⁴, Adam Korejwa¹, Judit Jané-Valbuena^{1,2}, Felipa A. Mapa⁴, Joseph Thibault⁵, Eva Bric-Furlong⁴, Pichai Raman⁴, Aaron Shipway⁵, Ingo H. Engels⁵, Jill Cheng⁶, Guoying K. Yu⁶, Jianjun Yu⁶, Peter Aspesi Jr⁴, Melanie de Silva⁴, Kalpana Jagtap⁴, Michael D. Jones⁴, Li Wang⁴, Charles Hatton³, Emanuele Palescandolo³, Supriya Gupta¹, Scott Mahan¹, Carrie Sougnez¹, Robert C. Onofrio¹, Ted Liefeld¹, Laura MacConaill³, Wendy Winckler¹, Michael Reich¹, Nanxin Li⁵, Jill P. Mesirov¹, Stacey B. Gabriel¹, Gad Getz¹, Kristin Ardlie¹, Vivien Chan⁶, Vic E. Myer⁴, Barbara L. Weber⁴, Jeff Porter⁴, Markus Warmuth⁴, Peter Finan⁴, Jennifer L. Harris⁵, Matthew Meyerson^{1,2,3}, Todd R. Golub^{1,3,7,8}, Michael P. Morrissey^{4*}, William R. Sellers^{4*}, Robert Schlegel^{4*} & Levi A. Garraway^{1,2,3*}

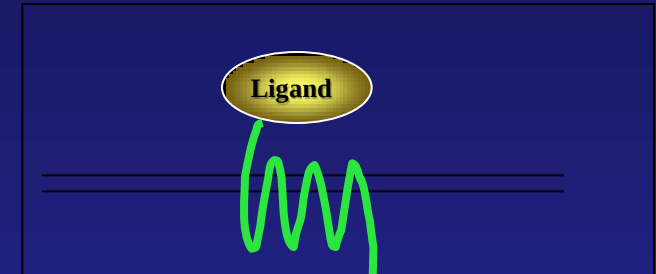
FZD10 is specifically up-regulated in synovial sarcoma

ESMO 2012

Genome-wide cDNA microarray of soft tissue sarcoma

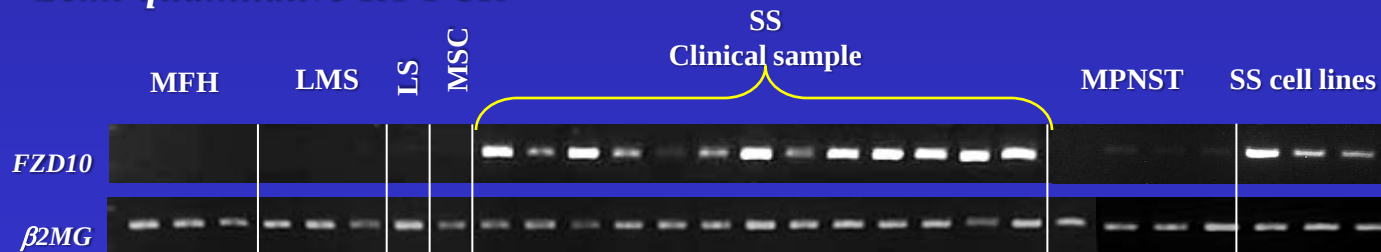


Frizzled Homologue 10 (FZD10)



A possible future strategy?
Passive immunotherapy for targets
with no known oncogenic role

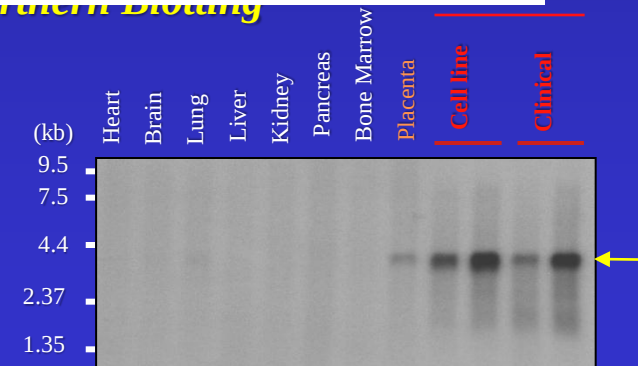
Semi-quantitative RT-PCR



MFH, Malignant fibrous histiocytoma; LMS, Leiomyosarcoma; LS, Liposarcoma; MSC, Mesenchymal stem cell; SS, Synovial Sarcoma; MPNST, Malignant peripheral nerve sheath tumor

Nagayama et al., Cancer Research 62, 5859-66 (2002)

Northern Blotting

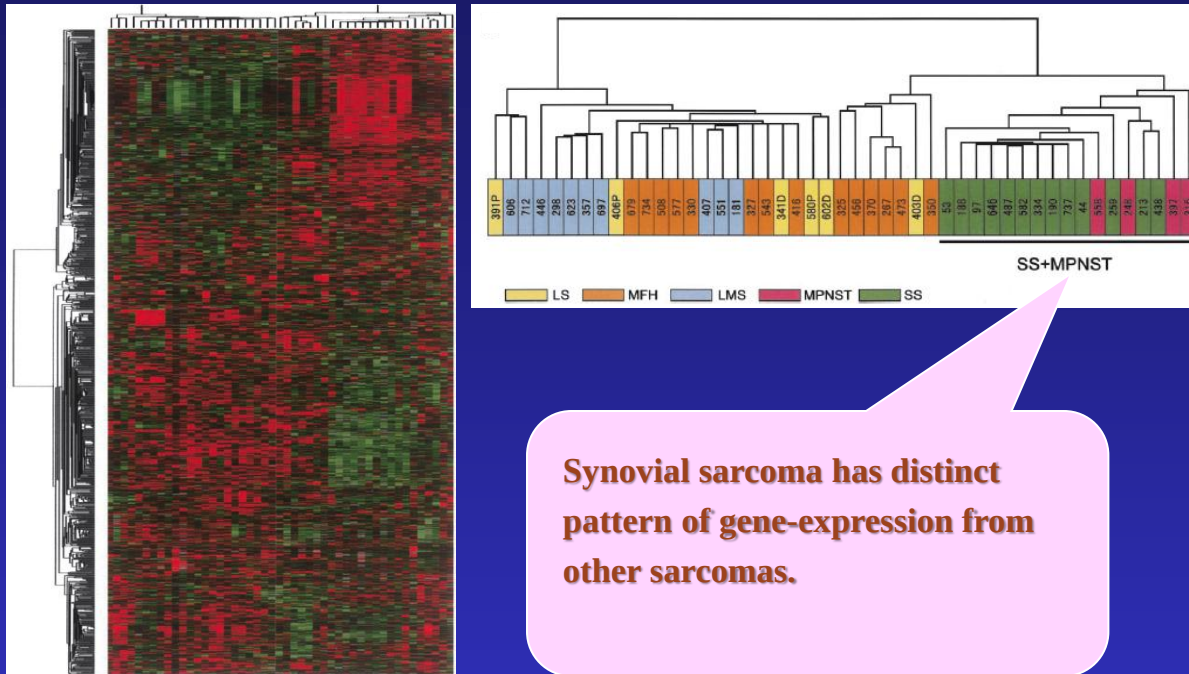


Nagayama et al., Oncogene 24, 6201-12 (2005)

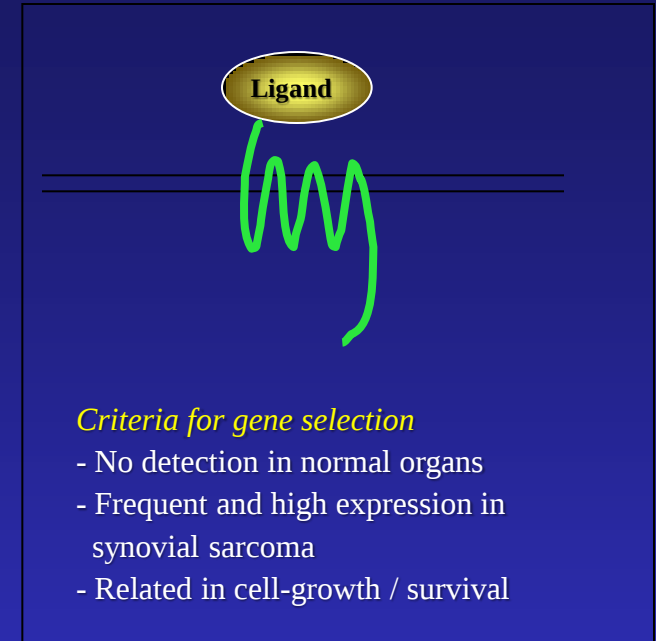
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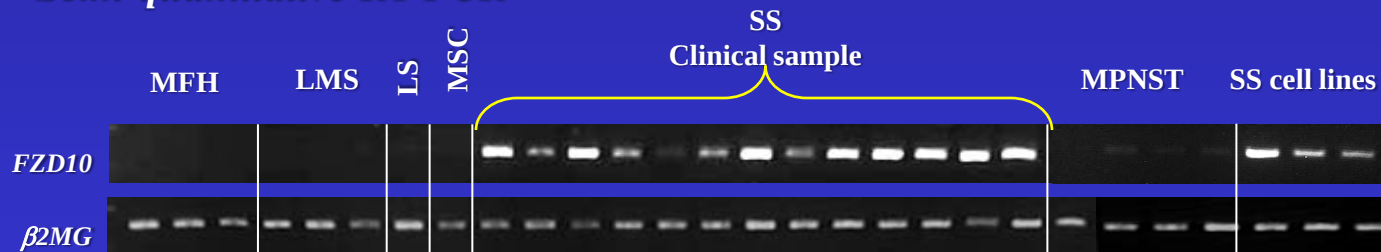
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Frizzled Homologue 10 (FZD10)



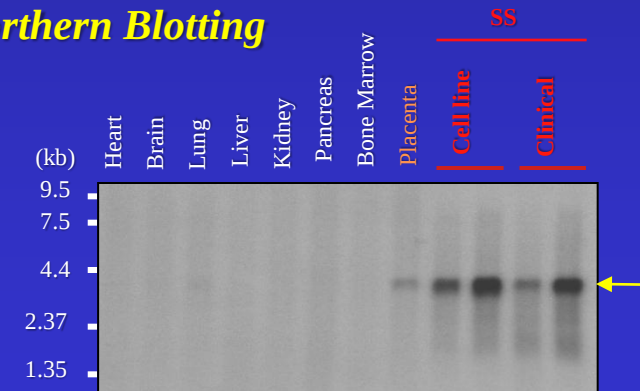
Semi-quantitative RT-PCR



MFH , Malignant fibrous histiocyteoma ; LMS , Leiomyosarcoma ; LS, Liposarcoma ; MSC , Mesenchymal stem cell ; SS , Synovial Sarcoma ; MPNST , Malignant peripheral nerve sheath tumor

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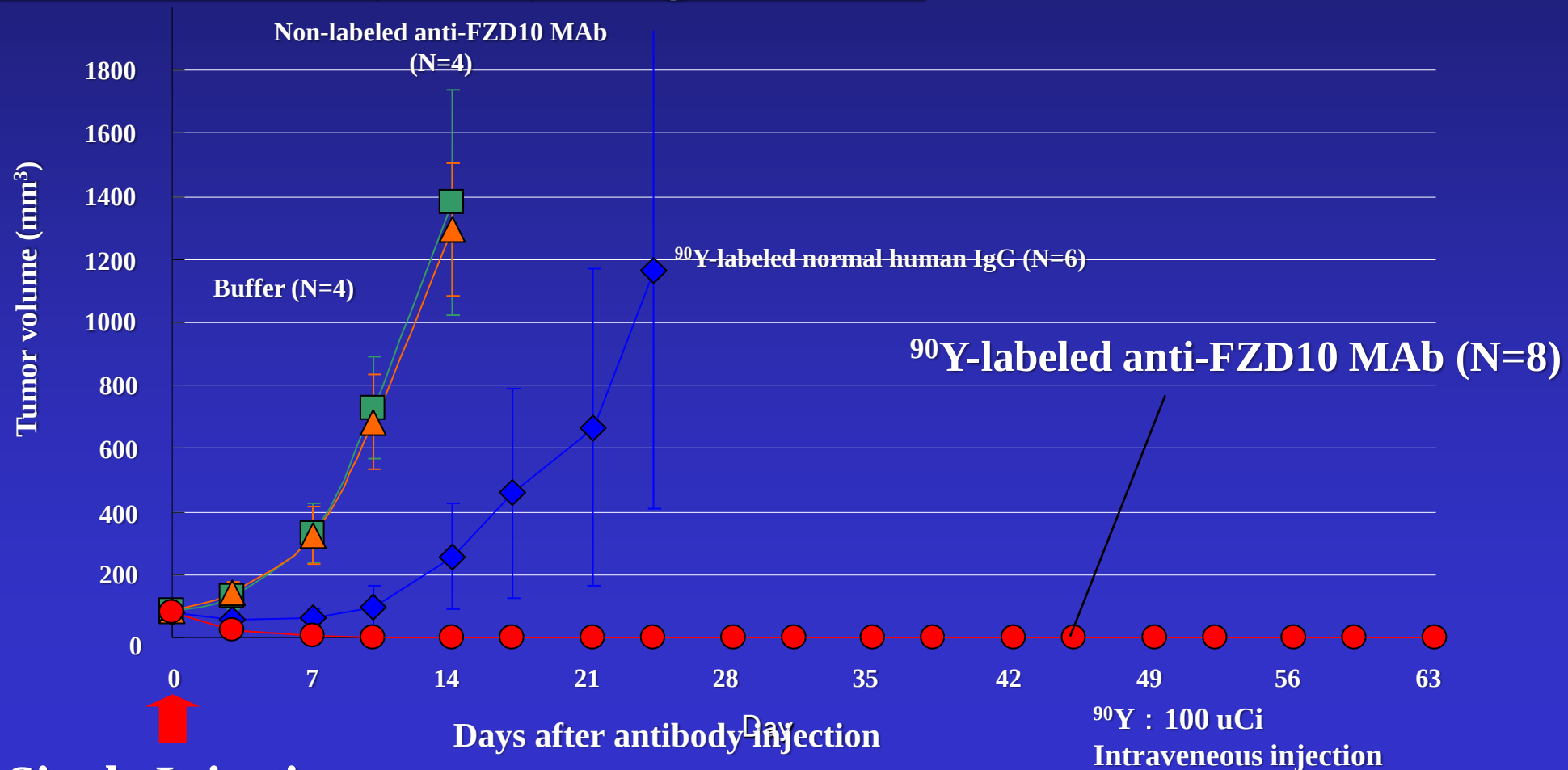


Nagayama et al., *Oncogene* 24, 6201-12 (2005)

Antitumor activity of ^{90}Y -labeled anti-FZD10 MAb ESMO 2012

Potent antitumor effect was shown in all xenograft mice by just single injection of ^{90}Y -labeled anti-FZD10 MAb.

Synovial sarcoma (SYO-1) xenograft mouse



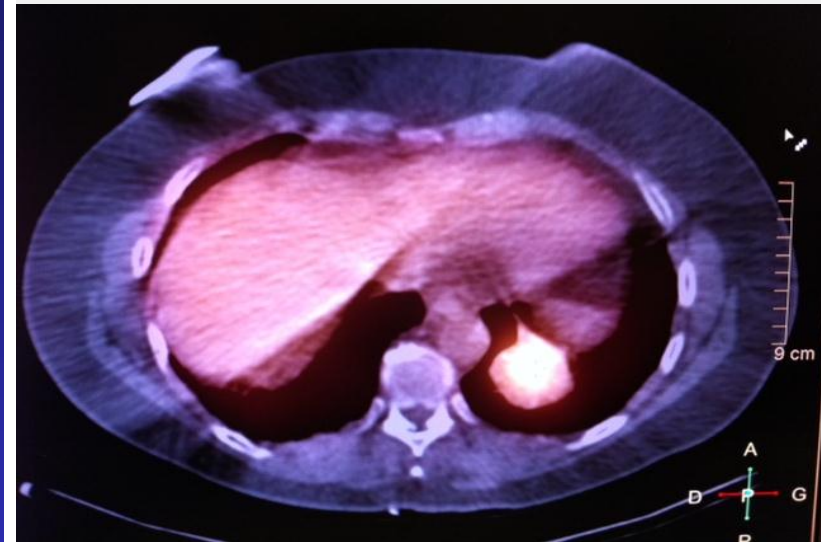
Single Injection

SYNFRIZZ Clinical Trial

SYNFRIZZ
Vial A (OTSA101-D)
Pour injection intraveineuse



Transversal image
24H uptake
Met1 has very good uptake



Distal image
24h uptake



144H
Met1 & Met 2



Distal image
144h uptake

Conclusions

- Rare and multiple heterogeneous subtypes and molecular subtypes
- Models for oncogenesis and clinical research
- Treatment adapted to driving molecular alterations
- Translational research to identify further driver mutations
- The challenge of resistance



Thanks to EORTC Soft Tissue and Bone Sarcoma Group

Peter Hohenberger, Ian Judson, Pancras Hogendorn, Jaap Verweij,
Winette van der Graf, Alessandro Gronchi, Martine Van Glabbeke
and all investigators

A World Sarcoma Network is needed

