

Direct targeting of RAS: small molecules

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Reader in Molecular Cancer Pharmacology

The Institute of Cancer Research / The Royal
Marsden Hospital

ESMO symposium on signalling pathways in cancer

Targeting the MAPK pathway: From RAS to MEK

13th March 2015

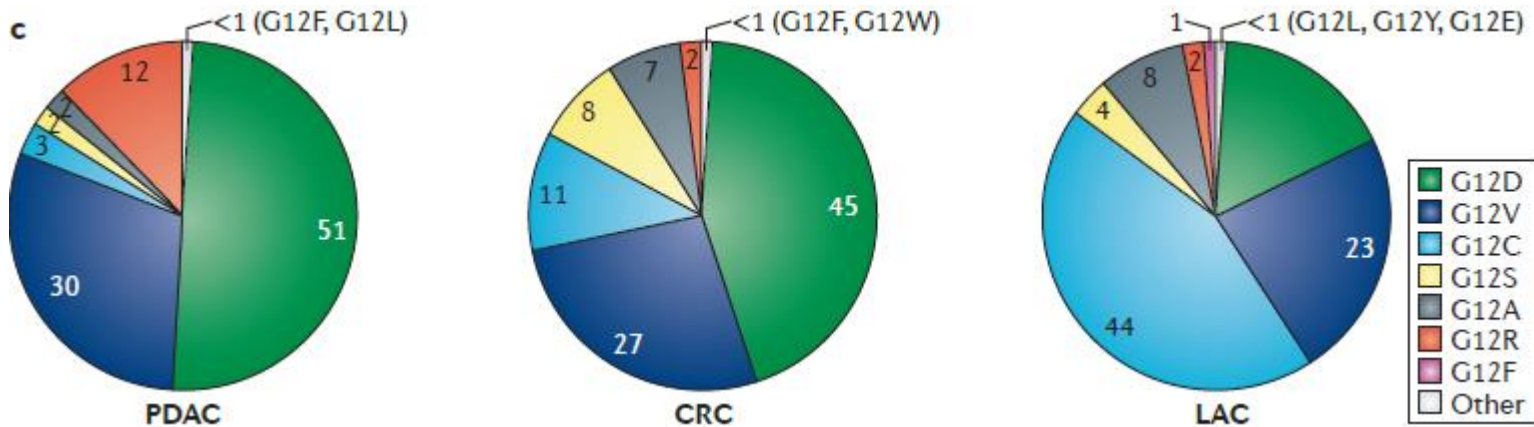
Conflicts of interest

- I have no conflict of interest related to the subject matter discussed in this talk

The usual suspects

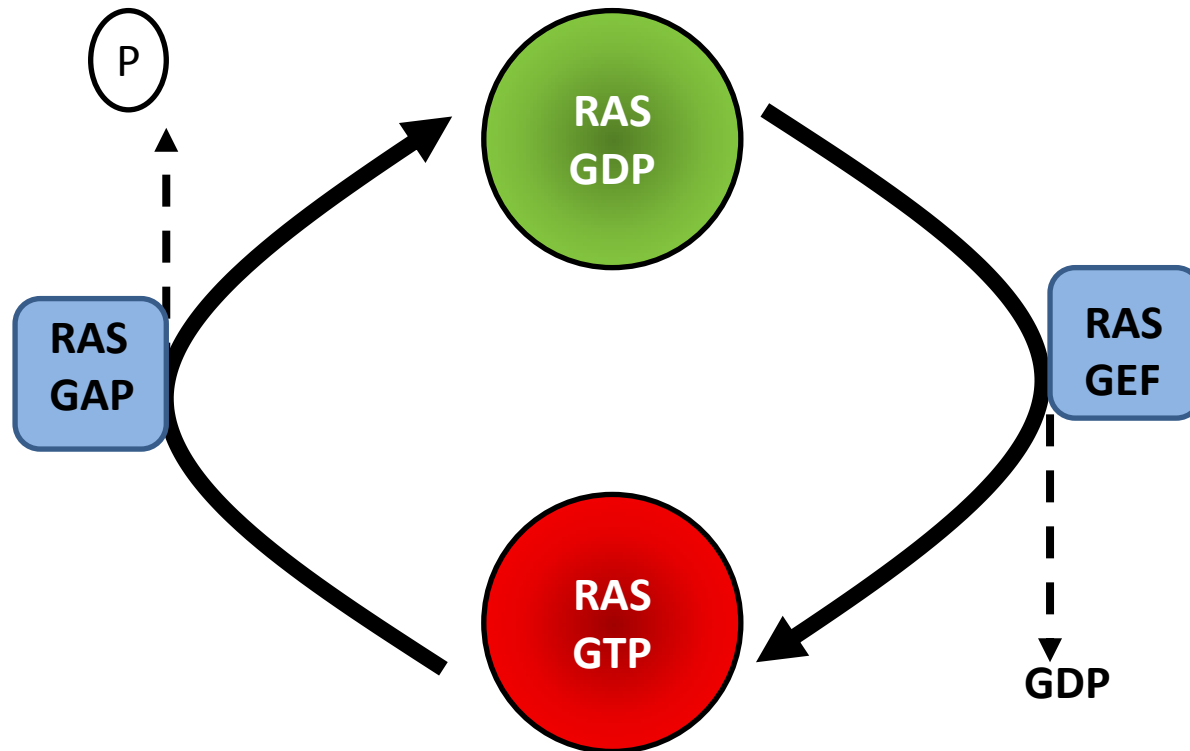


Tumour	% <i>KRAS</i>	% <i>NRAS</i>	% <i>HRAS</i>	% <i>RAS</i>
Pancreatic	97.7	0	0	97.7
Colorectal	44.7	7.5	0	52.2
Multiple Myeloma	22.8	19.9	0	42.6
Lung adenocarcinoma	30.9	0.9	0.3	32.2
Melanoma	0.8	27.6	1	29.1



- The frequency and distribution of *RAS* gene mutations are not uniform
- Often single base missense mutations on residues G12, G13 and Q61
- Drugs targeting *RAS* will have different effects on different isoforms and thus different effects on tumours

RAS- The physiological problem



- While ATP binds to protein kinases at low μM affinity, GTP binds to RAS with pM affinity
- GTP/GDP present in μM concentrations in cells, including cancer cells

RAS- The pathological problem

Proc. Natl. Acad. Sci. USA
Vol. 81, pp. 5704-5708, September 1984
Biochemistry

Intrinsic GTPase activity distinguishes normal and oncogenic *ras* p21 molecules

(p21 purification/oncogenic mutation/valine/threonine/autophosphorylating activity)

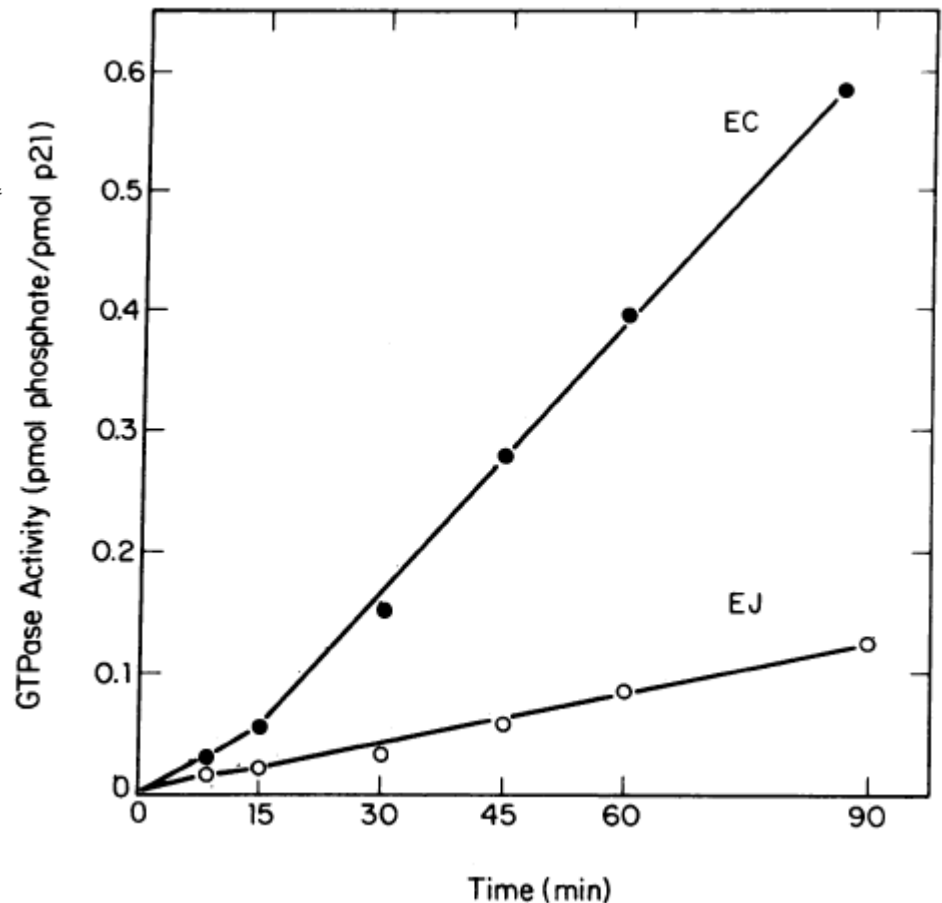
JACKSON B. GIBBS*, IRVING S. SIGAL*, MARTIN POE†, AND EDWARD M. SCOLNICK*

*Virus and Cell Biology Research, Merck Sharp & Dohme Research Laboratories, West Point, PA 19486; and †Department of Biophysics, Merck Sharp & Dohme Research Laboratories, Rahway, NJ 07065

Contributed by Edward M. Scolnick, June 11, 1984

p21 preparation	GTPase
1. EC	596 ± 170 (22)
2. EC	481 ± 42 (7)
3. EC	562 ± 190 (5)
4. EJ	129 ± 50 (13)
5. EJ	169 ± 14 (4)
6. EC/v-Ha	153 ± 110 (4)
7. EC/v-Ha	118 ± 20 (5)
8. EJ/v-Ha	18 ± 10 (4)

- Mutant RAS protein (RAS G12V) has a lower GTPase activity than wild type RAS, suggesting mutant RAS stays in an activated GTP bound form



Early low affinity inhibitors



Bioorganic & Medicinal Chemistry, Vol. 5, No. 1, pp 125–133, 1997
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0968-0896/97 \$17.00 + 0.00

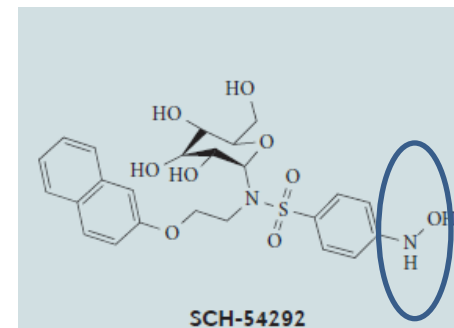
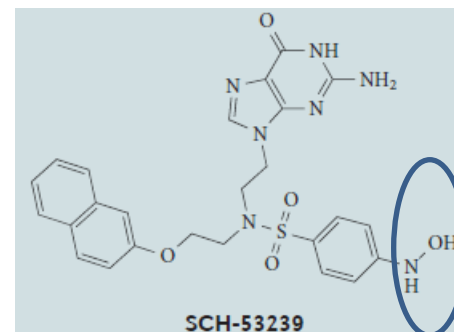
PII: S0968-0896(96)00202-7

Ras Oncoprotein Inhibitors: The Discovery of Potent, Ras Nucleotide Exchange Inhibitors and the Structural Determination of a Drug–Protein Complex

A. G. Taveras,^a S. W. Remiszewski,^a R. J. Doll^{a,*}, D. Cesarz,^a E. C. Huang,^a P. Kirschmeier,^a B. N. Pramanik,^a M. E. Snow,^a Y.-S. Wang,^a J. D. del Rosario,^a B. Vibulbhan,^a B. B. Bauer,^a J. E. Brown,^a D. Carr,^a J. Catino,^a C. A. Evans,^a V. Girijavallabhan,^a L. Heimark,^a L. James,^a S. Liberles,^a C. Nash,^a L. Perkins,^a M. M. Senior,^a A. Tsarbopoulos,^a A. K. Ganguly,^a R. Aust,^b E. Brown,^b D. Delisle,^b S. Fuhrman,^b T. Hendrickson,^b C. Kissinger,^b R. Love,^b W. Sisson^b, E. Villafranca^b and S. E. Webber^b

^aSchering-Plough Research Institute, 2015 Galloping Hill Road, Kenilworth, NJ 07033, U.S.A.

^bAgouron Pharmaceuticals, San Diego, CA, 92121, U.S.A.



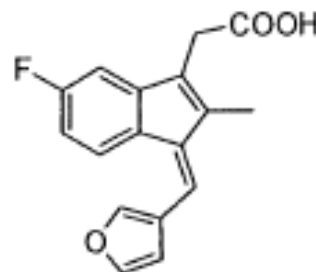
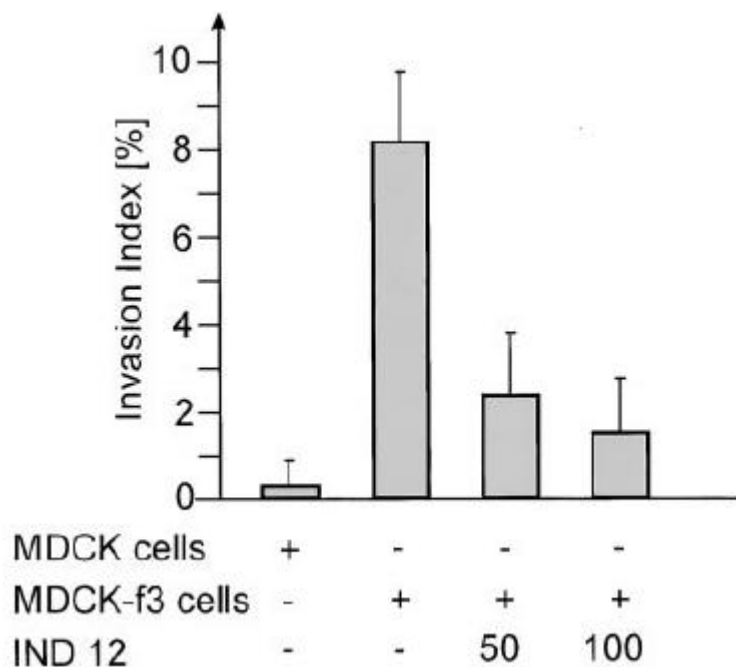
- Designed to bind nucleotide binding site but subsequently found to bind hydrophobic pocket near SII effector region
- Low affinity
- Contained hydroxylamine moiety which has toxicity and poor metabolic stability

Early low affinity inhibitors

[CANCER RESEARCH 62, 1718–1723, March 15, 2002]

The New Sulindac Derivative IND 12 Reverses Ras-induced Cell Transformation¹

Ioanna-Maria Karaguni, Peter Herter, Philip Debruyne, Slava Chtarbova, Alice Kasprzyński, Ulrike Herbrand, M-Reza Ahmadian, Karl-Heinz Glüsenkamp, Günther Winde, Marc Mareel, Tarik Möröy, and Oliver Müller²



- Non covalently bind RAS to inhibit the formation of the RAS-RAF complex
- Not very potent compounds
- Multiple other off target effects

RAS- GEF targeted inhibitors

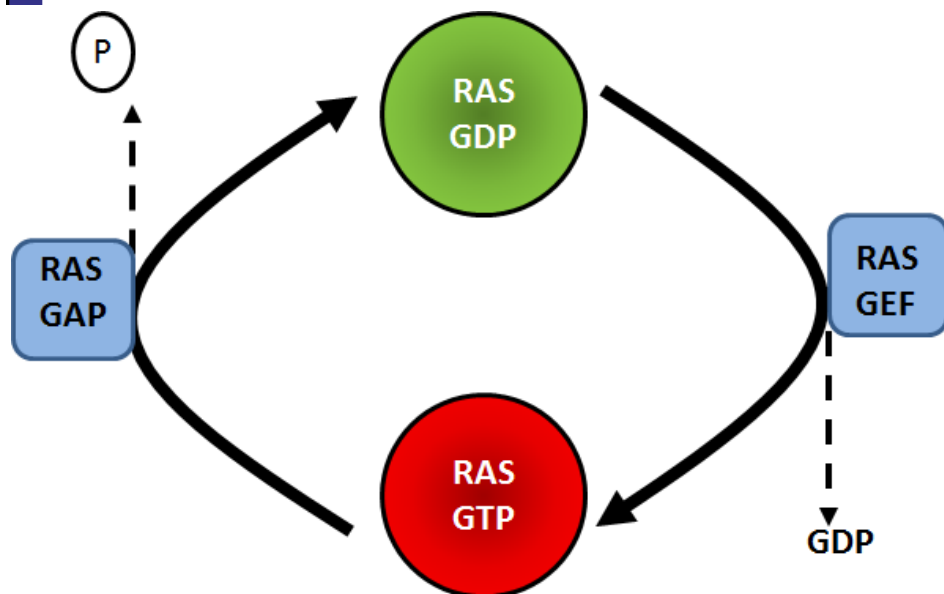
PNAS

Small-molecule ligands bind to a distinct pocket in Ras and inhibit SOS-mediated nucleotide exchange activity

Till Maurer^{a,1}, Lindsay S. Garrenton^{b,1}, Angela Oh^a, Keith Pitts^c, Daniel J. Anderson^b, Nicholas J. Skelton^d, Benjamin P. Fauber^d, Borlan Pan^d, Shiva Malek^c, David Stokoe^b, Mary J. C. Ludlam^b, Krista K. Bowman^a, Jiansheng Wu^a, Anthony M. Giannetti^a, Melissa A. Starovasnik^a, Ira Mellman^b, Peter K. Jackson^b, Joachim Rudolph^a, Weiru Wang^{b,2}, and Guowei Fang^{b,2}

^aStructural Biology, ^bResearch Oncology, ^cBiochemical and Cellular Pharmacology, ^dDiscovery Chemistry, and ^eProtein Chemistry, Genentech, Inc., One DNA Way, South San Francisco, CA 94080

Edited by* Sung-Hou Kim, University of California, Berkeley, CA, and approved February 2, 2012 (received for review October 6, 2011)



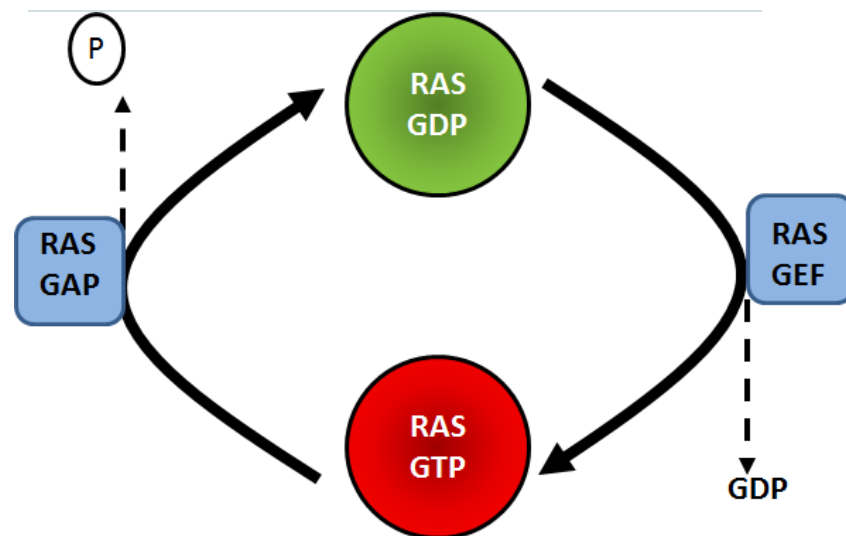
Angewandte
Communications

Fragment-Based Screening

DOI: 10.1002/anie.201201358

Discovery of Small Molecules that Bind to K-Ras and Inhibit Sos-Mediated Activation**

Qi Sun, Jason P. Burke, Jason Phan, Michael C. Burns, Edward T. Olejniczak, Alex G. Waterson, Taekyu Lee, Olivia W. Rossanese, and Stephen W. Fesik*



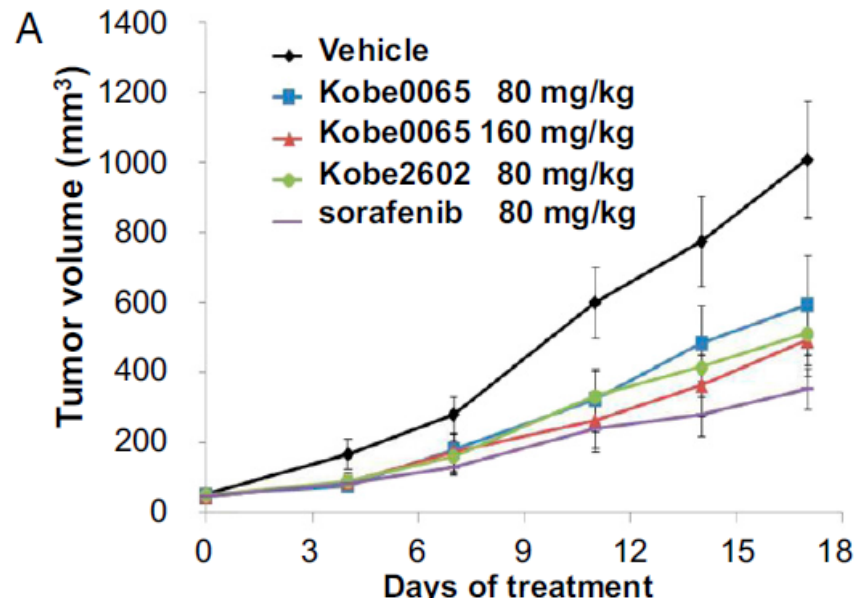
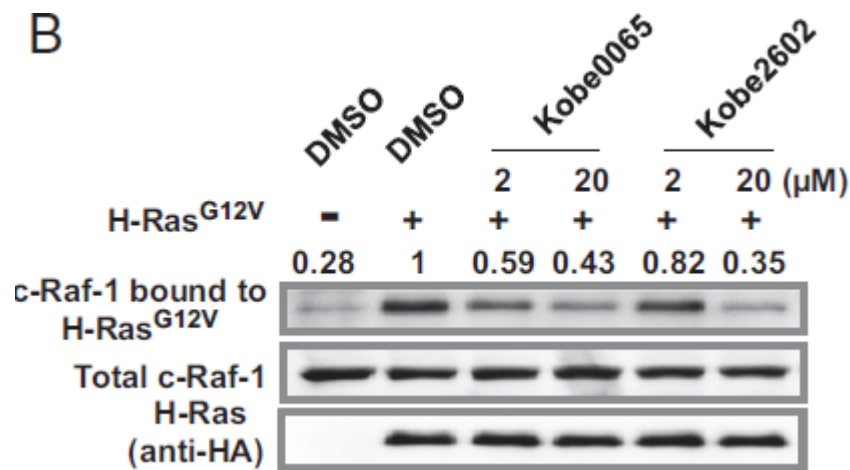
- Fragment based approaches have identified compounds such as DACI and VU0460009 which binds to KRAS and inhibited RAS-SOS1 mediated nucleotide exchange
- Only weak binding to RAS and possibility that they may not work in the setting of mutationally active RAS

RAS-RAF interaction

PNAS

In silico discovery of small-molecule Ras inhibitors that display antitumor activity by blocking the Ras-effector interaction

Fumi Shima^{a,1,2}, Yoko Yoshikawa^{a,1}, Min Ye^a, Mitsugu Araki^{a,b}, Shigeyuki Matsumoto^a, Jingling Liao^a, Lizhi Hu^a, Takeshi Sugimoto^a, Yuichi Ijiri^a, Azusa Takeda^a, Yuko Nishiyama^a, Chie Sato^a, Shin Muraoka^a, Atsuo Tamura^b, Tsutomu Osoda^c, Ken-ichiro Tsuda^d, Tomoya Miyakawa^c, Hiroaki Fukunishi^e, Jiro Shimada^e, Takashi Kumasaka^f, Masaki Yamamoto^g, and Tohru Kataoka^{a,2}



- Kobe 0065 was identified with a computer docking screen using a virtual library and was selected for its ability to inhibit HRAS-GTP binding to the RAF-RAS binding domain
- Binds to the SII region of RAS close to but in a distinct pocket from DACI and VU0460009

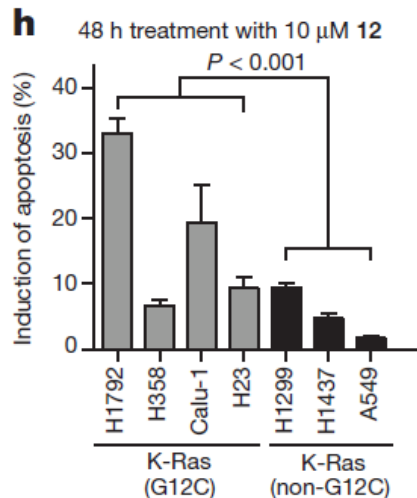
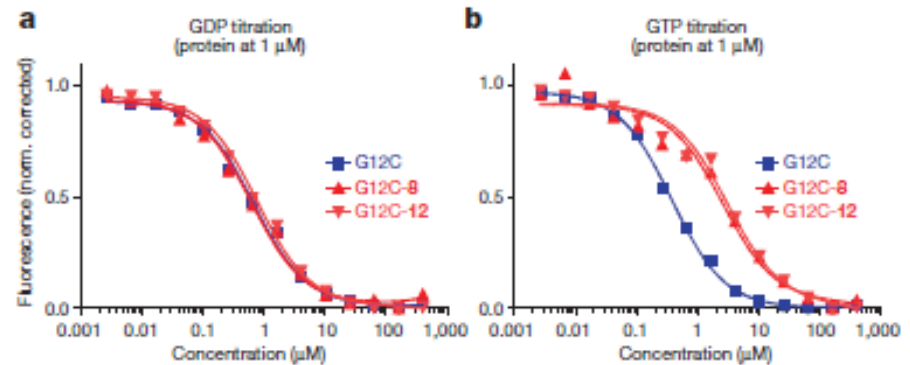
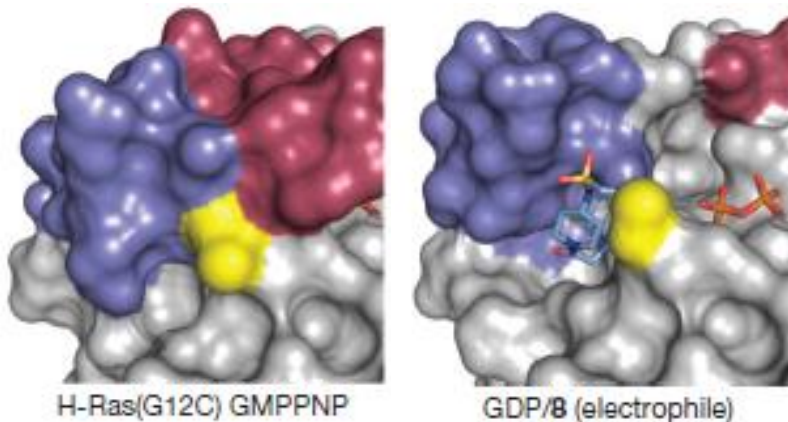
Mutant specific inhibitors

LETTER

doi:10.1038/nature12796

K-Ras(G12C) inhibitors allosterically control GTP affinity and effector interactions

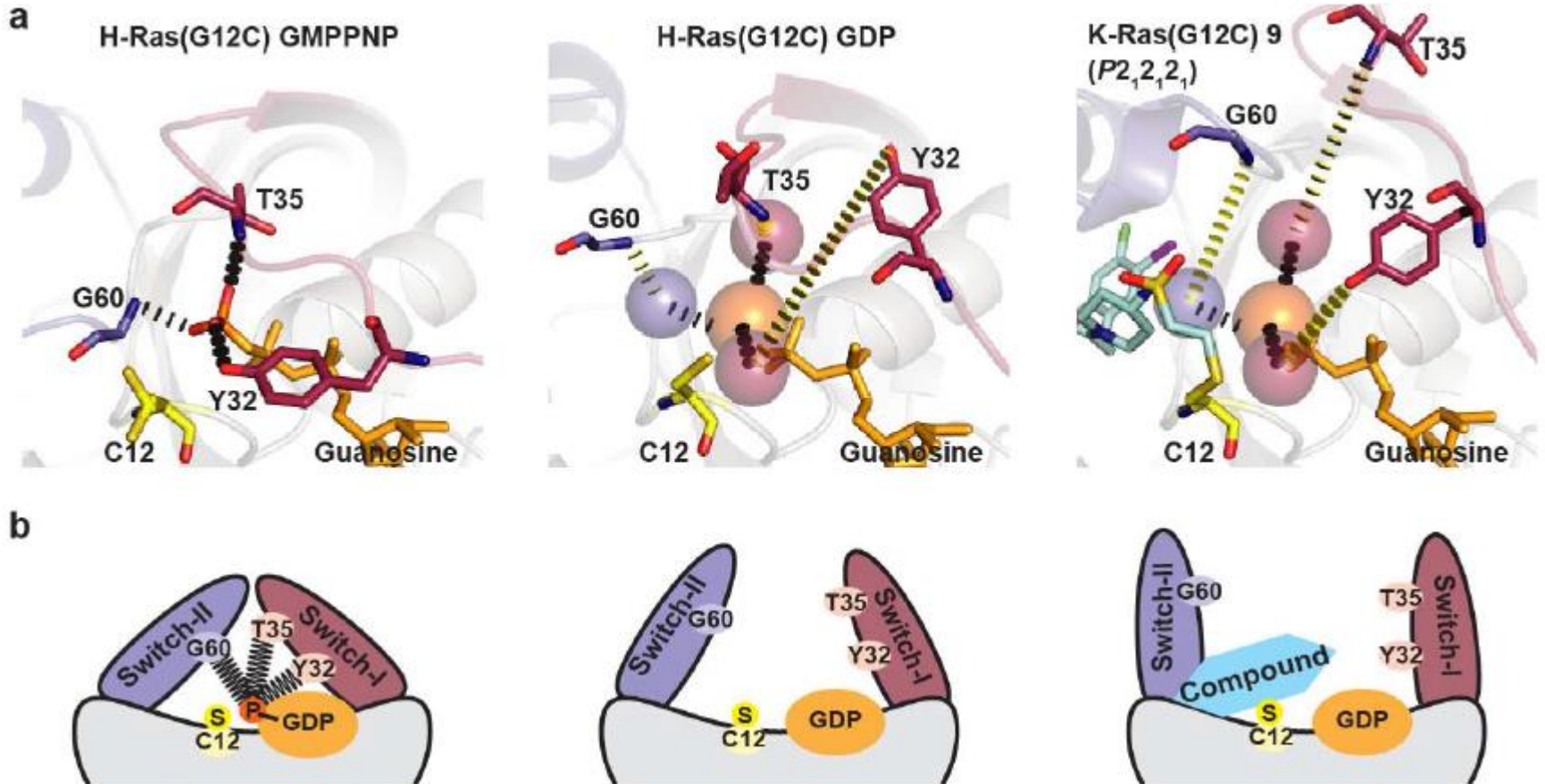
Jonathan M. Ostrem^{1*}, Ulf Peters^{1*}, Martin L. Sos¹, James A. Wells² & Kevan M. Shokat¹



- Compounds bind to pocket on the effector binding switch II region
- Allosteric binding makes mutant RAS favour GDP bound state
- Is more effective in mutations resulting in cysteine substitution rather than mutations that do not result in cysteine substitutions and wild type RAS

Ostrem JM et al Nature 2013, 503: 548-551

Mutant specific inhibitors



- γ phosphate interacts with Tyrosine 32, Threonine 30 and Glycine 60 to keep hold SII and SI in place
- Allosteric binding of compound leads to removal of γ phosphate leading to relaxation movement of SII

Mutant specific inhibitors

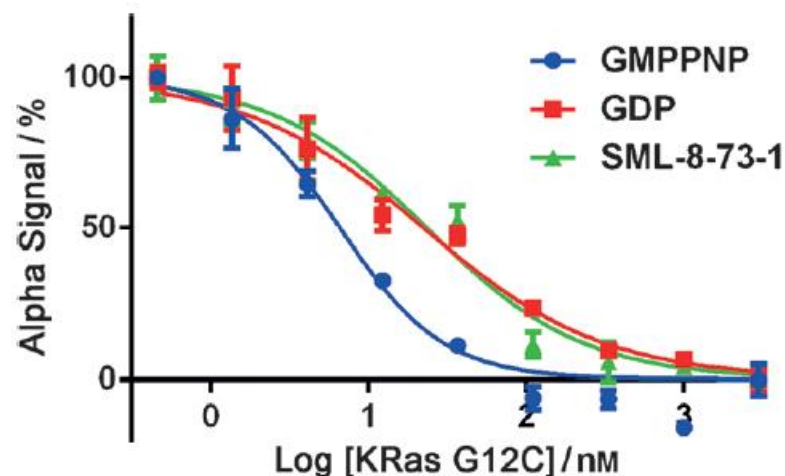
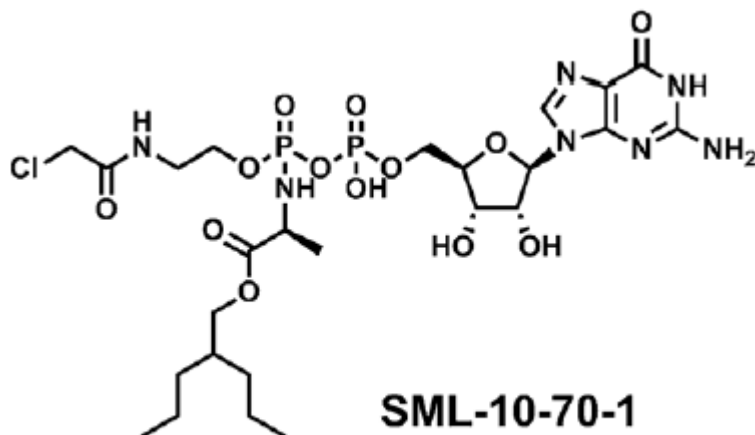


Drug Design Hot Paper

DOI: 10.1002/anie.201307387

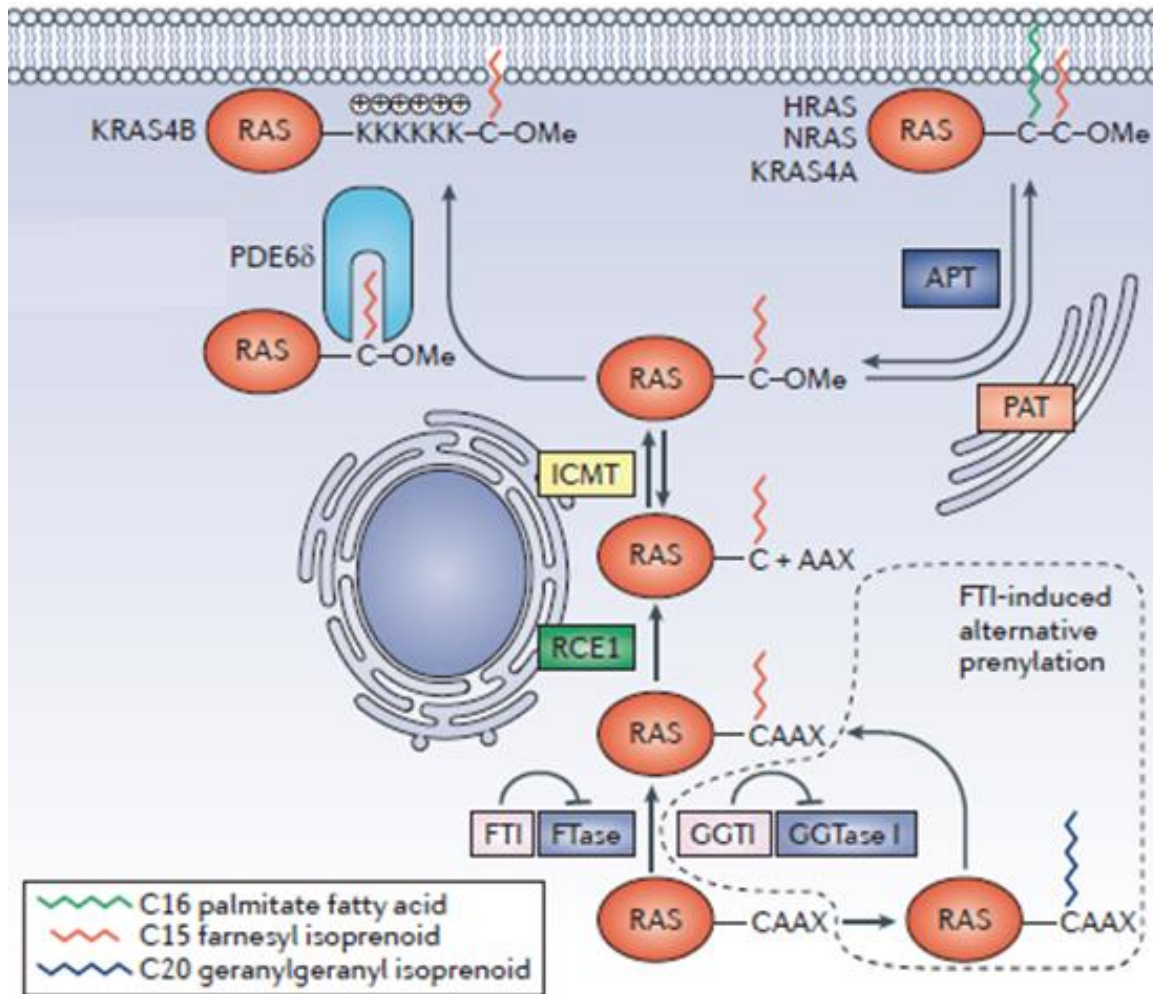
Therapeutic Targeting of Oncogenic K-Ras by a Covalent Catalytic Site Inhibitor**

Sang Min Lim, Kenneth D. Westover, Scott B. Ficarro, Rane A. Harrison, Hwan Geun Choi, Michael E. Pacold, Martin Carrasco, John Hunter, Nam Doo Kim, Ting Xie, Taebo Sim, Pasi A. Jänne, Matthew Meyerson, Jarrod A. Marto, John R. Engen, and Nathanael S. Gray*



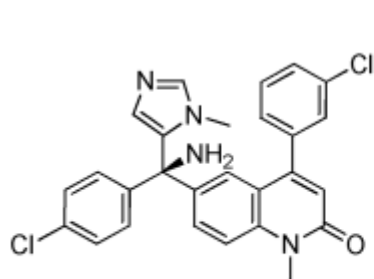
- Covalently binds to cysteine residue of G12C
- Does so in the presence of 1 mM concentration of GTP and GDP
- Not very cell permeable

Post translational modification of RAS

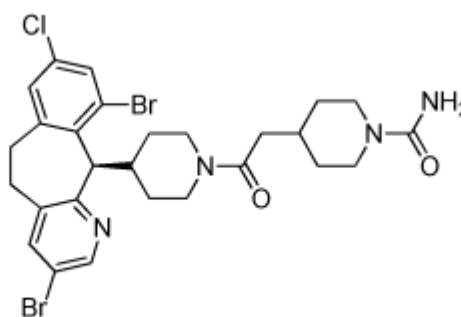


- All the 4 RAS proteins are synthesized as cytosolic inactive peptides
- HRAS and HREB is exclusively farnesylated
- RHOA, RHOC are exclusively geranylgeranylated
- RHOA, RHOC are both farnesylated and geranylgeranylated
- KRAS and NRAS are preferentially farnesylated but will be geranylgeranylated in the presence of a FT inhibitor

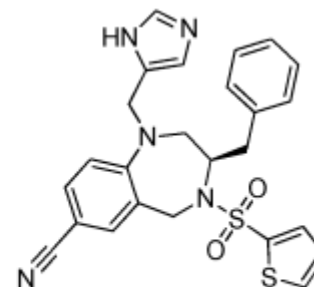
Farnesyl transferase inhibitors



24 (Tipifarnib)



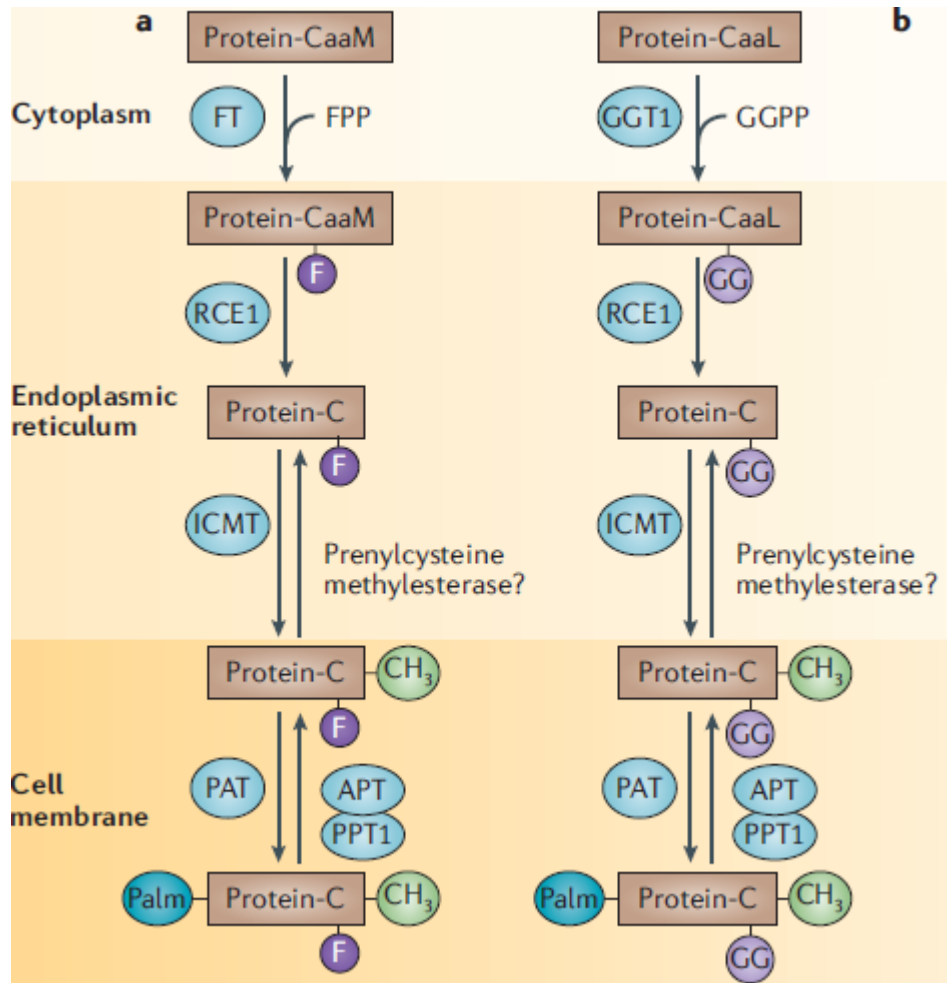
25 (Lonafarnib)



26 (BMS-214662)

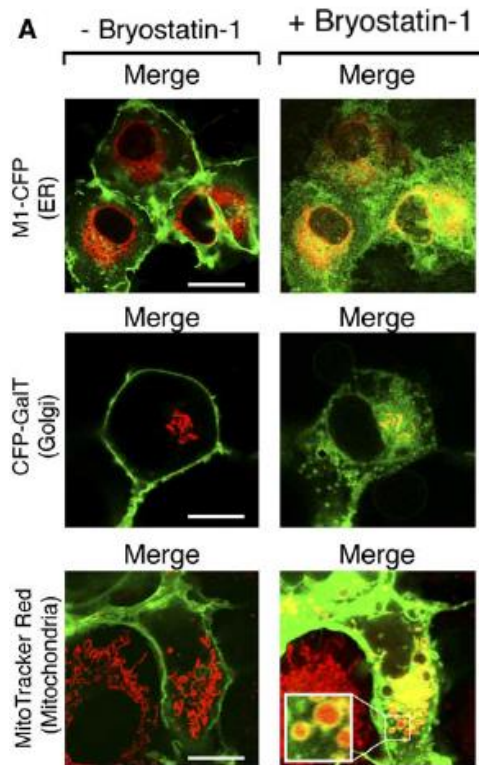
- Multiple phase I and phase II studies conducted
- Disappointing activity in solid tumours, due to the fact that most RAS mutations are KRAS or NRAS which can be prenylated by geranylgeranyl transferase upon FT inhibition
- Some activity seen in leukaemia's and haematological malignancies however these patients did not have RAS mutations, possibly due to the effects of farnesylation of other proteins
- Nausea, diarrhoea and fatigue are predominant toxicities and could be due to inhibition of farnesylation of multiple proteins.

Other targets that effect post-translational modification of RAS



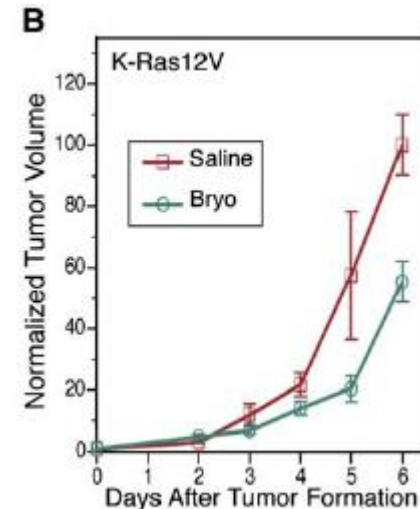
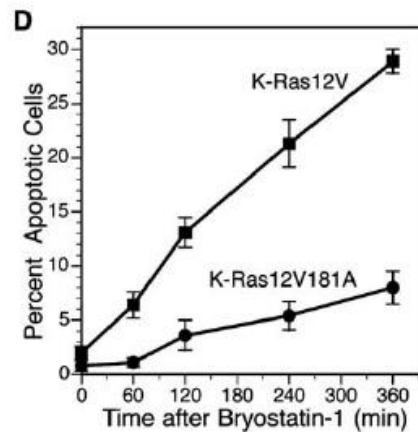
- Geranylgeranyl transferase adds a C-20 isoprenoid to selected RAS isoforms. Inhibitors include GGT1-2418
- RCE1 (RAS converting enzyme 1)
- ICMT (isoprenylcysteine-carboxymethyl transferase)
- Palmitoylation of KRAS4A, HRAS and NRAS is essential to their membrane association and function.
- One human palmitoyl acetyl transferase (DHH9-GCP16 complex) has activity against HRAS and NRAS

Other targets that effect post-translational modification of RAS: PKC α



Molecular Cell 21, 481–493, February 17, 2006 ©2006 Elsevier Inc. DOI 10.1016/j.molcel.2006.01.012

PKC Regulates a Farnesyl-Electrostatic Switch on K-Ras that Promotes its Association with Bcl-X_L on Mitochondria and Induces Apoptosis



- PKC α catalyses phosphorylation of KRAS4B at S181 within the C terminal.
- This phosphorylation causes the KRAS4 to disassociate from the plasma membrane and move to the endomembrane
- Bryostatin is a PKC agonist

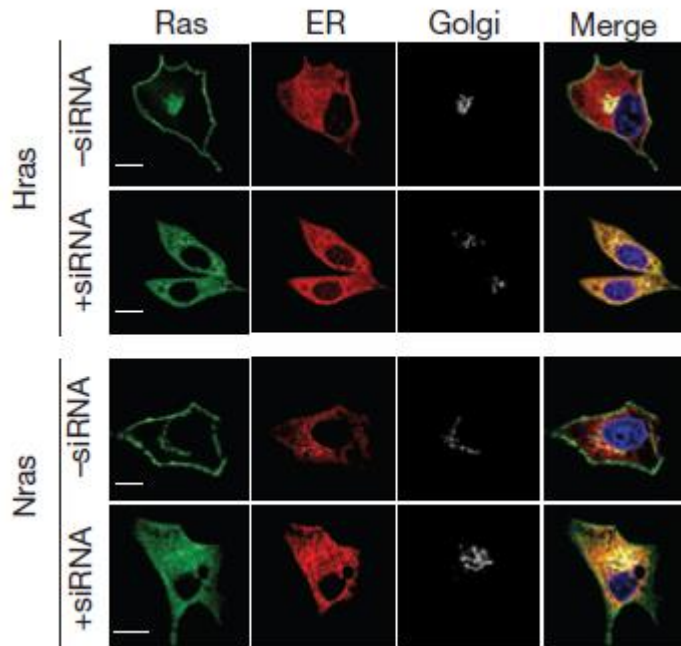
Other targets that effect post-translational modification of RAS: PDE6 δ

LETTER

doi:10.1038/nature12205

Small molecule inhibition of the KRAS–PDE δ interaction impairs oncogenic KRAS signalling

Gunther Zimmermann^{1*}, Björn Papke^{2*}, Shehab Ismail^{3*}, Nachiket Vartak², Anchal Chandra², Maike Hoffmann⁴, Stephan A. Hahn⁴, Gemma Triola¹, Alfred Wittinghofer³, Philippe I. H. Bastiaens^{2,5} & Herbert Waldmann^{1,5}

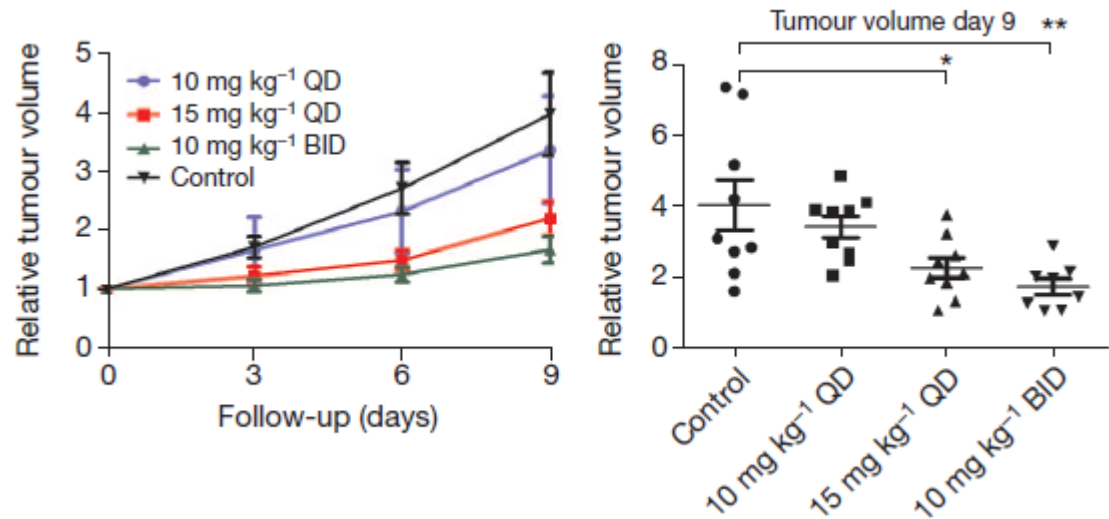


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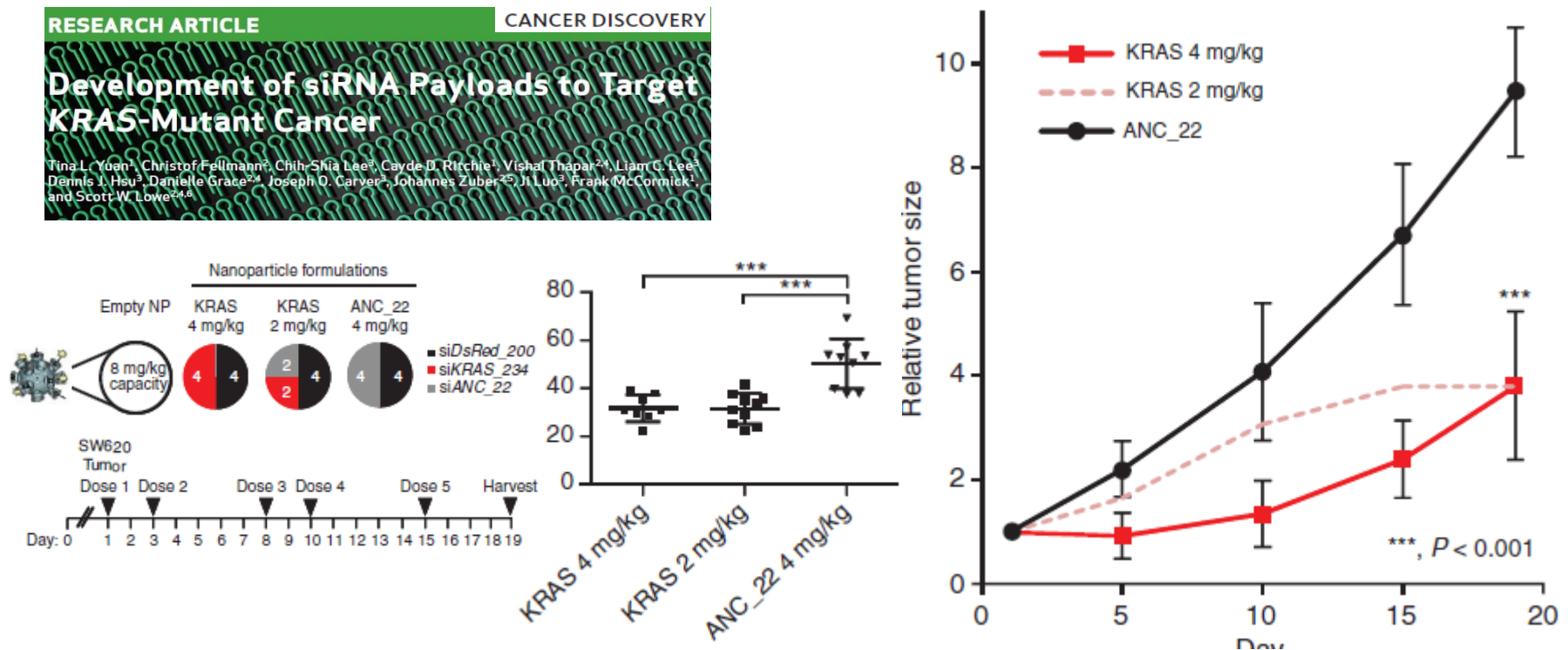
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siRNA payloads to target KRAS mutant cancers



- Preclinical efforts to try and deliver SiRNA payloads with nanoparticles are improving

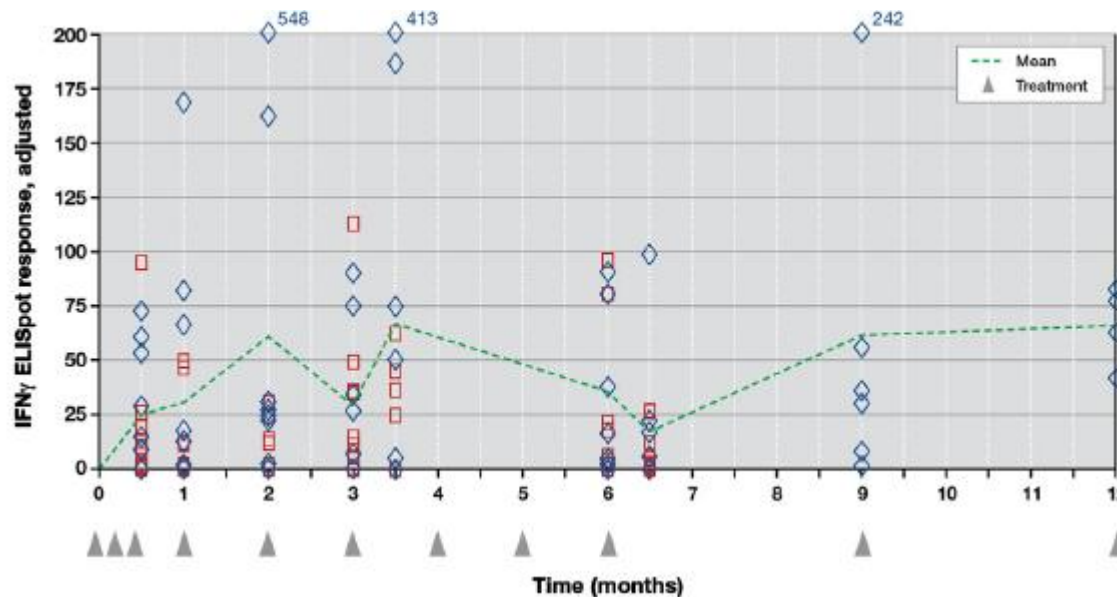
Nothing is complete without a mention of immunology – not even RAS!

Original Study



Phase II Study of the GI-4000 *KRAS* Vaccine
After Curative Therapy in Patients With Stage
I-III Lung Adenocarcinoma Harboring a *KRAS*
G12C, G12D, or G12V Mutation

Jamie E. Chaft,¹ Anya Litvak,¹ Maria E. Arcila,² Payal Patel,¹ Sandra P. D'Angelo,¹
Lee M. Krug,¹ Valerie Rusch,³ Alicia Mattson,⁴ Claire Coeshott,⁴ Bernard Park,³
David M. Apelian,⁴ Mark G. Kris,¹ Christopher G. Azzoli¹



- Vaccines usable only in a adjuvant setting
- Extracellular expression of mutant RAS not present however antigen presenting cells may display antigen
- Bystander effect on adjacent cancer cells

Conclusions

- Physiological and pathological problems to directly target RAS still limiting
- Fragment based screens have made more active compounds but not very potent but this has lead to improved understanding of druggability of RAS
- Mutant specific compounds are of interest
- Early efforts of drugging post translational modification of RAS with FT inhibitors proved disappointing and may have limited efficacy in *HRAS* mutant tumours, however other emerging areas such as PDE6 δ inhibitors are be interesting
- Combinations of downstream inhibitors have enabled some success
- Multiple synthetic lethal approaches are being pursed and have lead to interesting hypotheses that need new drugs or intelligent use of existing inhibitors to be tested in the clinic

Acknowledgements



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